

Human Mutation

ANO5 gene analysis in a large cohort of patients with anoctaminopathy: confirmation of male prevalence and high occurrence of the common exon 5 gene mutation

Journal:	Human Mutation
Manuscript ID:	humu-2013-0099.R1
Wiley - Manuscript type:	Research Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Sarkozy, Anna; Institute of Genetic Medicine, Hicks, Debbie; Institute of Genetic Medicine, Laval, Steven; Institute of Genetic Medicine, Barresi, Rita; NCG Diagnostic & Advisory Service for Rare Neuromuscular Diseases, Muscle Immunoanalysis Unit, Dental Hospital, Hilton-Jones, David; Department of Neurology, West Wing, John Radcliffe Hospital, Deschauer, Marcus; University of Halle-Wittenberg, Neurology Harris, Elizabeth; Institute of Genetic Medicine, Rufibach, Laura; Jain Foundation, Bashir, Rumaisa; University of Durham, Biological and Biomedical Sciences Walter, Maggie; Friedrich Baur Institute, Ludwig-Maximilians University, Krause, Sabine; Friedrich Baur Institute, Ludwig-Maximilians University, Van den Bergh, Peter; Université Catholique de Louvain, Brussels, Illa, Isabel; Servei Neurologia, Hospital Sta Creu Sant Pau, Universitat Autonoma de Barcelona; Penisson-Besnier, Isabelle; Département de Neurologie, CHU Dewaele, Liesbeth; Institute of Genetic Medicine, Turnbull, Doug; Newcastle University Centre for Brain Ageing and Vitality, Newcastle University, ; Wellcome Trust Centre for Mitochondrial Research, Institute for Ageing and Health, Newcastle University, Guglieri, Michela; Institute of Genetic Medicine, Schrank, Bertold; Deutsche Klinik für Diagnostik, Schoser, Benedikt; Friedrich Baur Institute, Ludwig-Maximilians University, Seeger, Jurgen; Deutsche Klinik für Diagnostik, Schreiber, Herbert; Neuropoint Patient Academy, (Myology Practice), Glaser, Dieter; Genetikum, Neu-Ulm, Eagle, Michele; Institute of Genetic Medicine, Bailey, Geraldine; Institute of Ge

1 2 3 4 5 6	
7 8	
9 10 11 12 13 14 15 16 17	
18	
20 21 22 23 24 25 26 27 28	
29 30 31 32 33 34 35	
35 36 37 38 39 40	
41 42 43 44 45	
46 47 48 49 50 51	
52 53 54 55 56 57	
58 59 60	

Hospital,
Muntoni, Francesco; MRC Centre for Neuromuscular Diseases, UCL Institut of Neurology and National Hospital for Neurology and Neurosurgery, Quee Square, ; Dubowitz Neuromuscular Centre, UCL Institute of Child Health
and Great Ormond Street Hospital for Children, Hanna, Michael; MRC Centre for Neuromuscular Diseases, UCL Institute o Neurology and National Hospital for Neurology and Neurosurgery, Queen Square, ; Department of Molecular Neuroscience, UCL Institute of
Neurology and National Hospital for Neurology and Neurosurgery, Queen Square,
Roberts, Mark; Hope Hospital, Departments of Neurology and Neuropathology
Bindoff, Laurence; Department of Clinical Medicine, University of Bergen and Department of Neurology, Haukeland University Hospital,
Brierley, Charlotte; Department of Neurology, Addenbrooke's Hospital, Cooper, Robert; University of Manchester Rheumatic Diseases Centre, Salford Royal Foundation Trust,
Cottrell, David; Department of Neurology, Frenchay Hospital, Davis, Nick; Birmingham Muscle and Nerve Centre, Queen Elizabeth Hospital,
Gibson, Andrew; Sheffield Teaching Hospitals NHS Foundation Trust, Gorman, Gráinne; Wellcome Trust Centre for Mitochondrial Research, Institute for Ageing and Health, Newcastle University,
Hammans, Simon; Wessex Neurological Centre, University Hospital of Southampton,
Jackson, Andrew; MRC Human Genetics Unit, MRC IGMM, University of Edinburgh,
Khan, Aijaz; Royal Hallamshire Hospital, Sheffield Teaching Hospitals, Lane, Russell; Charing Cross Hospital, Imperial College, Department of Clinical Neurosciences
McConville, John; Belfast City Hospital, McEntagart, Meriel; St George's University of London, Clinical Genetics Al-Memar, Ali; 34 St George's University Hospital,
Nixon, John; Department of Neurology, Royal Preston Hospital, Panicker, Jay; The Walton Centre For Neurology And Neurosurgery,
Parton, Matt; MRC Centre for Neuromuscular Diseases, UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen
Square, Petty, Richard; Department of Neurology, Southern General Hospital, Price, Christopher; Department of Neuroscience, Musgrove Park Hospital
Taunton, Rakowicz, Wojtek; Imperial College, Neurology Ray, Partha; The Walton Centre For Neurology And Neurosurgery,
Schapira, Anthony; UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen Square,
Swingler, Robert; Department of Neurology, Hospital & Medical School, Turner, Chris; MRC Centre for Neuromuscular Diseases, UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen
Square, ; Department of Molecular Neuroscience, UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen
Square, Wagner, Kathryn; Center for Genetic Muscle Disorders, The Kennedy Krieger Institute and the Departments of Neurology and Neuroscience, The
Johns Hopkins School of Medicine, Maddison, Paul; Nottingham University Hospitals, Shaw, Pamela; <none>,</none>
Straub, Volker; Newcastle University, Institute of Human Genetics; Bushby, Kate
Lochmüller, Hanns; 6Institute of Human Genetics, University of Newcastl upon Tyne, International Centre for Life,

