

1 New keratose sponges after the end-Permian extinction provide  
2 insights into biotic recoveries

3

4 **Abstract**

5 We challenge the prevailing view that the end-Permian extinction impeded the  
6 Triassic evolution of sponges. Here we report a deep-water community dominated by  
7 abundant keratose sponges in the lowest Triassic strata from Southwest China. The  
8 sponge fossils occur as dark elliptical imprints in mudstone with distinct oscula on  
9 their tops. The structure of preserved fibers suggests closest affinity with the extant  
10 Dictyoceratida, an aspiculate demosponge. The exceptional preservation plays a  
11 crucial role in retaining their exquisite structures. Sedimentary, taphonomic, pyrite  
12 framboid, and trace elemental analyses indicate that the sponges proliferated in an  
13 oxygen-poor habitat, demonstrating the high tolerance of sponges to severe conditions.  
14 Sponge proliferation is a signal of environmental upheaval but they also stabilized the  
15 ecosystem, driving the first phase of biotic recovery after the end-Permian extinction.

16

17 **Key words:** keratose sponge, environmental stress, recovery, end-Permian extinction,  
18 Southwest China

19

20 **1 INTRODUCTION**

21

22 The end-Permian extinction severely depleted marine biodiversity, and both

23 oxygen-poor and poisonous conditions prevailed in the earliest Triassic oceans,  
24 delaying biotic recovery (Chen and Benton, 2012; Huang et al., 2023); oceanographic  
25 conditions after the end-Permian crisis were far from ideal for the colonization of the  
26 seabed. Sponges, particularly those reef-building groups, suffered dramatic losses  
27 during the crisis (Finks, 2010). Global range charts show that sponges overall  
28 experienced an evolutionary gap in the aftermath of the end-Permian crisis (Velledits,  
29 2008; Finks, 2010), although there are sporadic reports of this clade from the Lower  
30 Triassic in western US and southern China (Brayard et al., 2017; Dai et al., 2023).  
31 Previous studies have reported some keratose sponge-like textures occasionally  
32 occurring in microbialites near the Permian–Triassic (P–Tr) boundary (Friesenbichler  
33 et al., 2018; Heindel et al. 2018; Baud et al., 2021; Wu et al., 2022). Similar keratose  
34 sponge-like fibers have received significant attention in recent years (e.g., Luo and  
35 Reitner, 2014; Luo et al., 2022; Lee and Riding, 2021; 2023) and have been found in  
36 an 890-million-year-old carbonate build up (Turner, 2021). However, such  
37 interpretations remain questionable (Neuweiler et al., 2023), and the identification of  
38 aspiculate sponge in other facies is much more challenging. Thus, previous records of  
39 keratose sponge fossils require critical appraisal to substantiate their reliability, and  
40 the survival status of sponges in earliest Triassic still remains obscure.

41 Here, we report abundant, well-preserved aspiculate sponges, together with  
42 abundant brachiopods and ammonoids from the lowest Triassic (Griesbachian) in the  
43 Kejiao section, Guizhou Province, South China. These fossils unexpectedly occurred  
44 immediately after the end-Permian extinction with a mode of preservation that agrees

45 well with Lagerstätte forms. Moreover, the taphonomic window of the Kejiao section,  
46 links low oxygen conditions with exceptional preservation similar to that of many  
47 Cambrian Lagerstätten (Harper et al., 2019; Zheng et al., 2022), and is compatible  
48 with the hypoxia tolerance of sponges.

49

## 50 **2 GEOLOGICAL SETTING**

51

52 The new sponge fossils were collected from the lowest Triassic strata in the  
53 Kejiao section of Huishui County, Guizhou Province, southwest China (Figure S1a),  
54 located in the northern Nanpanjiang Basin of the southern South China block that was  
55 situated at low latitudes in the eastern Paleo-Tethys region during the earliest Triassic  
56 (Enos et al., 2006; He et al., 2022; Figure 1). The upper Permian Talung Formation is  
57 composed of siliceous mudstone and carbonaceous shales, interbedded with tuff and  
58 calcareous mudstone (Figure 2), whereas the Lower Triassic Luolou Formation is  
59 dominated by yellowish mudstone and calcareous mudstone, with several layers of  
60 grayish green marlstone at the base. The Talung Formation yields abundant  
61 cephalopods, bivalves, brachiopods, radiolarians, foraminiferans, and sponge spicules.  
62 The cephalopod fauna including *Pseudotirolites* and *Pleuronodoceras*, indicates a  
63 latest Changhsingian age (Jiang et al., 2018). Fossil assemblages, along with the  
64 presence of laminations and distal Bouma turbidite sequences, indicate a deep-water  
65 environment (Wang et al., 2023). The Luolou Formation bed 74 to bed 83 is  
66 characterized by the presence of the ammonoids *Ophiceras* spp. and bivalve *Claraia*

67 *wangi*, assignable to the *Claraia wangi*-*C. gresbachi* assemblage and *Ophiceras*  
68 assemblage, respectively, equivalent to the conodont *Isarcicella staeschei* Zone to the  
69 *I. isarcica* Zone in the Meishan section, South China (Chen et al., 2015), which is the  
70 Global Stratotype Section and Point (GSSP) for the P–Tr boundary (PTB) (Yin et al.,  
71 2001; Shen et al., 2022), indicating an early Griesbachian age. The PTB therefore is  
72 placed at the base of Bed 74 in the Kejiao section (Wang et al., 2023).

73 The new sponge fossils, in association with an unidentifiable species of the  
74 ammonoid *Ophiceras*, bivalve *Claraia wangi* and brachiopod *Meishanorhynchia*  
75 (Chen et al., 2002; Guo et al., 2022), occur in strata 4.5–6 m above the PTB, beds  
76 82–84 in Luolou Formation (Figure 2, blue bar), and are therefore early Griesbachian  
77 in age.

78

## 79 MATERIAL AND METHODS

80

81 A total of 100 slabs were collected from 10 layers within the 1.5-m-thick  
82 sponge-bearing strata in Kejiao, and >600 well-preserved sponge individuals were  
83 measured. Taxonomic identification follows Antcliffe et al. (2014), with comparison  
84 with morphological features of modern sponge specimens illustrated in ‘The Sponge  
85 Guide’ (Zea et al., 2014) and dried specimens of modern sponges from the authors’  
86 personal collections.

87 Microfacies analysis and the paleoenvironmental interpretation were prepared  
88 based on observations of conventional petrologic thin-sections. To assess redox

89 conditions, 22 Kejiao samples were prepared for whole-rock trace element analysis  
90 (Figure 2). The trace element analysis was performed at Wuhan Sample Solution  
91 Analytical Technology Co., Ltd. using Agilent 7700e ICP-MS. V/Cr and V/(V + Ni)  
92 ratios are ideal indices of redox conditions in ancient mudstones (Hatch and Leventhal,  
93 1992; Jones and Manning, 1994). The V/Cr ratio varied through oxic ( $< 2$ ), dysoxic  
94 ( $2\sim 4.25$ ) and anoxic ( $> 4.25$ ) conditions (Jones and Manning, 1994). The V/(V+Ni)  
95 ratio varied through dysoxic (0.46~0.60), anoxic (0.54~0.82) and euxinic ( $> 0.84$ )  
96 conditions (Hatch and Leventhal, 1992).

97 A total of 60 rock samples from 22 horizons were collected for pyrite framboid  
98 analysis to help unravel the P–Tr redox history of the Kejiao succession. Pyrite  
99 framboids were observed and measured by scanning electron microscopy (SEM,  
100 Hitachi SU8000) at the State Key Laboratory of Biogeology and Environmental  
101 Geology (BGEG) in the China University of Geosciences (Wuhan). Criteria for the  
102 relationships between framboid size distributions and morphology, and redox states  
103 follow Bond and Wignall (2010). Dispersed framboids instead of clustered framboids  
104 are chosen to minimize the influence of diagenesis (Huang et al., 2017, 2019).

105 Well-preserved sponge samples were selected for Raman analysis using the  
106 WiTec alpha300 R confocal Raman spectrometer at the University of Göttingen in  
107 Germany. The Raman spectra were obtained under a 532 nm wavelength laser at a  
108 power of 4.974 mW in a range of  $75\text{--}4000\text{ cm}^{-1}$  at a  $1\text{ cm}^{-1}$  resolution (Jing et al.,  
109 2022).

110

## 111 **4 RESULTS**

112

### 113 **4.1 Description of sponge fossils**

114 Abundant sponge individuals are densely packed on the slab surfaces, showing a  
115 lack of preferred size or orientation (Figure 3a; Figure S2). Fragmentation ratio is low,  
116 only in 8.5% of 600 individuals. The new sponge fossils have a high abundance,  
117 blanketing the entire mudstone layer, sometimes covering up to >75 % of the surface  
118 (Figure 3a) and are dominated by one new taxon together with minor elements,  
119 including one brachiopod species and ammonoids. Sponge specimens are flattened,  
120 presented as circular-to-elliptical, thin carbonaceous films on mudstone surfaces, but  
121 preserving three-dimensional features (Figure 3f). A fragmented fossil specimen  
122 displays the thickness of the lateral wall, with the mesh penetrating through the lateral  
123 walls (Figure 3f). The fossils are composed of reticulate, organic networks (spongin  
124 skeleton), lacking identifiable spicules. The residual sponge material, overall, is  
125 sack-shaped with a circular opening on the top serving as an osculum (Figure 3; red  
126 arrows), which is typical of sponges. Complete individuals are 2–16 mm (average 5.6  
127 mm) wide and 3–30 mm (average 8.7 mm) high, with height/width ratios of 0.9–3.  
128 Oscula are 1–5 mm in diameters, occupying 20–70 % of sponge width. Organic  
129 frameworks are 0.5–1 mm wide.

130 The budding process of Kejiao sponges was preserved, manifest as an outgrowth  
131 from the parent body (Figure 3k-m). At the junction of the bud and the mother sponge,  
132 the base of the bud is slightly narrowed, while the bases of other ordinary individuals

133 are round and smooth, indicating that the narrow base is characteristic of the  
134 connection between the bud and the mother, rather than the two individuals simply  
135 overlapping. Therefore, this phenomenon may be due to the fact that the budding  
136 reproductive process of sponges has just been preserved. Budding is a common mode  
137 of asexual reproduction in sponges, representing an important reproductive strategy  
138 for some species, and maximizing the dispersal of these animals (Gaino et al., 2006).  
139 The Kejiao fossils represent the first record of budding reproduction in ancient  
140 keratose sponges.

141 The preserved fibers show thick primary and thinner secondary structures,  
142 suggesting that they represent a new keratose aspiculate demosponge which exhibits a  
143 best fit with the extant Dictyoceratida (see details in Supplementary materials, Figure  
144 4). A new genus and new species *Palaeosmenospongia kejiaoensis* n. gen, n. sp. is  
145 proposed. However, though the spongin network is well preserved, other soft tissues  
146 (e.g. epidermal cells) are not, which limit any valid data about the soft tissue function.

147 Raman spectroscopy is employed to understand the taphonomic pathway of the  
148 sponge material. The Raman spectra of the dark fossil remains possess two distinct  
149 bands at wave-length numbers  $1350\text{ cm}^{-1}$  and  $1580\text{ cm}^{-1}$  (Figure 5c-f), typical of  
150 organic carbon (Ferrari and Robertson, 2000; Jing et al., 2022). D (disordered carbon)  
151 and G (graphite) bands are very sharp and indicate strong maturation of the organic  
152 carbon, while the Raman spectra of the surrounding mudstone lack such bands. No  
153 silica signal was observed.

154

155 **4.2 Systematic paleontology**

156

157 Phylum Porifera Grant, 1836

158 Class Demospongiae Sollas, 1875

159 Order Dictyoceratida Minchin, 1900

160 Family Thorectidae Bergquist, 1978

161 Genus *Palaeosmenospongia* n. gen. Wu & Reitner

162

163 **Type species.** *Palaeosmenospongia kejiaoensis* n. gen., n. sp. Wu & Reitner

164 **Etymology.** Combined from “*palaeo*” = old (Latin) and “*Smenospongia*”, a  
165 comparable extant genus of the Family Thorectidae.

166 **Diagnosis.** Vase to cup shaped centimeter-sized sponges, with a large central osculum  
167 and a network of thick and thin fibers commonly crowded and interconnected; it is  
168 possible to distinguish between thick primary and thinner secondary fibers. The  
169 organic skeleton is thin and only one or two layers are distinguishable. The fibers do  
170 not show any agglutinated or generally allochthonous material. This sponge taxon  
171 commonly shows asexual buds.