Key Words: ANO5, LGMD2L, gender, common mutation
SCHOLARONE [™]
Manuscripts
John Wiley & Sons, Inc.

ANO5 gene analysis in a large cohort of patients with anoctaminopathy:

confirmation of male prevalence and high occurrence of the common exon 5 gene

mutation

Anna Sarkozy¹, Debbie Hicks¹, Judith Hudson¹, Steve H Laval¹, Rita Barresi², David Hilton-Jones³, Markus Deschauer⁴, Elizabeth Harris¹, Laura Rufibach⁵, Esther Hwang⁵, Rumaisa Bashir⁶, Maggie C Walter⁷, Sabine Krause⁷, Peter van den Bergh⁸, Isabella Illa⁹, Isabelle Pénisson-Besnier¹⁰, Liesbeth De Waele^{1*}, Doug Turnbull^{11,12}, Michela Guglieri¹, Bertold Schrank¹³, Benedikt Schoser⁷, Jürgen Seeger¹³, Herbert Schreiber¹⁴, Dieter Gläser¹⁵, Michelle Eagle¹, Geraldine Bailey¹, Richard Walters¹⁶, Cheryl Longman¹⁷, Fiona Norwood¹⁸, John Winer¹⁹, Francesco Muntoni²⁰⁻²¹, Michael Hanna^{20, 22}, Mark Roberts²³, Laurence A. Bindoff²⁴, Charlotte Brierley²⁵, Robert G Cooper²⁶, David A Cottrell²⁷, Nick P Davis¹⁹, Andrew Gibson²⁸, Gráinne S. Gorman¹², Simon Hammans²⁹, Andrew P. Jackson³⁰, Aijaz Khan³¹, Russell Lane³², John McConville³³, Meriel McEntagart³⁴, Ali Al-Memar³⁴, John Nixon³⁵, Jay Panicker³⁶, Matt Parton²⁰, Richard Petty³⁷, Christopher J Price³⁸, Wojtek Rakowicz³⁹, Partha Ray³⁶, Anthony H Schapira⁴⁰, Robert Swingler⁴¹, Chris Turner^{20,22}, Kathryn Wagner⁴², Paul Maddison⁴³, Pamela J Shaw⁴⁴, Volker Straub¹, Kate Bushby¹ and Hanns Lochmüller¹

1 Institute of Genetic Medicine, International Centre for Life, Newcastle upon Tyne, UK; 2 NCG Diagnostic & Advisory Service for Rare Neuromuscular Diseases, Muscle Immunoanalysis Unit, Dental Hospital, Newcastle upon Tyne, UK; 3Department of Neurology, West Wing, John Radcliffe Hospital, Oxford, UK; 4 Department of Neurology, Martin Luther University Halle-Wittenberg, Halle (Saale), Germany; 5 Jain Foundation Inc., Bellevue, WA, USA; 6 School of Biological and Biomedical Sciences, University of Durham, Durham, UK; 7 Friedrich Baur Institute, Ludwig-Maximilians University, Munich, Germany; 8 Neuromuscular Reference Center, Saint-Luc University Hospitals, Catholic University of Louvain, Brussels, Belgium; 9 Department of Neurology, Neuromuscular Unit Hospital de la Santa Creu i Sant Pau, U.A.B. and CIBERNED Barcelona, Spain; 10 Département de Neurologie, Centre de référence des Maladies Neuromusculaires, Centre Hospitalier Universitaire d'Angers, Angers, France; 11 Newcastle University Centre for Brain Ageing and Vitality, Newcastle University, Newcastle upon Tyne, UK; 12 Wellcome Trust Centre for Mitochondrial Research, Institute for Ageing and Health, Newcastle University, Newcastle upon Tyne, UK; 13 Deutsche Klinik für Diagnostik, Wiesbaden, Germany; 14 Neuropoint Patient Academy, (Myology Practice), Ulm, Germany; 15 Genetikum, Neu-Ulm, Germany; 16 Morriston Hospital, Morriston, Swansea, UK; 17 Ferguson Smith Centre for Clinical Genetics, Yorkhill Hospital, Glasgow, UK; 18 Department of Neurology, Kings College Hospital, London, UK; 19 Birmingham Muscle and Nerve Centre, Queen Elizabeth Hospital, Birmingham, UK; 20 MRC Centre for Neuromuscular Diseases, UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen Square, London, UK; 21 Dubowitz Neuromuscular Centre, UCL Institute of Child Health and Great Ormond Street Hospital, London, UK; 22 Department of Molecular Neuroscience, UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen Square, London, UK; 23 Department of Neurology, Salford Royal NHS Foundation Trust, Salford, United Kingdom; 24 Department of Clinical Medicine, University of Bergen and Department of Neurology, Haukeland University Hospital, Bergen, Norway;

Formatted: English (U.K.)

25 Department of Neurology, Addenbrooke's Hospital, Cambridge, UK; 26 University of Manchester Rheumatic Diseases Centre, Salford Royal Foundation Trust, Salford, UK; 27 Department of Neurology, Frenchay Hospital, Bristol, UK; 28 Sheffield Teaching Hospitals NHS Foundation Trust, Sheffield, UK; 29 Wessex Neurological Centre, University Hospital of Southampton, Southampton, UK; 30 MRC Human Genetics Unit, MRC IGMM, University of Edinburgh, UK; 31 Royal Hallamshire Hospital, Sheffield Teaching Hospitals, Sheffield, UK; 32 Division of Clinical Neurosciences, Imperial College London, UK; 33 Belfast City Hospital, Belfast, UK; 34 St George's University Hospital, London, UK; 35 Department of Neurology, Royal Preston Hospital, Preston, UK; 36 The Walton Centre For Neurology And Neurosurgery, Liverpool, UK; 37 Department of Neurology, Southern General Hospital, Glasgow, UK; 38 Department of Neuroscience, Musgrove Park Hospital, Taunton, Somerset, UK; 39 Department of Neurology, Charing Cross Hospital, Imperial College Healthcare NHS Trust, London, UK; 40 UCL Institute of Neurology and National Hospital for Neurology and Neurosurgery, Queen Square, London, UK; 41 Department of Neurology, Hospital & Medical School, Dundee, UK; 42 Center for Genetic Muscle Disorders, The Kennedy Krieger Institute and the Departments of Neurology and Neuroscience, The Johns Hopkins School of Medicine N., stitute 1. diation: Dep. , Belgium Baltimore, USA; 43 Nottingham University Hospitals, Nottingham, UK; 44 Academic Units of Neurology, Department of Neuroscience, Sheffield Institute for Translational Neuroscience (SITraN), University of Sheffield, Sheffield, UK; * current affiliation: Department of Paediatrics, section Paediatric Neurology, University Hospitals Leuven, Belgium

Corresponding author:

Prof Hanns Lochmüller

Institute of Genetic Medicine

International Centre for Life

Central Parkway

Newcastle upon Tyne

NE1 3BZ

Fax: +44 (0) 191 241 8770

Article Type: Research Article for Human Mutation

Running title: Clinical and molecular analysis of anoctaminopathies

Abstract

Limb girdle muscular dystrophy type 2L or anoctaminopathy is a condition mainly characterized by adult onset proximal lower limb muscular weakness and raised CK values, due to recessive ANO5 gene mutations. An exon 5 founder mutation (c.191dupA) has been identified in most of the British and German LGMD2L patients so far reported. We aimed to further investigate the prevalence and spectrum of ANO5 gene mutations and related clinical phenotypes, by screening 205 undiagnosed patients referred to our molecular service with a clinical suspicion of anoctaminopathy. A total of 42 unrelated patients had two ANO5 mutations (21%), while 14 carried a single change. We identified 34 pathogenic changes, 16-135 of which are novel. The c.191dupA mutation represents 61% of mutated alleles and appears to be less prevalent in non-Northern European populations. Retrospective clinical analysis corroborates the prevalently proximal lower limb phenotype, the male predominance and absence of major cardiac or respiratory involvement. Identification of cases with isolated hyperCKaemia and very late symptomatic male and female subjects further confirms the broadensextension of the phenotypic spectrum of the disease. Anoctaminopathy appears to be one of the most common adult muscular dystrophies in Northern Europe, with a prevalence of about 20-25% in unselected undiagnosed cases.