172

173 *Palaeosmenospongia kejiaoensis* n. gen, n. sp. Wu & Reitner

174 (Figure 3a-m and Figure 4a-c)

175

176 **Etymology.** The new species is named after the Kejiao section.

177 **Type specimens.** One complete specimen (BGEG KJ10301) is selected as holotype  
178 (Figure 4a). Other three specimens are selected as Paratype I (BGEG KJ10302; Figure  
179 4b), Paratype II (BGEG KJ10303; Figure 4c), and Paratype III (BGEG KJ10314;  
180 Figure 3i).

181

182 **Horizon and locality.** The greenish to brownish calcareous mudstone of the lowest  
183 Luolou Formation (lower Griesbachian); the Kejiao section of Duanshan Town in



184 Huishui County, Guizhou Province, southwest China.

185

186 **Remarks.** This new type of fossil aspiculate keratose demosponge is currently unique  
187 and has no obvious relatives. The new species is differentiated from the well-known  
188 keratose demosponges of the family Vauxiidae Walcott, 1920 as the latter are tubular  
189 and often branched (Rigby, 1980; Yang et al., 2017; Luo et al., 2020). The fiber  
190 skeleton of that taxon consists of a very regular reticulate network and shows,  
191 normally, a polygonal space between the fibers which is not observed in the new  
192 taxon. The fiber skeleton of the Vauxiidae is normally formed by two layers, a dermal  
193 and a choanosomal one, which was not observed in the new taxon. The Vauxiidae are  
194 most likely related to the Verongida or Verongimorpha *sensu* Erpenbeck et al. (2012)  
195 the other large phylogenetic group of aspiculate demosponges. The chemical  
196 composition of the fiber skeleton of the Vauxiidae has a chitin composition (Ehrlich et  
197 al., 2013) which is known in the Verongimorpha and not from the newly erected  
198 phylogenetic group “Keratosa” which includes the Dictyoceratida and Dendroceratida  
199 (Erpenbeck et al., 2012). The fiber skeleton of this group is composed only of spongin,  
200 a complex compound of scleroproteins. The key chemical signals that can be  
201 identified as specific organic components have been lost, but the morphological  
202 characteristics are closer to those of modern spongin textures. The new sponge  
203 exhibits some morphological similarities with the Cambrian taxon *Crumillospongia*.  
204 However, this taxon has a spicular skeleton with affinities to *Hazellia* (Rigby and  
205 Collins, 2004), which is probably related to the Halichondrida; a taxon that is  
206 explicitly not a keratose demosponge. Considering the similarity of the fiber structure,  
207 especially the arrangement of thick primary and thinner secondary fibers (Figure 4),  
208 our new taxon is most probably related to the modern genus *Smenospongia*  
209 Wiedenmayer, 1977 (Figure 4d-g).

210

## 211 **4.3 Redox interpretation**

### 212 *4.3.1 Elemental geochemistry*

213 Both trace elemental proxies: V/Cr and V/ (V + Ni) ratios are employed to  
214 unravel redox changes across the end-Permian mass extinction in Kejiao. Most V/Cr  
215 ratios from the uppermost Permian are rather low, ranging from 1.23 to 1.70. The low  
216 values continue up into the lowest Triassic and there is a transition between bed 78  
217 and bed 79. The V/Cr ratios from bed 78 to bed 85 range from 2.13 to 4.08, within the  
218 dysoxic window (Figure 2).

219 The V/ (V + Ni) ratios derived from the Permian samples range from 0.075 to  
220 0.54 (averaging 0.36), and the curve rose gradually following the PTB. Except for two  
221 points that are lower, the V/ (V + Ni) ratios of the mudstone are greater than 0.84  
222 (Figure 2), indicating euxinia (Hatch and Leventhal, 1992; Shi et al., 2016), which is  
223 reinforced by beds that consist of dark, organic-rich mudstone with fine, horizontal  
224 laminae (Fig. S4E-F). Thus, the sponge-bearing layers were most probably deposited  
225 in a basin with oxygen-poor condition.

226

#### 227 *4.3.2 Pyrite Framboid Analysis and Redox Interpretation*

228 A total of 17 out of 22 (77%) samples yield pyrite framboids (Figure 2). Overall,  
229 the Lower Triassic yellowish mudstones tend to yield more abundant and smaller  
230 framboids than the uppermost Permian dark-colored siliceous mudstones. No  
231 framboids were found in the lowest Triassic greenish mudstone. Framboids derived  
232 from the six uppermost Permian samples have mean diameters (MD) of 7.15–8.96  $\mu\text{m}$   
233 with standard deviations (SD) of 2.28–3.65  $\mu\text{m}$ , indicative of dysoxic conditions  
234 (Bond and Wignall, 2010; Figure S3). Eleven of the 16 Lower Triassic samples fall

235 within the anoxic field with MDs of 3.20–5.84  $\mu\text{m}$  and SDs of 0.96–2.24  $\mu\text{m}$ .  
236 Framboids from sponge-bearing horizons (Beds 82–84) all fall into the anoxic field  
237 (Figure 2; Figure S3). No framboids were found in Beds 73–78, which straddle the  
238 P–Tr boundary. These beds, containing ammonoids and bivalves, may represent a  
239 more oxic environment (Figure 2). The coeval oxygen-rich layers between two stages  
240 of anoxia also occur in the PTB beds at the GSSP in Meishan, South China and the  
241 Guryul Ravine section, Kashmir, India (Chen et al., 2015; Huang et al., 2017, 2018),  
242 suggesting that the redox history during the P–Tr transition was consistent with that  
243 elsewhere in the Paleo-Tethys region.

244

## 245 **5 DISCUSSION**

### 246 **5.1 Taphonomy**

247 The spongin skeletons of these soft-bodied sponges are well preserved, without  
248 any evidence of preferred orientation, deformation or lateral compression, suggesting  
249 *in situ* burial. Raman measurements show a high content of organic matter in the dark  
250 sponge remains (Figure 5), clearly distinguished from the surrounding matrix,  
251 indicating the high preservation potential for organic skeletons. Considering the  
252 morphological attributes, the preserved material is most likely to be the former  
253 spongin skeleton of an aspiculate demosponge.

254 Three factors are crucial for the exceptional preservation of these sponges. First  
255 is the preservation potential of the sponge skeleton. Unlike other soft tissues like  
256 epidermal cells, the spongin skeleton of keratose sponges is a tough material that is

257 nevertheless soft, elastic, absorbent, and insoluble in water and acids, and shows  
258 strong resistance to enzymatic degradation (Jesionowski et al., 2018). Second, high  
259 V/Cr and V/(V + Ni) ratios from the sponge-bearing strata indicate dysoxic to anoxic  
260 conditions, which are considered as a prerequisite for the exceptional preservation of  
261 other Lagerstätten (e.g., Gaines et al., 2005, 2012). Third, in longitudinal section, the  
262 fossil-bearing mudstones have elongated silt-sized lenses embedded within a clay-rich  
263 dark matrix (Figure 6e-f), completely different from the underlying layers (Figure  
264 6a-d). Flume experiments showed that such lenticular fabrics in shales originated  
265 from transport of surficial water-rich muds by currents (Schieber et al., 2010). Such  
266 lenticular fabrics were also found in other Lagerstätten (e.g., Gabbott et al., 2008,  
267 2010). The slow-moving imbricated mud chips thus created a confined space,  
268 conducive to fossil preservation. Besides, this proves that the surrounding water was  
269 not stagnant.

270

## 271 **5.2 Sponges after mass extinction event**

272 The proliferation of the Kejiao sponges may have acted as a direct reflection of  
273 environmental deterioration after the end-Permian extinction. The sponges, together  
274 with tiny, thin-shelled brachiopods (Figure 3n-o) and few ammonoids, formed a  
275 typical sponge-dominated ecosystem (Figure 7) flourishing as opportunists in stressful  
276 environments. Keratose sponge fabrics found in PTB microbialites (Friesenbichler et  
277 al., 2018; Heindel et al., 2018; Baud et al., 2021; Chen et al., 2022; Wu et al., 2022a, b)  
278 are under discussion (Neuweiler et al. 2023). Luo et al. (2022), however, have

279 introduced a taphonomic concept which explains the preservation of complex spongin  
280 skeletons and related this to the assumptions made in Neuweiler et al. (2023). The  
281 Kejjiao fauna provides first hard evidence that sponges proliferated after end-Permian  
282 mass extinction, confirming that survival ability of Keratosa, at least in deep-water  
283 settings. This study bridges the gap in our knowledge of sponges within the earliest  
284 Triassic and opens a limited window for incorporating sponges into the narrative  
285 surrounding post-extinction ecosystems. In addition, the proliferation of sponges after  
286 disaster events is a common phenomenon: e.g., end-Ordovician, Late Devonian,  
287 end-Triassic extinction events and modern El Niño events (Vishnevskaya et al., 2002;  
288 Delecat & Reitner, 2005; Delecat et al. 2011; Kelmo et al., 2013; Ritterbush et al.,  
289 2015; Li et al., 2015; Botting et al., 2017), suggesting that some sponges  
290 demonstrated remarkable resilience in adapting to and surviving in these extreme  
291 environmental conditions.

292       The P–Tr great dying was associated with several extreme environmental and  
293 climate upheavals, such as expanded marine anoxia (Zhang et al., 2018). In Kejjiao,  
294 V/Cr ratio and taphonomic evidence indicate that the preservation facilitated by low  
295 levels of oxygen Lagerstätte-type sponges, suggested that they inhabited a relatively  
296 oxygen-poor environment. In fact, some modern demosponges can survive under  
297 low-oxygen conditions (Hoffmann et al. 2005; Mills et al., 2014) and, the earliest  
298 crown-group sponges had already evolved a great tolerance of oxygen-deficient  
299 habitats (Mills et al., 2014). The tolerance of low-oxygen conditions is also a property  
300 of the Cambrian Burgess-Shale-type sponge group (Gaines et al., 2005; Li et al.,

301 2021). Sponges have a proven tolerance of temperature and pH change (Duckworth et  
302 al., 2012; Kelmo et al., 2013), which would help survival of disruptive environmental  
303 change following the extinction event. Furthermore, sponges possess efficient  
304 filtration systems that ensure their survival in nutrient-depleted environments (Leys et  
305 al., 2011). Abundant keratose sponges occupied the post-extinction oxygen-depleted  
306 habitats in Kejiao and proliferated when most other marine animals suffered massive  
307 casualties. One or more of the above abilities were conducive to their survival in  
308 stressful environments and proliferate after catastrophic events.

309

### 310 **5.3 The Ecological Function and Significance of Kejiao Sponge**

311 The phylogenomic dataset available supports sponges at the base of the  
312 metazoan tree of life (Simion et al., 2017). The Ediacaran-Cambrian rise of sponges is  
313 believed to have played a role in the initiation of seawater oxygenation by  
314 redistributing organic carbon while oxygenating ambient environments through their  
315 ability to filter suspended organic matter from seawater, aiding the diversification of  
316 modern-type ecosystems (Chang et al., 2019; Zheng et al., 2022). Modern  
317 experiments show that sponges have the potential to support the whole ecosystem by  
318 transforming dissolved organic matter into particulate detritus that are beneficial for  
319 higher trophic levels (de Goeij et al., 2013), and this ecological role extends even to  
320 deep-sea environments (Bart et al., 2021). This ability positioned sponges as part of a  
321 highly efficient recycling pathway, allowing barren marine ecosystem develop high  
322 diversity (de Goeij et al., 2013). Additionally, modern studies demonstrate that sponge

323 caves could serve as refuges and ‘seed banks’ for nearby dwindling corals (Slattery et  
324 al., 2013), confirming the potential contribution of sponges in biotic recovery.  
325 Furthermore, the physical structure of sponge colonies could provide shelter and  
326 substrate for a diverse array of organisms, creating microhabitats that support the  
327 settlement and growth of numerous species (Chin et al., 2020).

328 In recent years, there has been a notable resurgence of scientific interest in  
329 sponges, driven by a growing recognition of their significant contributions to the  
330 biodiversity and functioning of marine ecosystems (Bell et al., 2018; Lesser and  
331 Slattery, 2020). Extensive research has yielded fresh insights into the pivotal role  
332 played by deep-water sponges (Rooks et al., 2020; Pierrejean et al., 2020). Modern  
333 investigations have found that deep-water sponge grounds could serve as reservoirs of  
334 biodiversity, creating habitats and refuges for other organisms (Hogg et al., 2010).  
335 And, keratose sponges have become the dominant species in some modern deep-sea  
336 habitats (Enrichetti et al., 2019), validating the adaptability and ecological potential of  
337 this group of sponges.