Key words:

ANO5, LGMD2L, gender, common mutation

Introduction

Limb girdle muscular dystrophy type 2L (LGMD2L) is a condition characterized by proximal weakness affecting primarily the pelvic girdle and leg muscles, with less prominent distal leg weakness and high CK values (Hicks et al, 2011). LGMD2L, together with a more distal variant also known as non-dysferlin Miyoshi muscular dystrophy type 3 (MMD3) as well as some cases of isolated hyperCKaemia and pseudometabolic myopathy, have been shown to be associated with recessive mutations in the ANO5 gene (OMIM accession number 608662; Bolduc et al, 2010; Hicks et al, 2011; Milone et al, 2012; Deschauer et al, 2012; Schessl et al, 2012; Penttilä et al, 2012; Bouquet et al, 2012; Magri et al, 2012; Pénisson-Besnier et al, 2012; Wahbi et al, 2012; Witting et al, 2012; Little et al, 20132; Neusch et al, 2012). The prevalence of ANO5 gene related conditions, or socalled anoctaminopathies, is not yet fully established, but preliminary data from Northern England indicate that LGMD2L is the 3rd most common type of LGMD with a minimum prevalence of 0.27/100 000 (Hicks et al, 2011). To date, 35-67 variants (35 pathogenic; www.lovd.nl/ANO5) mutations have been detected all over the ANO5 gene, with one a common mutation (c.191dupA) in exon 5, likely the result of a founder effect of Northern European origin (Hicks et al, 2011), and another in exon 20 (c.2272C>T) more frequent in the Finnish population (Penttilä et al, 2012). Early clinical and MRI studies indicated wide clinical heterogeneity and a gender difference in expression, with anoctaminopathies appearing to be less frequent and less severe in females (Sarkozy et al, 2012).

In the present study, we investigated the prevalence and spectrum of *ANO5* gene mutations in the so far largest and clinically most heterogeneous cohort of

Formatted: Font: Times New Roman, Not Bold, Not Highlight

Human Mutation

patients with anoctaminopathy, in order to better describe the phenotype, gender predominance and allelic variability of this emerging disease.

Materials and Methods

The study was performed on a total of 205 unrelated individuals of both sexes, referred for *ANO5* gene analysis to the Newcastle upon Tyne Limb-Girdle Referral Centre in view of their phenotypes compatible with anoctaminopathy, ranging from isolated hyperCKaemia, myalgia and raised CK, to MMD3 and LGMD2L. Patients were referred from different clinical centers, but primarily from the UK, Germany, France, Spain, the USA and Belgium. The cohort previously reported by Hicks et al. was not included in this study (Hicks et al, 2011). Clinical details for patients with *ANO5* mutation were collected retrospectively. The clinical suspicion of anoctaminopathy was made following clinical examination, muscle assessment and clinical investigations including CK analysis, EMG, muscle MRI and/or muscle biopsy. Possible alternative diagnoses, such as LGMD2B, LGMD2I and LGMD2A, were excluded as appropriate. DNAs of affected and unaffected relatives were collected and segregation analysis performed where available.

DNA extraction and direct sequencing of the *ANO5* gene (Gene Bank accession number NM 213599.2) was performed as described elsewhere (Hicks et al,

2011). Sequence variants were coded reflecting cDNA numbering with +1 corresponding to the A of the ATG translation initiation codon in the reference sequence, according to guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. Novelty of each sequence variant was checked in the <u>ANO5 locus specific</u> database (www.lovd.nl/ANO5). To investigate whether novel missense, synonymous or intronic variations not located in splice sites could possibly represent pathogenic Formatted: Font: 12 pt, Not Bold, Font color: Auto, English (U.S.)

Formatted: Font: 12 pt, Not Bold, Font color: Auto, English (U.S.)

Formatted: Font: Italic

Formatted: Font: 12 pt, Not Bold, English (U.S.), Not Highlight

sequence variants, we performed in silico analysis using the Alamut mutation analysis software (Interactive Biosoftware, v2.1).

Genetic analysis

ANO5 gene sequence variants were identified in 90/205 unrelated individuals (46%) and 5 affected relatives (Table 1, Suppl. Table 1). Fifty-one patients were found to carry 2 or more ANO5 variants in homozygosity or compound heterozygosity, while further 39 patients showed one single ANO5 change (Tables 1 and 2). A total of 42 ANO5 gene variants were identified, 32-27 of which novel (Tables 1 and 2). Twenty variants were missense (47%), 4 frameshift (9%), 6 splice site (14%), 2 stop (5%), 1 synonymous (2%) and 10 intronic (24%) (Table 2). The pathogenicity of 16-135 novel variants is supported by their predicted effect on protein product, by their recurrence in multiple unrelated patients, by segregation in affected family members, absence in SNPs databases and in silico analysis. Four missense changes, two of which were previously reported in literature as pathogenic (c.155A>G and c.2387C>T)), were considered benign variants based on *in silico* analysis and/or their frequency in control population (Table 2) (Wahbi et al, 2012). It was not possible to further comment on the pathogenicity of a missense (c.968C>G) and a synonymous change (c.2256G>A) as well as 10 intronic variants in absence of segregation studies or further cDNA investigations.

Forty-two-unrelated patients (N.1-N.42) carried 2 likely pathogenic ANO5 gene changes, 24-19 in homozygosity and 20-283 in compound heterozygosity (Table 1). Patients N.21B, N.22B, N.28B, N.40B-C carried the pathogenic variants found in their affected relative (i.e. N.21A, N.22A, N.28A and N.40A). In 14 patients (N.43-N.56) only one single pathogenic variant was identified. cDNA analysis or

Results

genomic dosage analysis was not performed to rule out a possible 2^{nd} deep intronic change or deletion/duplication of the part/entire *ANO5* gene. Seven novel pathogenic variants were recurrent (Tables 1 and 2). The common c.191dupA exon 5 mutation was found in homozygosity in 16 patients, in compound heterozygosity with another identified mutation in 19, and in heterozygosity without a 2^{nd} detectable mutation in further 5 individuals. Geographic origin of patients, indicated in Table 1, confirms that the common variant is present also in populations other than the British and German populations. Additionally, two already reported (c.692G>T, c.1927dupA) and 2 novel (c.1898+1G>A, c.2417A>G) mutations were also identified in multiple patients, and further 3 novel changes were found in homozygosity in 3 individuals (Tables 1 and 2). Parental DNA screening or *ANO5* genomic dosage analysis was not performed yet for the homozygous patients, and hemizygosity of parts or the entire *ANO5* locus cannot be ruled out.

Phenotypic analysis

Among patients with 2 likely pathogenic changes (total 47 patients, including 42 unrelated subjects and 5 affected relatives), 39 were male (83%) and 8 were female (17%). Clinical information was available for 4144_patients and relatives with 2 pathogenic changes 44 patients (34 males, and 7 females) (table 3). Thirty four patients (83%) were male and 7 were female (17%). Age at onset ranged between teens and late 70s, with an average age at onset of 35 years. At time of genetic diagnosis, 3/7 females and 3/374 males were completely asymptomatic or very mildly symptomatic. Average CK values were 4000 IU/l, ranging from around 200 IU/l up to about 30.000 IU/L. Detailed information about last muscle assessment At time of last assessment was available for 38/47 individuals., a A predominantly proximal involvement was observed in 22/38 (57%) patients, while 4/38 (10%) and 3/38 (8%)

patients showed a more distal or proximal plus distal involvement, respectively. No or very mild clinical symptoms (myalgia and/or raised CK) were reported in 9 patients with 2 *ANO5* mutations (25%).

Symptoms at onset and overall clinical presentation in males were similar to what has been previously described for patients with *ANO5* gene mutations (Table 3; Hicks et al, 2011). Although age at onset was similar on average (34 years), in this cohort we observed a wider range, with patients showing first symptoms in their teens to patients only symptomatic in late adulthood, with one patient (Pt. N.38) showing first symptoms of proximal lower limb weakness and wasting at the age of 77 years only. Moreover, the phenotype was extremely mild in 3 individuals, only showing raised CK values (Pt. N.15 and N.36) or some mild proximal leg weakness after a disease course of 32 years (Pt. N40B). Patient N.21B was diagnosed as part of family screening and showed no symptoms of the disease. CK values were not available at the time of study.

Five female individuals (N.03, N.21A, N.27, N.28B, 40C), with an age at last assessment ranging from 14 to 68 years, showed raised CK values only or asymmetric calf atrophy, while female patients N.02 and N.14, aged 68 and 42 years respectively, showed a more pronounced LGMD type weakness. Female patient N.08 (not included in table 3 in view of lack of -detailed clinical information) was diagnosed as part of a family screening of a previously reported LGMD2L patient. At the age of 44 years she does not report any clinical symptoms, but no further clinical information was available at time of the study.

A total of 7 unrelated patients were confirmed with anoctaminopathy, but did not carry the common c.191dupA mutation on either allele. Age at onset was 44 years on average. Four patients showed a milder phenotype, with a very late onset (77

years), normal power or only mild proximal weakness at last assessment. Muscle wasting was present in half of them, while upper limb function was normal in all.

Clinical information were was available for 11/14 <u>unrelated</u> patients with one single pathogenic variant (data not shown). Five individuals showed a phenotype compatible with a diagnosis of LGMD2L, while 3 subjects only showed raised CK values with or without calf hypertrophy. In one individual (Pt N.50) this variant was also found in further two family members also affected by isolated hyperCKaemia.

Clinical analysis of the 7 patients with putatively non pathogenic exonic mutations (Pt. N57 63) showed phenotypes not typical for anoctaminopathy, with lower CK values, more severe upper limb involvement or early age at onset (data not shown). Phenotypic analysis of patients with intronic changes only (Pt. N 64 90) was not performed.