338 To sum up, there are some key examples that suggest that sponges acted as a  
339 prelude to highly diverse communities in early ecosystem evolution and following  
340 ecological crises. However, further research is needed to fully understand the extent  
341 of their role in these processes. The mechanism of how ecosystems began to recover  
342 during the Early Triassic is currently not well understood and the long-neglected  
343 sponge communities might also have participated in biogeochemical cycling in the  
344 earliest Triassic. The appearance of sponges in different facies after disaster events

345 implies that low-trophic-level ecosystems could have persisted in some environments,  
346 maintaining the original dynamic of biotic recovery, ready to drive the ecosystem to a  
347 higher level. Kejiao sponge assemblages represent a symbol of the collapse of marine  
348 ecosystem, but also have the potential to serve as a hotbed of marine life and the  
349 engine of the reboot of the marine ecosystem. Moreover, whether the proliferation of  
350 sponges is a local event or reached a global scale requires further investigation, which  
351 will help us better understand the ecological dynamics of the Early Triassic

352

## 353 **6 CONCLUSIONS**

354

355 Abundant keratose sponges occur in the lowest Triassic mudstone in a  
356 deep-water basin facies in Southwest China. The Kejiao sponge represents a  
357 remarkable taphonomic phenomenon with important implications for ecosystem  
358 evolution after major biotic crises. The new sponge is closest to the extant  
359 Dictyoceratida, and also exhibits a budding mode of asexual reproduction. The  
360 well-preserved fiber skeleton shows a centimeter-sized vase-shaped habit commonly  
361 with a central osculum. Sedimentary, taphonomic and trace elemental analyses show a  
362 low-oxygenated fossil Lagerstätte mode of preservation, and indicate the sponges  
363 proliferated in a deep-water, dysoxic to anoxic habitat, highlighting the high tolerance  
364 of these sponges. The first batch of sponges after disaster events may have contributed  
365 to the return of other sessile suspension feeders through their specialized ecological  
366 functions, and acted as ecosystem engineers driving biotic recovery after the



367 end-Permian extinction. It appears that further evaluation is warranted regarding the  
368 implications of the great dying for sponge taxa, as well as their subsequent Triassic  
369 evolution and ecological expansion.

370

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380

## 381 **REFERENCES CITED**

382 Antcliffe, J. B., Callow, R. H., & Brasier, M. D. (2014). Giving the early fossil record  
383 of sponges a squeeze. *Biological Reviews*, 89(4), 972-1004.  
384 <https://doi.org/10.1111/brv.12090>

385 Bart, M. C., Hudspith, M., Rapp, H. T., Verdonshot, P. F., & De Goeij, J. M. (2021).  
386 A deep-sea sponge loop? Sponges transfer dissolved and particulate organic carbon  
387 and nitrogen to associated fauna. *Frontiers in Marine Science*, 8, 604879.  
388 <https://doi.org/10.3389/fmars.2021.604879>.

389 Baud, A., Richoz, S., Brandner, R., Krystyn, L., Heindel, K., Mohtat, T.,  
390 Mohtat-Aghai, P., & Horacek, M. (2021). Sponge takeover from end-Permian  
391 mass extinction to Early Induan time: Records in Central Iran microbial buildups.  
392 *Frontiers in Earth Science*, 9, 586210. <https://doi.org/10.3389/feart.2021.586210>.

393 Bell, J. J., Rovellini, A., Davy, S. K., Taylor, M. W., Fulton, E. A., Dunn, M. R.,  
394 Bennett, H. M., Kan & Webster, N. S. (2018). Climate change alterations to  
395 ecosystem dominance: how might sponge-dominated reefs function? *Ecology*,  
396 99(9), 1920-1931. <https://doi.org/10.1002/ecy.2446>

397 Bergquist, P. (1978). *Sponges*. University of California Press, Los Angeles.

398 Bond, D. P. G., & Wignall, P. B. (2010). Pyrite framboid study of marine  
399 Permian–Triassic boundary sections: a complex anoxic event and its relationship to  
400 contemporaneous mass extinction. *Geological Society of America Bulletin*,  
401 122(7-8), 1265-1279. <https://doi.org/10.1130/B30042.1>

402 Botting, J. P., Muir, L. A., Zhang, Y., Ma, X., Ma, J., Wang, L., Zhang, J., Song, Y., &  
403 Fang, X., (2017). Flourishing sponge-based ecosystems after the end-Ordovician  
404 mass extinction. *Current Biology*, 27(4), 556-562.  
405 <https://doi.org/10.1016/j.cub.2016.12.061>.

406 Burton, M. (1934). *Sponges*. Great Barrier Reef Expedition, 1928–29. *Scientific*  
407 *Reports*, 4(14), 513-621.

408 Brayard, A., Krumenacker, L. J., Botting, J. P., Jenks, J. F., Bylund, K. G., Fara, E.,  
409 Vennin, E., Olivier, N., Goudemand, N., Saucède, T., Charbonnier, S., Romano,  
410 C., Doguzhaeva, L., Thuy, B., Hautmann, M., Stephen, D. A., Thomazo, C. &

411 Escarguel, G. (2017). Unexpected Early Triassic marine ecosystem and the rise  
412 of the Modern evolutionary fauna. *Science Advances*, 3(2), e1602159.  
413 <https://doi.org/10.1126/sciadv.1602159>

414 Chang, S., Zhang, L., Clausen, S., Bottjer, D. J., & Feng, Q. (2019). The  
415 Ediacaran-Cambrian rise of siliceous sponges and development of modern oceanic  
416 ecosystems. *Precambrian Research*, 333, 105438.  
417 <https://doi.org/10.1016/j.precamres.2019.105438>.

418 Chen, Z. Q., & Benton, M. J. (2012). The timing and pattern of biotic recovery  
419 following the end-Permian mass extinction. *Nature Geoscience*, 5(6), 375-383.  
420 <https://doi.org/10.1038/ngeo1475>.

421 Chen, Z. Q., Shi, G. R., & Kaiho, K. (2002). A new genus of rhynchonellid  
422 brachiopod from the Lower Triassic of South China and implications for timing the  
423 recovery of Brachiopoda after the end-Permian mass extinction. *Palaeontology*,  
424 45(1), 149-164. <https://doi.org/10.1111/1475-4983.00231>

425 Chen, Z. Q., Yang, H., Luo, M., Benton, M. J., Kaiho, K., Zhao, L. S., Huang, Y. G.,  
426 Zhang, K. X., Fang, Y. H., Jiang, H. S., Qiu, H., Li, Y., Tu, C. Y., Shi, L., Zhang, L.,  
427 Feng, X. Q., & Chen, L. (2015). Complete biotic and sedimentary records of the  
428 Permian–Triassic transition from Meishan section, South China: ecologically  
429 assessing mass extinction and its aftermath. *Earth-Science Reviews*, 149, 67-107.  
430 <https://doi.org/10.1016/j.earscirev.2014.10.005>

431 Chen, Z. Q., Fang, Y. H., Wignall, P. B., Guo, Z., Wu, S. Q., Liu, Z. L., Wang, R. Q.,  
432 Huang, Y. G., & Feng, X. Q. (2022). Microbial blooms triggered pyrite framboid

433 enrichment and oxygen depletion in carbonate platforms immediately after the  
434 latest Permian extinction. *Geophysical Research Letters*, 49(7), e2021GL096998.  
435 <https://doi.org/10.1029/2021GL096998>

436 Chin, Y. Y., Prince, J., Kendrick, G., & Abdul Wahab, M. A. (2020). Sponges in  
437 shallow tropical and temperate reefs are important habitats for marine invertebrate  
438 biodiversity. *Marine Biology*, 167(11), 164.  
439 <https://doi.org/10.1007/s00227-020-03771-1>

440 Dai, X., Davies, J. H., Yuan, Z., Brayard, A., Ovtcharova, M., Xu, G. H., Liu, X. K.,  
441 Smith, C. P. A., Schweitzer, C. E., Li, M. T., Perrot, M. G., Jiang, S. Y., Miao, L.  
442 Y., Cao, Y. R., Yan, J., Bai, R. Y., Wang, F. Y., Guo, W., Song, H. Y., Tian, L.,  
443 Corso, J. D., Liu, Y. T., Chu, D. L., & Song, H. (2023). A Mesozoic fossil  
444 lagerstätte from 250.8 million years ago shows a modern-type marine ecosystem.  
445 *Science*, 379(6632), 567-572. <https://doi.org/10.1126/science.adf1622>

446 Delecat, S., & Reitner, J. (2005). Sponge communities from the Lower Liassic of  
447 Adnet (Northern Calcareous Alps). *Facies*, 51, 385-404.  
448 <https://doi.org/10.1007/s10347-005-0045-x>

449 Delecat, S., Arp, G., & Reitner, J. (2011). Aftermath of the Triassic–Jurassic  
450 Boundary Crisis: Spiculite Formation on drowned Triassic steinplatte reef-slope  
451 by communities of hexactinellid sponges (Northern Calcareous Alps, Austria). In:  
452 Reitner, J., Queric, N.-V., & Arp, G. (eds), *Advances in Stromatolite Geobiology*.  
453 *Lecture Notes in Earth Sciences*, 131, 355-390.  
454 [https://doi.org/10.1007/978-3-642-10415-2\\_23](https://doi.org/10.1007/978-3-642-10415-2_23)

455 De Goeij, J. M., Van Oevelen, D., Vermeij, M. J., Osinga, R., Middelburg, J. J., De  
456 Goeij, A. F., & Admiraal, W. (2013). Surviving in a marine desert: the sponge loop  
457 retains resources within coral reefs. *Science*, 342(6154), 108-110.  
458 <https://doi.org/10.1126/science.1241981>.

459 Duckworth, A. R., West, L., Vansach, T., Stubler, A., & Hardt, M. (2012). Effects of  
460 water temperature and pH on growth and metabolite biosynthesis of coral reef  
461 sponges. *Marine Ecology Progress Series*, 462, 67-77.  
462 <https://doi.org/10.3354/meps09853>.

463 Ehrlich, H., Rigby, J. K., Botting, J., Tsurkan, M., Werner, C., Schwille, P., Petrášek,  
464 Z., Pisera, A., Simon, P., & Sivkov, V. (2013). Discovery of 505-million-year old  
465 chitin in the basal demosponge *Vauxia gracilentia*. *Scientific Reports*, 3(1), 3497.  
466 <https://doi.org/10.1038/srep03497>

467 Enos, P., Lehrmann, D. J., Jiayong, W., Youyi, Y., Jiafei, X., Chaikin, D. H., Minzoni,  
468 M., Berry, A. K., & Montgomery, P. (2006). Triassic evolution of the Yangtze  
469 platform in Guizhou Province, People's Republic of China, Geological Society of  
470 America, Boulder.

471 Enrichetti, F., Bavestrello, G., Betti, F., Coppari, M., Toma, M., Pronzato, R., Canese,  
472 S., Bertolino, M., Costa, G., Pansini, M., & Bo, M. (2020). Keratose-dominated  
473 sponge grounds from temperate mesophotic ecosystems (NW Mediterranean Sea).  
474 *Marine Ecology*, 41(6), e12620. <https://doi.org/10.1111/maec.12620>

475 Erpenbeck, D., Sutcliffe, P., Cook, S. D. C., Dietzel, A., Maldonado, M., van Soest, R.  
476 W., Hooper, J. N., & Wörheide, G. (2012). Horny sponges and their affairs: On the

477 phylogenetic relationships of keratose sponges. *Molecular Phylogenetics and*  
478 *Evolution*, 63(3), 809-816. <https://doi.org/10.1016/j.ympev.2012.02.024>

479 Ferrari, A. C., & Robertson, J. (2000). Interpretation of Raman spectra of disordered  
480 and amorphous carbon. *Physical review B*, 61(20), 14095.  
481 <https://doi.org/10.1103/PhysRevB.61.14095>.

482 Finks, R. M. (2010). Hypercalcified demosponges and the end-Permian extinction.  
483 *Global and Planetary Change*, 73(1-2), 141-148.  
484 <https://doi.org/10.1016/j.gloplacha.2010.03.009>.

485 Friesenbichler, E., Richoz, S., Baud, A., Krystyn, L., Sahakyan, L., Vardanyan, S.,  
486 Peckmann, J., Reitner, J., & Heindel, K. (2018). Sponge-microbial build-ups from  
487 the lowermost Triassic Chanakhchi section in southern Armenia: Microfacies and  
488 stable carbon isotopes. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 490,  
489 653-672. <https://doi.org/10.1016/j.palaeo.2017.11.056>.