Discussion

In this study we report on the clinical analysis and molecular screening of the *ANO5* gene in the largest cohort of anoctaminopathy patients identified so far. Our results confirm the high frequency of the common c.191dupA mutation which represents 51/84 (61%) of mutated alleles in the cohort of patients with two mutations (Tables 1 and 2). Occurrence of this Northern European founder mutation in patients of different geographic origin indicates that it has likely spread worldwide, although at lower frequencies compared to the British and German populations. This is corroborated by the observation that the mutation has not been observed in homozygosity in other populations, with the exception of two unrelated French patients (Wahbi et al, 2012). However, our result could have been biased by the predominance of British and German patients in our cohort.

We identified 34-<u>27</u> novel variants in the *ANO5* gene, <u>16-135</u> of which are likely pathogenic (Tables 1 and 2). Mutations are spread across the gene, indicating the absence of additional mutation hot spots, although some mutations appear to be recurrent (Tables 1 and 2). The frequency of the second most common variant (c.692G>T) suggests that it could represent an additional common change in our population (Table 1). Interestingly, the common Finnish mutation appears to be rare in our cohort, with a single occurrence in a German patient in compound heterozygosity with the c.191dupA mutation (Pt N.35). Mutations leading to loss of protein function (frameshift, splice and stop mutations) were the most common changes in the "2-mutation" group of patients (66/84 alleles, 79%) although the relative frequency of missense changes (18/84 alleles, 21%) could imply further pathogenetic mechanisms.

We have identified several exonic changes that appear to be benign variants or to have uncertain pathogenicity. Among these, the non pathogenic variant c.155A>G (p.Asn52Ser) in exon 4 (dbSNP rs143777403), found with allelic frequency of 0.0001-0.001 in North American and African American control populations, was identified in 5 patients from our cohort. The c.2387C>T (p.Ser796Leu) in exon 22, also a benign variant by *in silico* analysis, was found in another individual. Interestingly, these two missense variants were described as pathogenic by Wahbi and coauthors (2012) in 3 unrelated subjects, one of which was compound heterozygous for this and the c.155A>G variant. These findings point towards caution when novel missense changes are found (especially if in compound heterozygosity), and no further data corroborating the pathogenicity of the change is available. We identified 3 patients who were homozygous for a novel variant and 14 patients with 1 heterozygous change only. One of the limitations of this study is the lack of cDNA

Human Mutation

studies to exclude 2nd deep intronic changes or gene dosage analysis, but deletion/duplication analysis is currently in progress in some of these individuals.

We have previously described the clinical phenotype of anoctaminopathies as an adult onset disease, characterized by slowly progressive muscle weakness mainly affecting the pelvic girdle and the lower limbs (Hicks et al, 2011). Clinical analysis of the present cohort of patients extends our initial findings to a large group of patients not restricted to the common founder mutation (Table 3). Being our center the only currently offering ANO5 screening in the UK, the prevalence of proximal disease is unlikely to be caused by the referral bias previously suggested (Hicks et al, 2011). No major significant cardiac or respiratory disease was observed in this cohort, suggesting that these are not major features of the condition. We provide further evidence for male predominance in anoctaminopathies, females being overall less frequently and less severely affected than males (Hicks et al, 2011, Penttilä et al, 2012). Phenotypic analysis of the here reported affected females reveals a milder clinical picture, with 5 patients showing only raised CK values or calf hypertrophy. The oldest female individual, homozygous for the common exon 5 mutation, showed a late disease onset (64 years), and at the age of 68 years she was still ambulant and she was able to stand on tiptoes and heels with minor difficulties. These findings suggest that gender differences could be in part due to the milder phenotype of affected females that is therefore less likely to be ascertained. In fact, two asymptomatic females were identified after a molecular diagnosis was reached in their brothers. Recruitment bias could also be in part be responsible for gender difference, as patients seen in specialized centers usually show a more severe phenotype, and therefore females with milder phenotypes might not come as frequently to our attention. In this study, we also identified 4 male subjects showing a very mild

phenotype, ranging from isolated hyperCKaemia to mild proximal leg weakness with onset in late adulthood (Table 3). In particular, one subject only showed symptoms in his late 70s, confirming that indeed age at onset and severity of anoctaminopathies can be extremely variable also in males (Hicks et al, 2010; Schessl et al, 2012; Penttilä et al, 2012).

No genotype-phenotype correlations were noted in anoctaminopathy so far. Nevertheless, a clinical comparison between patients carrying the common exon 5 mutation and those carrying different changes revealed some interesting findings (Table 1 and 3). Pattern of muscle involvement and female/male ratio appear similar, although the phenotype is somehow milder, with onset at later age in patients carrying mutations other than the c.191dupA. Unfortunately the relatively low number of patients with different ANO5 changes does not allow any statistically significant conclusions. Conversely, phenotypic analysis of heterozygous patients did not evidence major differences compared to patients with two mutations, although 3 heterozygous patients presented raised CK values and mild calf hypertrophy only. While we cannot exclude a second non-yet identified change in these subjects, family analysis of one individual (Pt. N.50) showed that raised CK values segregated with the heterozygous mutation in 2 other family members (data not shown). An association between isolated hyperCKaemia and ANO5 heterozygous carrier status was suggested by Milone and coworkers, who identified an otherwise asymptomatic carrier of the c.191dupA mutation (Milone et al, 2011). Likewise, isolated hyperCKaemia or very mild phenotypes were also found in a few male and female patients carrying two pathogenic ANO5 mutations (Hicks et al, 2011; Schessl et al, 2012; Penttilä et al, 2012, Wahbi et al, 2012). Systematic analysis of ANO5 gene in

John Wiley & Sons, Inc.

 wider cohort of subjects with isolated hyperCKaemia, as well as CK measurements in otherwise healthy heterozygous family members could help to clarify these findings.

Our results show that *ANO5* gene mutations are responsible for about ¹/₄ of cases of undiagnosed muscular dystrophy with adult onset and raised CK, screened at our service over the last two years (present series and Hicks et al, 2011). This value is similar to what reported in the Finnish population (Penttilä et al, 2012) and indicates anoctaminopathy as one of the most common form of LGMD in our population, possible reflection of the founder effects observed in Northern Europe (Hicks et al, 2011). Indeed, observed incidences in the UK and Finland are higher than in Italy (Magri et al, 2012) where only about 2% of undiagnosed LGMD patients were shown to carry *ANO5* mutations. Based on its relative frequency, screening of the *ANO5* gene now represents an early step in the diagnostic work-up of these patients. However, in view of the increasing genetic heterogeneity and prevalence of families without the common mutation (14%), analysis should not be limited to founder mutations only (Penttilä et al, 2012). More systematic studies on isolated hyperCKaemia patients will also help in giving better prevalence data for anoctaminopathies in general.

In conclusion, we confirmed that *ANO5* gene mutations are responsible for about ¹/₄ of cases of undiagnosed muscular dystrophy in our population, being the exon 5 change the most prevalent. We expanded the allelic heterogeneity of the disease and recognized a broader clinical spectrum of the disease, ranging from isolated hyperCKaemia to full blown LGMD2L, with clear male predominance both in terms of overall prevalence and severity of the disease.

Acknowledgments

The authors would like to thank the patients who took part in this study. Diagnostic facilities at the Newcastle Muscle Centre are supported by the National Specialised Commissioning Team (NSCT) for rare neuromuscular disorders. The Institute of Genetic Medicine in Newcastle is part of the MRC Centre for Neuromuscular Diseases and the TREAT-NMD Alliance (www.treat-nmd.eu). > conflicts of intered. There are no conflicts of interest to declare.

John Wiley & Sons, Inc.

2
3
3 4 5 6 7
5
6
0
1
8
9
10
8 9 10 11 12 13 14 15 16 17 18
10
12
13
14
15
16
17
18
10
19 20 21 22 23 24 25 26 27 28 30 31 32 33 34 35 36 37 38 39 40
20
21
22
23
21
24
20
26
27
28
29
30
24
31
32
33
34
35
36
27
31
38
39
40
41
42
43
40
44
45
46
47
48
49
50
50
51
52
53
54
50 51 52 53 54 55 56 57 58 59
56
50
5/ 52
58
59

60

References

- Bolduc V, Marlow G, Boycott KM, Saleki K, Inoue H, Kroon J, Itakura M, Robitaille
 Y, Parent L, Baas F, Mizuta K, Kamata N, et al. 2010. Recessive mutations in
 the putative calcium-activated chloride channel Anoctamin 5 cause proximal
 LGMD2L and distal MMD3 muscular dystrophies. Am J Hum Genet 86:213-21.
- Bouquet F, Cossée M, Béhin A, Deburgrave N, Romero N, Leturcq F, Eymard B. 2012. Miyoshi-like distal myopathy with mutations in anoctamin 5 gene. Rev Neurol (Paris) 168:135-41.
- Deschauer M, Joshi PR, Gläser D, Hanisch F, Stoltenburg G, Zierz S. 2011. Muscular dystrophy due to mutations in anoctamin 5 : Clinical and molecular genetic findings. Nervenarzt 82:1596-603.
- Hicks D, Sarkozy A, Muelas N, Koehler K, Huebner A, Hudson G, Chinnery PF,
 Barresi R, Eagle M, Polvikoski T, Bailey G, Miller J, et al. 2011. A founder
 mutation in Anoctamin 5 is a major cause of limb-girdle muscular dystrophy.
 Brain 134:171-82.
- Little AA, McKeever PE, Gruis KL. 201<u>3</u>2. Novel mutations in the anoctamin 5 gene (ANO5) associated with limb-girdle muscular dystrophy 2L. Muscle Nerve

47:287-91[Epub ahead of print]