490 Gabbott, S. E., Zalasiewicz, J., & Collins, D. (2008). Sedimentation of the phyllopod  
491 bed within the Cambrian Burgess Shale Formation of British Columbia. *Journal*  
492 *of the Geological Society*, 165(1), 307-318.

493 Gabbott, S. E., Zalasiewicz, J., Aldridge, R. J., & Theron, J. N. (2010). Eolian input  
494 into the Late Ordovician postglacial Soom Shale, South Africa. *Geology*, 38(12),  
495 1103-1106. <https://doi.org/10.1130/G31426.1>.

496 Gaines, R. R., Hammarlund, E. U., Hou, X., Qi, C., Gabbott, S. E., Zhao, Y., Peng, J.,  
497 & Canfield, D. E. (2012). Mechanism for Burgess Shale-type preservation.  
498 *Proceedings of the National Academy of Sciences, United States of America*,

499 109(14), 5180-5184. <https://doi.org/10.1073/pnas.1111784109>

500 Gaines, R. R., Kennedy, M. J., & Droser, M. L. (2005). A new hypothesis for organic  
501 preservation of Burgess Shale taxa in the middle Cambrian Wheeler Formation,  
502 House Range, Utah. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 220(1-2),  
503 193-205. <https://doi.org/10.1016/j.palaeo.2004.07.034>.

504 Gaino, E., Scalera Liaci, L., Sciscioli, M., & Corriero, G. (2006). Investigation of the  
505 budding process in *Tethya citrina* and *Tethya aurantium* (Porifera, Demospongiae).  
506 *Zoomorphology*, 125(2), 87. <https://doi.org/10.1007/s00435-006-0015-z>.

507 Grant, R. E., & Todd, R. B. (1836). *Animal Kingdom. The Encyclopaedia of Anatomy*  
508 *and Physiology*, 1, 107-118.

509 Guo, Z., Chen, Z. Q., Harper, D. A. T., & Huang, Y. G. (2022). Permian–Triassic  
510 phylogenetic and morphologic evolution of rhynchonellide brachiopods.  
511 *Paleobiology*, 48(1), 99-119. <https://doi.org/10.1017/pab.2021.25>

512 Harper, D. A.T., Hammarlund, E. U., Topper, T. P., Nielsen, A. T., Rasmussen, J. A.,  
513 Park, T. Y. S., & Smith, M. P. (2019). The Sirius Passet Lagerstätte of North  
514 Greenland: a remote window on the Cambrian explosion. *Journal of the Geological*  
515 *Society*, 176(6), 1023-1037. <https://doi.org/10.1144/jgs2019-043>.

516 Hatch, J. R., & Leventhal, J. S. (1992). Relationship between inferred redox potential  
517 of the depositional environment and geochemistry of the Upper Pennsylvanian  
518 (Missourian) Stark Shale Member of the Dennis Limestone, Wabaunsee County,  
519 Kansas, USA. *Chemical Geology*, 99(1-3), 65-82.  
520 [https://doi.org/10.1016/0009-2541\(92\)90031-Y](https://doi.org/10.1016/0009-2541(92)90031-Y).

521 He, Z. L., Li, S. J., Li, Y. Q., & Gao, J. (2022). Multi-directional and multiphase  
522 tectonic modification, and hydrocarbon differential enrichment in the middle-Upper  
523 Yangtze Region. *Journal of Earth Science*, 33(5), 1246–1259. <https://doi.org/10.1007/s12583-022-1738-6>

524

525 Heindel, K., Foster, W. J., Richoz, S., Birgel, D., Roden, V. J., Baud, A., Brandner, R.,  
526 Krystyn, L., Mohtat, T., Koşun, E., Twitchett, J. R., Reitner, J., & Peckmann, J.  
527 (2018). The formation of microbial-metazoan bioherms and biostromes following  
528 the latest Permian mass extinction. *Gondwana Research*, 61, 187-202.  
529 <https://doi.org/10.1016/j.gr.2018.05.007>

530 Hogg, M. M., Tendal, O. S., Conway, K. W., Pomponi, S. A., Van Soest, R. W. M.,  
531 Gutt, J., Krautter, M., & Roberts, J. M. (2010). Deep-sea sponge grounds:  
532 reservoirs of biodiversity. *UNEP-WCMC Biodiversity Series*, 32, 1-86.

533 Hoffmann, F., Larson, O., Thiel, V., Rapp, H. T., Pape, T., Michaelis, W., & Reitner, J.  
534 (2005). An anaerobic world in sponges. *Geomicrobiology Journal*, 22, 1-10.  
535 <https://doi.org/10.1080/01490450590922505>

536 Huang, Y. G., Chen, Z. Q., Wignall, P. B., & Zhao, L. (2017). Latest Permian to  
537 Middle Triassic redox condition variations in ramp settings, South China: Pyrite  
538 framboid evidence. *Geological Society of America Bulletin*, 129(1-2), 229-243.  
539 <https://doi.org/10.1130/B31458.1>

540 Huang, Y. G., Chen, Z. Q., Algeo, T. J., Zhao, L., Baud, A., Bhat, G. M., Zhang, L., &  
541 Guo, Z. (2019). Two-stage marine anoxia and biotic response during the  
542 Permian–Triassic transition in Kashmir, northern India: pyrite framboid evidence.



543 Global and Planetary Change, 172, 124-139.  
544 <https://doi.org/10.1016/j.gloplacha.2018.10.002>

545 Huang, Y., Chen, Z. Q., Roopnarine, P. D., Benton, M. J., Zhao, L., Feng, X., & Li, Z.  
546 (2023). The stability and collapse of marine ecosystems during the  
547 Permian-Triassic mass extinction. *Current Biology*, 33(6), 1059-1070.  
548 <https://doi.org/10.1016/j.cub.2023.02.007>

549 Jesionowski, T., Norman, M., Żółtowska-Aksamitowska, S., Petrenko, I., Joseph, Y.,  
550 & Ehrlich, H. (2018). Marine spongin: Naturally prefabricated 3D scaffold-based  
551 biomaterial. *Marine Drugs*, 16(3), 88. <https://doi.org/10.3390/md16030088>.

552 Jiang, Y., Wu, H. T., Xiao, Y. F., Yang, T. L., He, W. H., Wang, H., Sun, X., & Wu, S.  
553 B. (2018), Ammonoid zonation of Permian-Triassic boundary strata from Kejiao  
554 section, Huishui County, southern Guizhou Province, South China and regional  
555 correlation: *Geological Science and Technological Information*, 37, 65-73.  
556 <https://doi.org/10.19509/j.cnki.dzkq.2018.0608>.

557 Jing, Y. H., Chen, Z. Q., & Tu, C. Y. (2022). A late Paleoproterozoic microfossil  
558 community from siliceous granules, Dahongyu Formation, North China.  
559 *Precambrian Research*, 377, 106723.  
560 <https://doi.org/10.1016/j.precamres.2022.106723>.

561 Jones, B., & Manning, D. A. (1994). Comparison of geochemical indices used for the  
562 interpretation of palaeoredox conditions in ancient mudstones. *Chemical geology*,  
563 111(1-4), 111-129. [https://doi.org/10.1016/0009-2541\(94\)90085-X](https://doi.org/10.1016/0009-2541(94)90085-X).

564 Kelmo, F., Bell, J. J., & Attrill, M. J. (2013). Tolerance of sponge assemblages to

565 temperature anomalies: resilience and proliferation of sponges following the  
566 1997–8 El-Nino southern oscillation. PLoS One, 8(10), e76441.  
567 <https://doi.org/10.1371/journal.pone.0076441>.

568 Lee, J. H., & Riding, R. (2021). The ‘classic stromatolite’ *Cryptozoön* is a keratose  
569 sponge-microbial consortium. *Geobiology*, 19(2), 189-198.  
570 <https://doi.org/10.1111/gbi.12422>

571 Lee, J. H., & Riding, R. (2023). Recognizing sponge in *Spongiostroma* Gürich, 1906  
572 from the Mississippian of Belgium. *Journal of Paleontology*, 97(1), 26-37.  
573 <https://doi.org/10.1017/jpa.2022.73>

574 Lehrmann, D. J., Payne, J. L., Felix, S. V., Dillett, P. M., Wang, H., Yu, Y., & Wei, J.  
575 (2003). Permian–Triassic boundary sections from shallow-marine carbonate  
576 platforms of the Nanpanjiang Basin, South China: implications for oceanic  
577 conditions associated with the end-Permian extinction and its aftermath. *Palaios*,  
578 18(2), 138-152.  
579 [https://doi.org/10.1669/0883-1351\(2003\)18<138:PBSFSC>2.0.CO;2](https://doi.org/10.1669/0883-1351(2003)18<138:PBSFSC>2.0.CO;2).

580 Lesser, M. P., & Slattery, M. (2020). Will coral reef sponges be winners in the  
581 Anthropocene?. *Global Change Biology*, 26(6), 3202-3211.  
582 <https://doi.org/10.1111/gcb.15039>

583 Leys, S. P., Yahel, G., Reidenbach, M. A., Tunnicliffe, V., Shavit, U., & Reiswig, H.  
584 M. (2011). The sponge pump: the role of current induced flow in the design of the  
585 sponge body plan. *PloS one*, 6(12), e27787.  
586 <https://doi.org/10.1371/journal.pone.0027787>

587 Li, L., Feng, H., Janussen, D., & Reitner, J. (2015). Unusual deep water sponge  
588 assemblage in South China—witness of the end-Ordovician mass extinction.  
589 *Scientific Reports*, 5, 16060. <https://doi.org/10.1038/srep16060>.

590 Li, Z. H., Zhang, M., Chen, Z. Q., Algeo, T. L., Zhao, L. S., & Zhang, F. F. (2021).  
591 Early Cambrian oceanic oxygenation and evolution of early animals: A critical  
592 review from the South China Craton. *Global and Planetary Change*, 204, 103561.  
593 <https://doi.org/10.1016/j.gloplacha.2021.103561>

594 Luo, C., & Reitner, J. (2014). First report of fossil “keratose” demosponges in  
595 Phanerozoic carbonates: preservation and 3-D reconstruction. *Naturwissenschaften*,  
596 101, 467-477. <https://doi.org/10.1007/s00114-014-1176-0>.

597 Luo, C., Pei, Y., Richoz, S., Li, Q., and Reitner, J., 2022, Identification and current  
598 palaeobiological understanding of “Keratosa”-type nonspicular demosponge fossils  
599 in carbonates: with a new example from the lowermost Triassic, Armenia. *Life*, 12,  
600 1348. <https://doi.org/10.3390/life12091348>.

601 Luo, C., Zhao, F., & Zeng, H. (2020). The first report of a vauxiid sponge from the  
602 Cambrian Chengjiang Biota. *Journal of Paleontology*, 94(1), 28-33.  
603 <https://doi.org/10.1017/jpa.2019.52>.

604 Mills, D. B., Ward, L. M., Jones, C., Sweeten, B., Forth, M., Treusch, A. H., &  
605 Canfield, D. E. (2014). Oxygen requirements of the earliest animals. *Proceedings*  
606 *of the National Academy of Sciences, United States of America*, 111(11),  
607 4168-4172. <https://doi.org/10.1073/pnas.1400547111>.

608 Minchin, E. A., & Lankester, E. R. (1900). Chapter III. Sponges. *A Treatise on*

609 Zoology. Part II. The Porifera and Coelenterata, 2, 1-178. Neuweiler, F., Kershaw,  
610 S., Boulvain, F., Matysik, M., Sendino, C., McMenamin, M., & Munnecke, A.  
611 (2023). Keratose sponges in ancient carbonates – A problem of interpretation.  
612 Sedimentology, 70, 927-968. <https://doi.org/10.1111/sed.13059>

613 Pierrejean, M., Grant, C., Neves, B. D. M., Chaillou, G., Edinger, E., Blanchet, F. G.,  
614 Maps, F., Nozais, C., & Archambault, P. (2020). Influence of deep-water corals and  
615 sponge gardens on infaunal community composition and ecosystem functioning in  
616 the Eastern Canadian Arctic. *Frontiers in Marine Science*, 7, 495.  
617 <https://doi.org/10.3389/fmars.2020.00495>

618 Rigby, J. K. (1980). The new Middle Cambrian sponge *Vauxia magna* from the  
619 Spence Shale of northern Utah and taxonomic position of the Vauxiidae. *Journal of*  
620 *Paleontology*, 54, 234-240.

621 Rigby, J.K., & Collins, D. (2004). Sponges of the Middle Cambrian Burgess Shale  
622 and Stephen Formations, British Columbia. Toronto, Royal Ontario Museum,  
623 1-155.

624 Ritterbush, K. A., Rosas, S., Corsetti, F. A., Bottjer, D. J., & West, A. J. (2015).  
625 Andean sponges reveal long-term benthic ecosystem shifts following the  
626 end-Triassic mass extinction. *Palaeogeography, Palaeoclimatology, Palaeoecology*,  
627 420, 193-209. <https://doi.org/10.1016/j.palaeo.2014.12.002>.

628 Rooks, C., Fang, J. K. H., Mørkved, P. T., Zhao, R., Rapp, H. T., Xavier, J. R., &  
629 Hoffmann, F. (2020). Deep-sea sponge grounds as nutrient sinks: denitrification is  
630 common in boreo-Arctic sponges. *Biogeosciences*, 17(5), 1231-1245.

631 <https://doi.org/10.5194/bg-17-1231-2020>

632 Schieber, J., Southard, J. B., & Schimmelmann, A. (2010). Lenticular shale fabrics  
633 resulting from intermittent erosion of water-rich muds—interpreting the rock  
634 record in the light of recent flume experiments. *Journal of Sedimentary Research*,  
635 80(1), 119-128. <https://doi.org/10.2110/jsr.2010.005>.

636 Scotese, C. R., (2014). Paleomap Project: Early Triassic Paleomap. Updated at:  
637 <https://www.scotese.com/newpage8.htm>

638 Shen, S. Z., Fan, J. X., Wang, X. D., Zhang, F. F., Shi, Y. K., & Zhang, S. H. (2022).  
639 How to build a high-resolution digital geological timeline? *Journal of Earth  
640 Science*, 33, 1629-1632. <https://doi.org/10.1007/s12583-022-1315-z>

641 Shi, L., Feng, Q. L., Shen, J., Ito, T., & Chen, Z. Q. (2016). Proliferation of  
642 shallow-water radiolarians coinciding with enhanced oceanic productivity in  
643 reducing conditions during the Middle Permian, South China: evidence from the  
644 Gufeng Formation of western Hubei Province. *Palaeogeography,  
645 Palaeoclimatology, Palaeoecology*, 444, 1-14.  
646 <https://doi.org/10.1016/j.palaeo.2015.11.031>

647 Simion, P., Philippe, H., Baurain, D., Jager, M., Richter, D. J., Di Franco, A., Roure,  
648 B., Satoh, N., Quéinnec, É., & Ereskovsky, A. (2017). A large and consistent  
649 phylogenomic dataset supports sponges as the sister group to all other animals.  
650 *Current biology*, 27(7), 958-967. <https://doi.org/10.1016/j.cub.2017.02.031>.

651 Slattery, M., Gochfeld, D. J., Easson, C. G., & O'Donahue, L. R. (2013). Facilitation  
652 of coral reef biodiversity and health by cave sponge communities. *Marine Ecology*

653 Progress Series, 476, 71-86. <https://doi.org/10.3354/meps10139>.

654 Sollas, W. (1875). Sponges *in* Encyclopaedia Britannica, ninth edition. London.

655 Sun, W. D., & Zhang, L. P. (2022). Preface: Pacific plate subduction and the  
656 Yanshanian movement in Eastern China. *Journal of Earth Science*, 33(3), 541–543.  
657 <https://doi.org/10.1007/s12583-022-1311-3>

658 Turner, E. C. (2021). Possible poriferan body fossils in early Neoproterozoic  
659 microbial reefs. *Nature*, 596(7870), 87-91.  
660 <https://doi.org/10.1038/s41586-021-03773-z>.

661 Velledits, F. (2008). Evolution of the Triassic reef communities. *Hantkeniana*, 6, 9-16.

662 Vishnevskaya, V., Pisera, A., & Racki, G. (2002). Siliceous biota [radiolarians and  
663 sponges] and the Late Devonian biotic crisis: The Polish reference. *Acta*  
664 *Palaeontologica Polonica*, 47(2), 211-226.

665 Walcott, C. D. (1920). *Cambrian Geology and Paleontology: Middle Cambrian*  
666 *Spongia*, Smithsonian Institution.

667 Wang, H., He, W. H., Xiao, Y. F., Yang, T. L., Zhang, K. X., Wu, H. T., Huang, Y.F.,  
668 Peng, X. F., & Wu, S. B. (2023). Stagewise collapse of biotic communities and its  
669 relations to oxygen depletion along the north margin of Nanpanjiang Basin during  
670 the Permian–Triassic transition. *Palaeogeography, Palaeoclimatology,*  
671 *Palaeoecology*, 621, 111569. <https://doi.org/10.1016/j.palaeo.2023.111569>

672 Wiedenmayer, F. (1977). Shallow-water sponges of the western Bahamas: *Experientia*  
673 *Supplementum*, 1-287.

674 Wu, S. Q., Chen, Z. Q., Su, C. M., Fang, Y. H., & Yang, H. (2022a). Keratose sponge

675 fabrics from the lowermost Triassic microbialites in South China: Geobiologic  
676 features and Phanerozoic evolution. *Global and Planetary Change*, 211, 103787.  
677 <https://doi.org/10.1016/j.gloplacha.2022.103787>.

678 Wu, S. Q., Chen, Z. Q., Fang, Y. H., Pei, Y., & Foster, W. J. (2022b). Benthic  
679 Pleurocapsales (cyanobacteria) blooms catalyzing carbonate precipitation and  
680 dolomitization following the end-Permian mass extinction. *Geophysical Research*  
681 *Letters*, 49, e2022GL100819. <https://doi.org/10.1029/2022GL100819>

682 Yang, X., Zhao, Y., Babcock, L. E., & Peng, J. I. N. (2017). A new vauxiid sponge  
683 from the Kaili Biota (Cambrian Stage 5), Guizhou, South China. *Geological*  
684 *Magazine*, 154(6), 1334-1343. [10.1017/S0016756816001229](https://doi.org/10.1017/S0016756816001229).

685 Yin, H. F., Zhang, K. X., Tong, J. N., Yang, Z. Y., & Wu, S. B. (2001). The global  
686 stratotype section and point (GSSP) of the Permian-Triassic boundary. *Episodes*,  
687 24(2), 102-114. <https://doi.org/10.18814/epiiugs/2001/v24i2/004>

688 Zea, S., Henkel, T. P., & Pawlik, J.R. (2014). *The Sponge Guide: A Picture Guide to*  
689 *Caribbean Sponges*. Available online at [www.spongeguide.org](http://www.spongeguide.org).

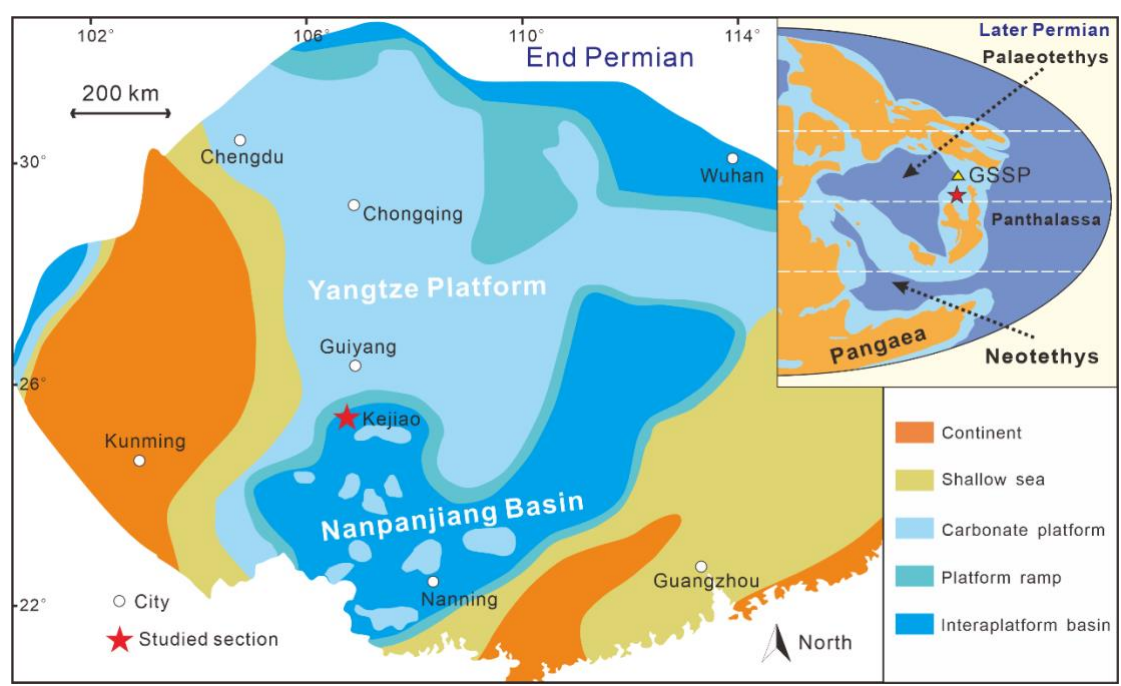
690 Zhang, F. F., Algeo, T. J., Romaniello, S. J., Cui, Y., Zhao, L., Chen, Z. Q., & Anbar,  
691 A. D. (2018). Congruent Permian-Triassic  $\delta^{238}\text{U}$  records at Panthalassic and  
692 Tethyan sites: Confirmation of global-oceanic anoxia and validation of the  
693 U-isotope paleoredox proxy. *Geology*, 46(4), 327-330.  
694 <https://doi.org/10.1130/G39695.1>.

695 Zheng, S., Feng, Q., van de Velde, S., Chang, S., Zhang, L., & Gao B. (2022).  
696 Microfossil assemblages and indication of the source and preservation pattern of

697 organic matter from the Early Cambrian in South China. *Journal of Earth Science*,  
698 33(3), 802–819. <https://doi.org/10.1007/s12583-020-1117-0>

699  
700  
701

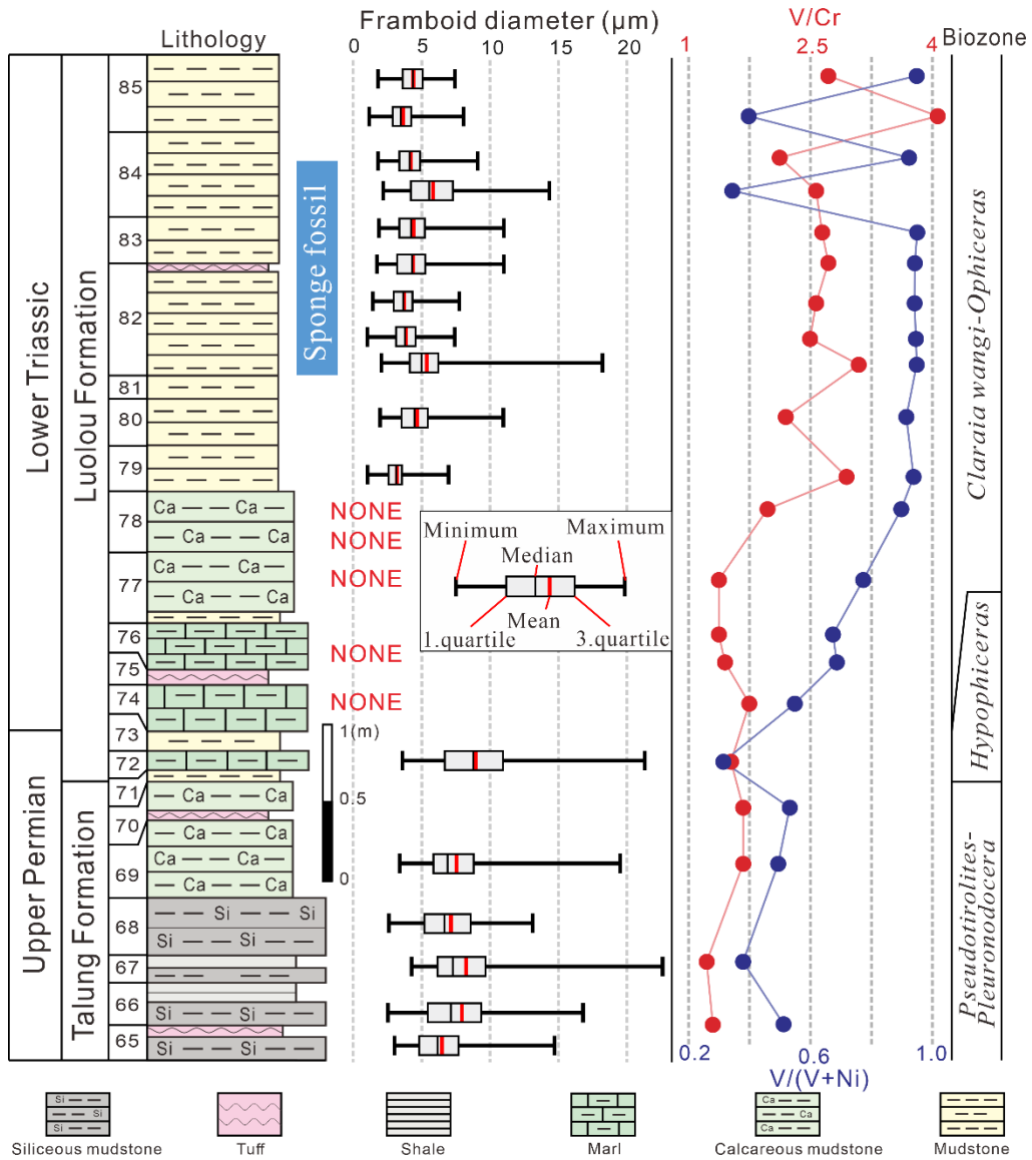
702 **FIGURE CAPTIONS**



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**Figure 1** The later Permian paleogeographic map of the Paleo-Tethys (Scotese, 2014) and South China (Enos et al., 2006; Sun & Zhang, 2022) showing the site of the Kejiao section.





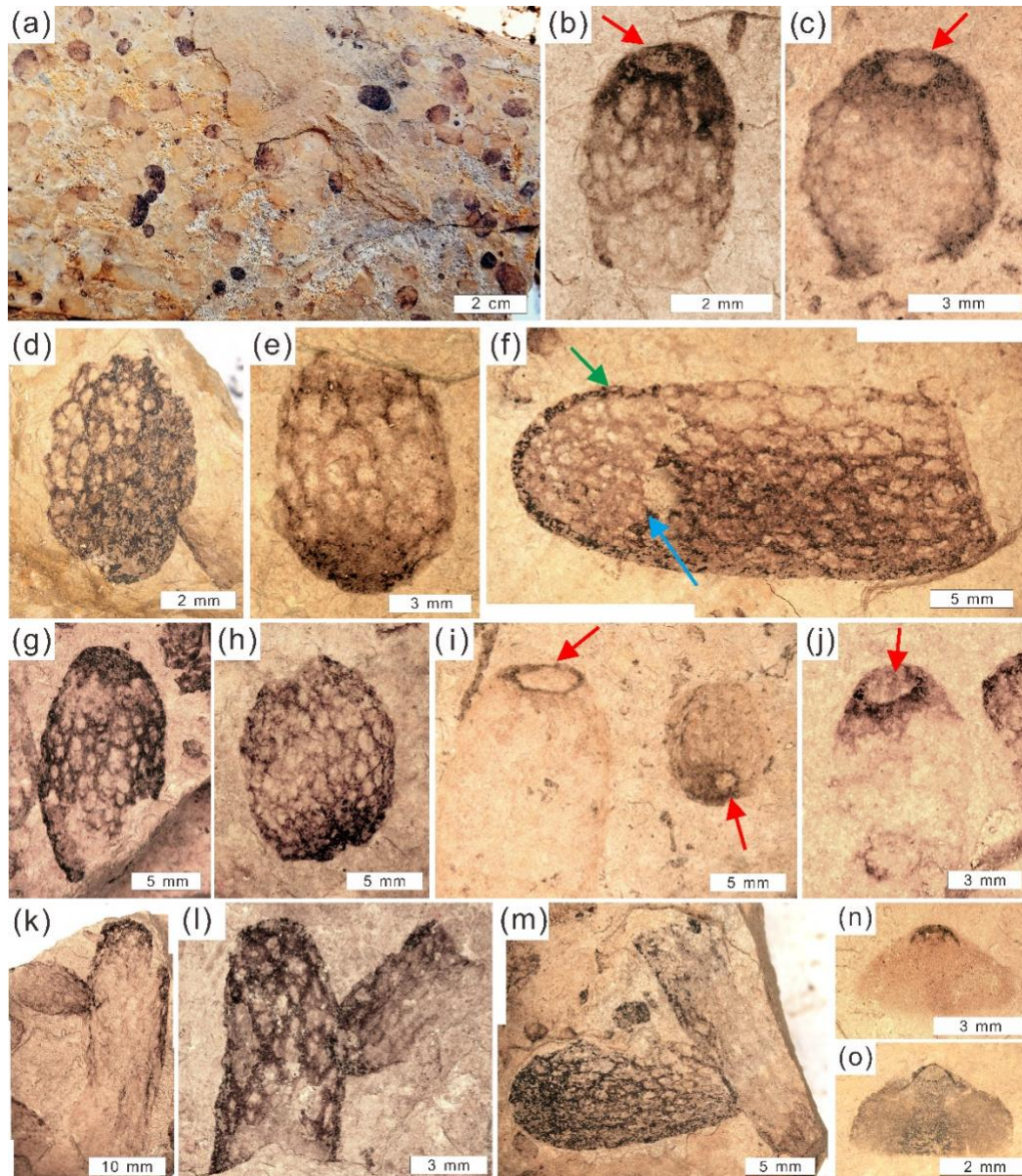
707 **Figure 2** The P–Tr boundary succession exposed at Kejiao showing lithology, fossil

708 distribution, pyrite framboid size distributions, and trace element ratios.

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713 **Figure 3** Selected specimens of the sponge and brachiopod fossils from the lowest

714 Triassic in the Kejiao section. (a) Distribution of sponge fossils on the bedding

715 surface. (b-j) Selected sponge specimens, red arrows point out the oscula (BGEG

716 KJ10304-10313). (f) A broken specimen (BGEG KJ10305) showing thickened walls

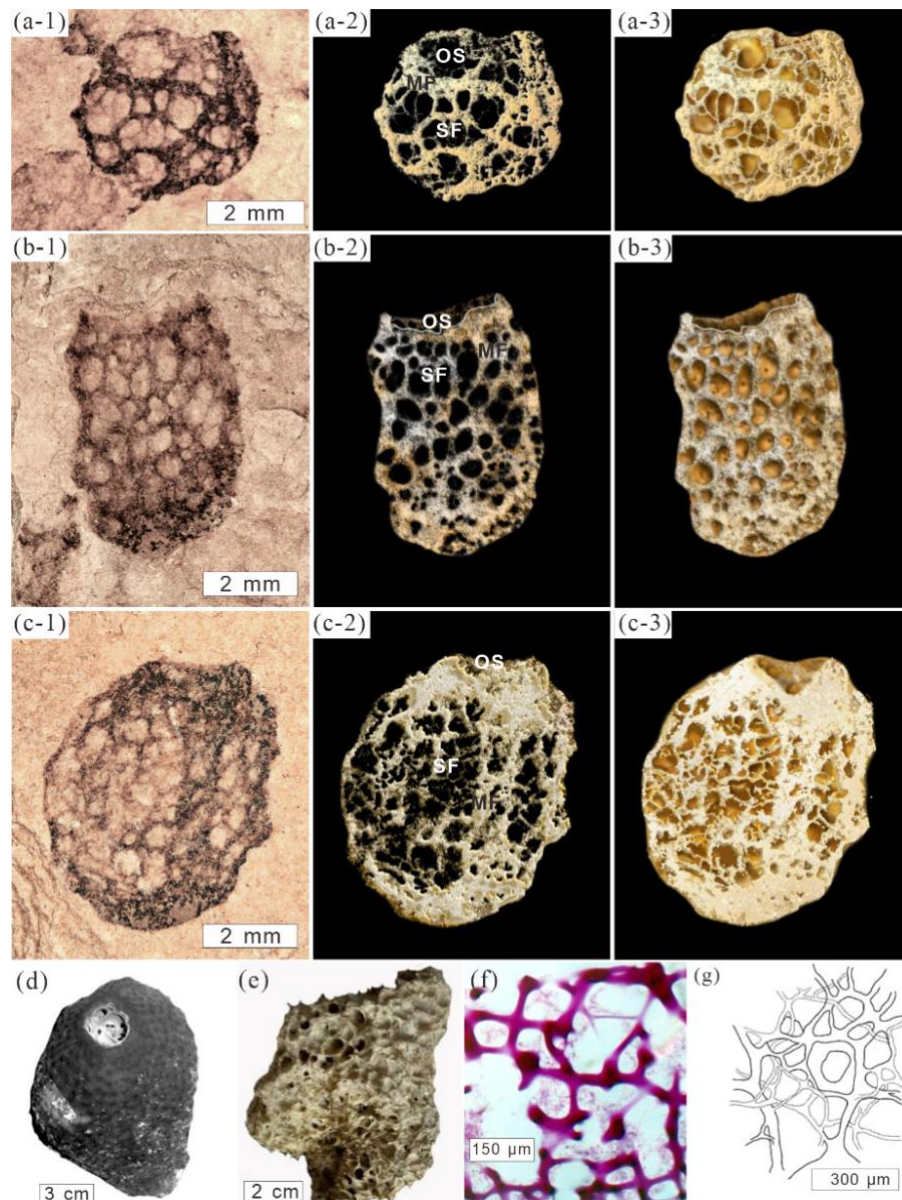
717 (green arrow) and interior view of intake network (blue arrow). (k-m) Branching

718 sponges (BGEG KJ10314-10316, Paratype III) indicating a budding model of

719 reproduction. (n-o) Brachiopod *Meishanorhynchia* shells associated with sponges on

720 the same slab.

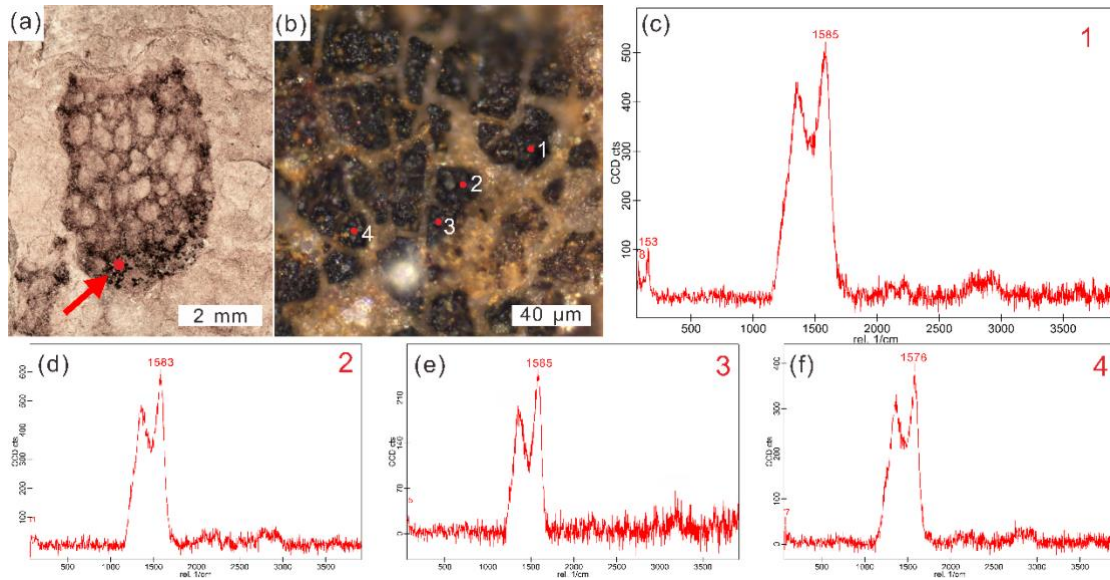




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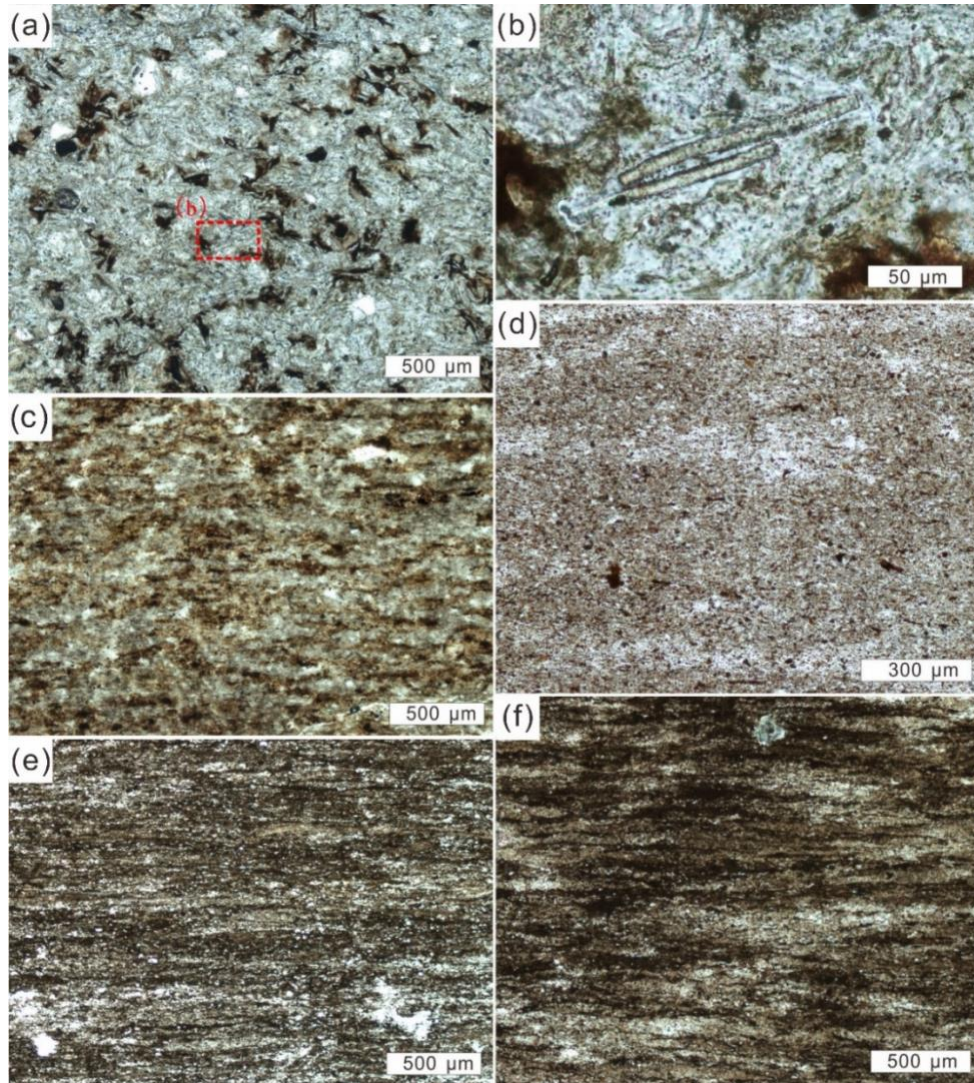
722 **Figure 4.** Fossil keratose sponges and their modern analogues. (a) A complete  
 723 specimen (BGEG KJ10301; Holotype) in optical microscopic view (a-1) and  
 724 reconstructions (a-2; a-3). (b) A complete specimen (BGEG KJ10302; Paratype I) in  
 725 optical microscopic view (b-1) and reconstructions (b-2; b-3). (c) A complete  
 726 specimen (BGEG KJ10303; Paratype II) (c-1), reconstructions (c-2; c-3). (a-2, b-2,  
 727 c-2) are directly extracted image and (a-3, b-3, c-3) are 3D reconstructions. OS =  
 728 osculum, MF = primary fiber, SF = secondary fiber. (d) *Smenospongia conulosa*  
 729 Pulizer-Finali, 1986; picture modified after *The Sponge Guide* (Zea et al., 2014). (e)

730 *Smenospongia* sp., dried specimen. (f) Stained spongin skeleton of *Smenospongia* sp.  
 731 showing thick primary and thin secondary fibers. (g) Drawing of the fiber skeleton of  
 732 *Smenospongia conulosa*; modified after *The Sponge Guide* (Zea et al., 2014).  
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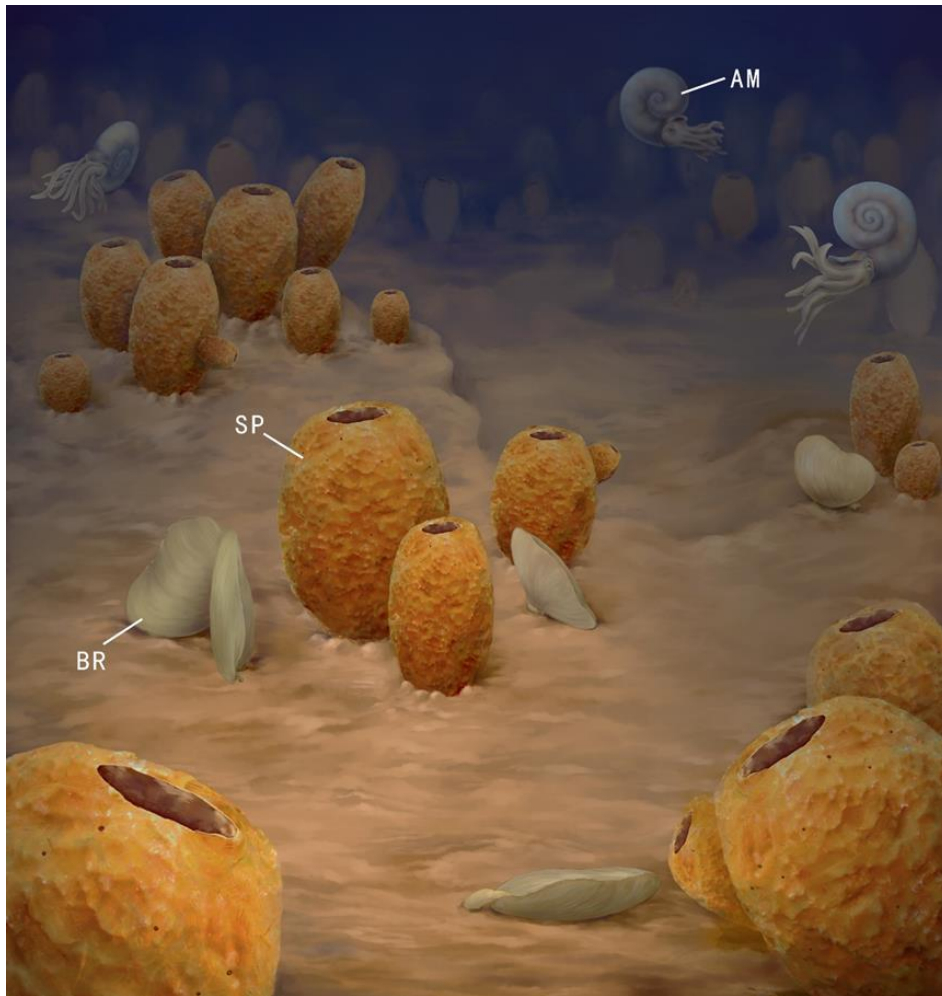
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 735 **Figure 5** (a) Selected sponge fossil (BGEK KJ10302) for Raman spectrum analysis.  
 736 (b) Close-up of the dark sponge remains in (a) marked with Raman spectrum points  
 737 1-4. (c-f) Raman spectrum of points 1-4.  
 738





739

740 **Figure 6** Microphotographs showing microfacies of the lowest Triassic strata in the  
 741 Kejiao section. (a) Siliceous rock containing abundant sponge spicules (bed 65). (b)  
 742 Close-up of boxed area in a showing details of sponge spicules. (c) Calcareous  
 743 mudstone with oriented mineral arrangement (bed 69). (d) Slightly oriented Mineral  
 744 arrangement (bed 81). (e) Horizontal bedding (bed 82). (f) Horizontal bedding (Bed  
 745 85).



746

747 **Figure 7** Artistic reconstruction of the marine ecosystem immediately after the  
748 end-Permian mass extinction in Kejiao section showing abundant keratose sponges  
749 and associated brachiopods and ammonoids. SP = sponge, BR = brachiopod, AM =  
750 ammonoid.

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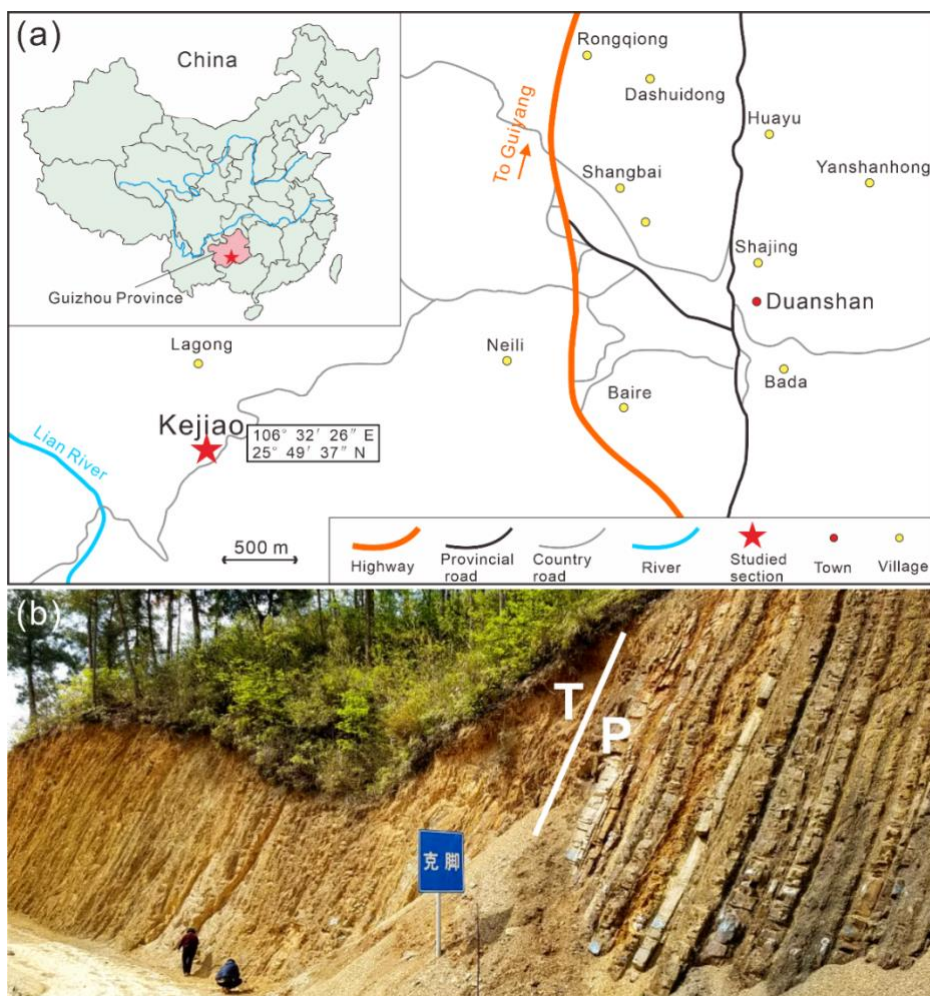


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## Supplemental Material for

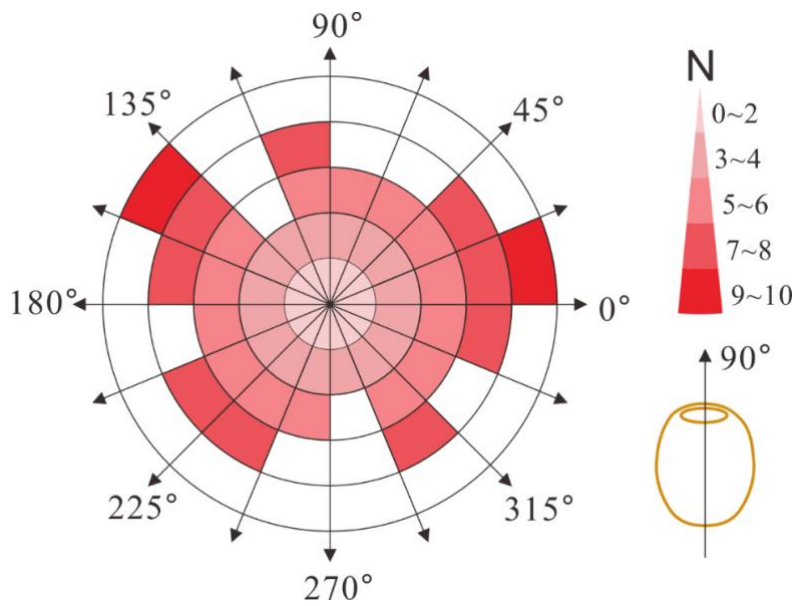
New keratose sponges after the end-Permian extinction provide insights into biotic recoveries

Siqi Wu<sup>1,2</sup>, Joachim Reitner<sup>3\*</sup>, Zhong-Qiang Chen<sup>2\*</sup>, David A. T. Harper<sup>2,4</sup>, and Jianxin Yu<sup>2</sup>



7 **Supplementary Figure S1.** (a) Location of the Kejiao section, Huishui County,  
8 Guizhou Province, South China. (b) Field photo showing the succession of the  
9 uppermost Permian Dalong Formation to the lowest Triassic Daye Formation in the  
10 Kejiao section. Note the Permian-Triassic boundary is marked as P/T in the field  
11 photo.

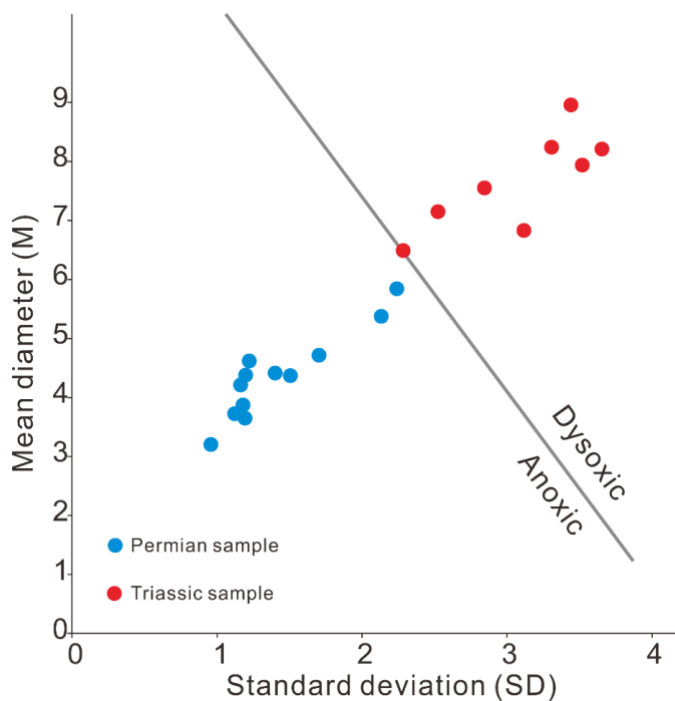
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13 **Supplementary Figure S2.** Rose diagram shows orientation of sponge specimens  
14 distributed on the surface of one slab. N=104.

15

16



17 **Supplementary Figure S3.** M-SD plot of mean diameter versus standard deviation of  
18 pyrite framboid sizes for the Kejiao section (see pyrite framboid size distributions in  
19 Figure 2).



20 **Relationship of the new fossil keratose sponge**

21 The new taxon exhibits similarities of the fiber structure, especially the arrangement  
22 of thick primary and thinner secondary fibers, with modern taxa related to the  
23 Dictyoceratida (Minchin 1900) and specially with the Thorectidae erected by  
24 Bergquist 1978. The Thorectidae have various growth forms and show often vase- and  
25 mound shaped morphologies. In modern taxa the choanocyte chambers are  
26 characteristic and taxon specific which are not preserved in the fossil representatives.  
27 The spongin fibers of the Thorectidae often form an anastomosing skeleton (Fig. 4e,  
28 g). The central region of the primary thick fibers often shows a diffuse area often  
29 enriched with agglutinated material which is, however, not observed in the new  
30 sponge. Many primary fibers are often extremely thick and arranged with thinner,  
31 maybe secondary fibers, and form basket-like structures (Figures. 3-4). This spongin  
32 fiber arrangement is often seen in the new fossil sponge and resemble strongly to the  
33 fiber skeleton known from the Thorectidae. This was the main reason to classify the  
34 new taxon within the Thorectidae. The new fossil sponge is most similar to the  
35 modern genus *Smenospongia* first described by Wiedenmayer (1977) (Figure 4d-g).  
36 The irregular arrangements of the primary thick and thinner secondary fibers and  
37 prominent large oscular pores on top of the sponge baskets (Figures 3b-c, i-j, n, o; 4a-  
38 c, d) support this alignment.

39

40 **Phylogenetic status of the aspiculate “keratose” demosponges.**

41 The term “Keratosa” or “horny sponges” was introduced by Grant (1861), and was  
42 further elaborated by several workers (Bowerbank, 1866; Lendenfeld, 1889; Minchin,  
43 1900). Biochemical and molecular phylogenetic data reveal that these sponges are  
44 probably the sister group of the spicular demosponges (Erpenbeck et al., 2020;  
45 Erpenbeck et al., 2012; Lavrov et al., 2008). Aspiculate demosponges are categorized  
46 as two clades: the “Keratosa” and Myxospongida. The former includes the orders  
47 Dictyoceratida and Dendroceratida, exhibiting organic skeletons made of spongin.  
48 Spongin is an enigmatic proteinaceous material that contains halogenated residues and

49 until recently had not been sequenced (Ehrlich, 2019). The latter embraces the orders  
50 Verongida, Halisarcida and Chondrosida, and forms chitinous fiber skeletons (Ehrlich  
51 et al., 2010; Ehrlich et al., 2013; Morrow and Cárdenas, 2015; Vacelet et al., 2019).  
52 Due to the widespread homoplasy in the skeletal morphology of these sponges  
53 (Erpenbeck et al., 2020; Vacelet et al., 2019) and the loss of biochemical information  
54 during taphonomic processes, it is hard to differentiate the two clades in fossil record.

55

### 56 **Description of Holotype and Paratypes I, II and III**

57

#### 58 **Holotype**

59 BGEG KJ10301. The holotype (Figure 4a) is more or less spherical in outline, with 5  
60 mm in width/horizontal diameter and 4.6 mm in vertical height. The main osculum  
61 has a diameter of nearly 2 mm. The margin of the oscular area is enhanced, typical for  
62 the main opening and shows an inner ring structure. The entire sponge skeleton looks  
63 like a basket. The organic skeleton fibers exhibit only one distinct front and rear  
64 network layer, caused by taphonomic compression of the sponge body. The organic  
65 fibers show clearly the main thick fibers of 200-250  $\mu\text{m}$  in diameter. The thinner ones,  
66 developed from the thicker ones, are 50-150  $\mu\text{m}$  in diameter. Generally, the fibers are  
67 crowded and anastomose. The spaces between the fibers show an average diameter of  
68 400  $\mu\text{m}$ –1mm. The fibers are composed of kerogenous-like black organic carbon  
69 without any allochthonous inclusions or spicules.

70

#### 71 **Paratype I**

72 BGEG KJ10302. Paratype I (Figure 4b), 1.2 cm high and 0.8 cm wide, is larger and  
73 more elongated than the holotype. The network of the preserved fibers exhibits the same  
74 dimensions as the holotype. The fibers also anastomose and show thick and thinner  
75 ones. The spaces between the fibers show an average diameter of 0.5–1 mm. The central  
76 oscular area is not completely preserved and with a diameter of 5.3 mm wider than  
77 expected. The base of the sponge exhibits an enhancement of the fiber network and this

78 feature is interpreted as hold fast character. The fibers are also preserved in black  
79 organic carbon (kerogenous material).

80

### 81 **Paratype II**

82 BGEG KJ10303. Paratype II (Figure 4c), 1.3 cm wide and 1.53 cm high, is larger and  
83 more elongated than the holotype and paratype I. The network of the preserved fibers  
84 exhibits the same dimensions as the holotype and paratype I. The fibers also anastomose  
85 and show thick and thinner ones. The spaces between the fibers show an average  
86 diameter of 0.5–1 mm. However, the central part of the sponge is not very well  
87 preserved and the fiber skeleton is due to incomplete preservation. However, the central  
88 osculum is completely preserved and with a diameter of 4 mm. The margin of the  
89 oscular area is enhanced, typical for this opening. The base of the sponge also shows  
90 an enhancement of the fiber network and this feature is interpreted as a hold fast. The  
91 fibers are also preserved in black organic carbon (kerogenous material).

92

### 93 **Paratype III**

94 BGEG KJ103014. Paratype III (Figure 3i) has the appearance of a tower and has a  
95 height of 1 cm and a width of 5 mm. The oscular area is developed and the main  
96 osculum has diameter of 2 mm. The base of the sponge is irregular and fragmented. The  
97 spaces between the fibers vary from 300  $\mu\text{m}$  to 1mm. Most of the fibers are between  
98 100-250  $\mu\text{m}$  thick. Thin fibres are rare. Part of the left fiber skeleton is damaged  
99 together with the basal areas. The sponge shows, on its right side a probable branch  
100 possibly representing an asexual bud. The base of the contact area with the mother  
101 sponge is well developed and does not exhibit anastomosing fibers. The branch is 6  
102 high and 3 mm wide. The central osculum is not preserved and the top area is  
103 fragmented. The right lower part of the branch is not well preserved and the fiber  
104 structure is excised.

105

### 106 **REFERENCES CITED**

107 Bergquist, P. (1978). Sponges: University of California Press.

108 Bowerbank, J.S. (1866). A monograph of the British Spongiadae: The Ray Society,  
109 London.

110 Ehrlich, H. (2019). Marine biological materials of invertebrate origin (Vol. 13). Cham:  
111 Springer International Publishing.

112 Ehrlich, H., Ilan, M., Maldonado, M., Muricy, G., Bavestrello, G., Kljajic, Z.,  
113 Carballo, J., Schiaparelli, S., Ereskovsky, A., & Schupp, P. (2010). Three-  
114 dimensional chitin-based scaffolds from Verongida sponges (Demospongiae:  
115 Porifera). Part I. Isolation and identification of chitin. International Journal of  
116 Biological Macromolecules, 47(2), 132-140.  
117 <https://doi.org/10.1016/j.ijbiomac.2010.05.007>

118 Ehrlich, H., Rigby, J. K., Botting, J., Tsurkan, M., Werner, C., Schwille, P., Petrášek, Z.,  
119 Pisera, A., Simon, P., & Sivkov, V. (2013). Discovery of 505-million-year old  
120 chitin in the basal demosponge *Vauxia gracilentia*. Scientific reports, 3(1), 3497.  
121 <https://doi.org/10.1038/srep03497>

122 Erpenbeck, D., Galitz, A., Ekins, M., Cook, S. D. C., van Soest, R. W., Hooper, J. N.,  
123 & Wörheide, G. (2020). Soft sponges with tricky tree: On the phylogeny of  
124 dictyoceratid sponges. Journal of Zoological Systematics and Evolutionary  
125 Research, 58(1), 27-40. <https://doi.org/10.1111/jzs.12351>

126 Erpenbeck, D., Sutcliffe, P., Cook, S. D. C., Dietzel, A., Maldonado, M., van Soest, R.  
127 W., Hooper, J. N., & Wörheide, G. (2012). Horny sponges and their affairs: On  
128 the phylogenetic relationships of keratose sponges. Molecular Phylogenetics  
129 and Evolution, 63(3), 809-816. <https://doi.org/10.1016/j.ympev.2012.02.024>

130 Grant, R. E., 1861, Tabular view of the primary divisions of the Animal Kingdom,  
131 intended to serve as an outline of an elementary course of recent zoology (caino-  
132 zoology); or, the natural history of existing animals, Walton and Maberly.

133 Lavrov, D. V., Wang, X., & Kelly, M. (2008). Reconstructing ordinal relationships in  
134 the Demospongiae using mitochondrial genomic data. Molecular phylogenetics  
135 and evolution, 49(1), 111-124. <https://doi.org/10.1016/j.ympev.2008.05.014>.

- 136 Lendenfeld, R. (1889). A monograph of the horny sponges, Royal Society. Trübner &  
137 Co., London.
- 138 Luo, C., Zhao, F., & Zeng, H. (2020). The first report of a vauxiid sponge from the  
139 Cambrian Chengjiang Biota. *Journal of Paleontology*, 94(1), 28-33.  
140 <https://doi.org/10.1017/jpa.2019.52>.
- 141 Minchin, E. A., & Lankester, E. R. (1900). Chapter III. Sponges. A treatise on zoology.  
142 Part II. The Porifera and Coelenterata, 2, 1-178.
- 143 Morrow, C., & Cárdenas, P. (2015). Proposal for a revised classification of the  
144 Demospongiae (Porifera). *Frontiers in Zoology*, 12, 1-27.  
145 <https://doi.org/10.1186/s12983-015-0099-8>
- 146 Vacelet, J., Erpenbeck, D., Diaz, C., Ehrlich, H., & Fromont, J. (2019). New family and  
147 genus for *Dendrilla*-like sponges with characters of Verongiida. Part I  
148 redescription of *Dendrilla lacunosa* Hentschel 1912, diagnosis of the new family  
149 Ernstillidae and *Ernstilla* ng. *Zoologischer Anzeiger*, 280, 14-20.  
150 <https://doi.org/10.1016/j.jcz.2019.03.001>
- 151 Wiedenmayer, F. (1977). Shallow-water sponges of the western Bahamas: *Experientia*  
152 Supplementum, 1-287.
- 153



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<https://doi.org/10.1111/gbi.12582>

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