- Magri F, Bo RD, D'Angelo MG, Sciacco M, Gandossini S, Govoni A, Napoli L, Ciscato P, Fortunato F, Brighina E, Bonato S, Bordoni A, et al. 2012. Frequency and characterisation of anoctamin 5 mutations in a cohort of Italian limb-girdle muscular dystrophy patients. Neuromuscul Disord 22:934-43
- Milone M, Liewluck T, Winder TL, Pianosi PT. 2012. Amyloidosis and exercise intolerance in ANO5 muscular dystrophy. Neuromuscul Disord 22:13-5

Penttilä S, Palmio J, Suominen T, Raheem O, Evilä A, Muelas Gomez N, Tasca G,
Waddell LB, Clarke NF, Barboi A, Hackman P, Udd B. 2012. Eight new
mutations and the expanding phenotype variability in muscular dystrophy caused
by ANO5. Neurology 78:897-903
Pénisson-Besnier I, Saint-André JP, Hicks D, Sarkozy A, Croué A, Hudson J,
Lochmüller H, Dubas F. 2012. Myopathy caused by anoctamin 5 mutations and
necrotizing vasculitis. J Neurol 259:1988-90.

Sarkozy A, Deschauer M, Carlier RY, Schrank B, Seeger J, Walter MC, Schoser B, Reilich P, Leturq F, Radunovic A, Behin A, Laforet P, et al. 2012. Muscle MRI findings in limb girdle muscular dystrophy type 2L. Neuromuscul Disord 22 Suppl 2:S122-9.

- Schessl J, Kress W, Schoser B. Novel ANO5 mutations causing hyper-CK-emia, limb girdle muscular weakness and Miyoshi type of muscular dystrophy. 2012. Muscle Nerve 45:740-2.
- Wahbi K, Béhin A, Bécane HM, Leturcq F, Cossée M, Laforêt P, Stojkovic T, CarlierP, Toussaint M, Gaxotte V, Cluzel P, Eymard B et al. 2012. Dilatedcardiomyopathy in patients with mutations in anoctamin 5. Int J Cardiol [Epubahead of print]
- Witting N, Duno M, Born AP, Vissing J. 2012. LGMD2L with bone affection: overlapping phenotype of dominant and recessive ANO5-induced disease. Muscle Nerve 46:829-30.

John Wiley & Sons, Inc.

 Table N1. Patients with ANO5 gene mutations

Patient	allele 1	allele 2	Geographic origin	_		
Patients with 2 n	athogenic variants					
N.01-N.16	c.191dupA	c.191dupA*	Great Britain (12); Germany (4)			Formatted: Highlight
N.17-N22B	c.191dupA	c.692G>T	Great Britain (7)			Formatted: Highlight
N.23	c.191dupA	c.762+1G>A	Great Britain			
N.24	c.191dupA	c.1391C>A	USA	-		(-
N.25	c.191dupA	c.1407+5G>T	Belgium	-	/	Formatted
N.26-N.27	c.191dupA	c.1627dupA	Spain, n.a.			Formatted
N.28A-N.28B	c.191dupA	c.1643C>T	Germany (2)			Formatted
N.29	c.191dupA	c.1733T>C	Great Britain			Formatted
N.30-N.32	c.191dupA	c.1898+1G>A	Great Britain, Poland and USA			Formatted
N.33	c.191dupA	c.2395C>T	Great Britain			Formatted
N.34	c.191dupA	c.2417A>G	USA	-		Formatted
N.35	c.191dupA	c.2272C>T	Germany	-		Formatted
N.36	c.401A>G	c.1898+1G>A	Great Britain		,	Formatted
N.37	c.1639C>T	c.1639C>T	n.a.			Formatted
N.38	c.2030-1G>T	c.2030-1G>T	Great Britain			Formatted
N.39	c.2236-13_2236- 4delinsATTCTTCTGGC	c.2236-13_2236- 4delinsATTCTTCTGGC	Great Britain			Formatted
N.40A-N.40C	c.242A>G	c.1097A>G	Germany (3)			Formatted
N.41	c.400C>T	c.2235+1G>A	France		/	Formatted
N.42	c.989dupTex10	c.2018A>G	Great Britain			Formatted
						Formatted
Patients with 1 p	athogenic variant					Formatted
N.43-N.44	c.191dupA	c.155A>G	Great Britain; n.a.			Formatted
N.45-N.47	c.191dupA		Germany (2); Great Britain			Formatted
N.48-N.50	c.692G>T		Great Britain (3)			Formatted
N.51	c.762+1G>A	c.155A>G	USA		N	Formatted
N.52	c.1925G>T	c.1181-21T>A	Great Britain			Formatted
N.53	c.1640G>A	_	Great Britain	1		Formatted

translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. Legend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity.					
N 55 C 2321C>G - Great Britain N 56 C 2393A - Pakistan Patients with coole and thronic changes with ancestain pathogenicity - Pakistan N 57: N 53 C 135A-0 - Great Britain (2) N 50 C 2393A - - Pakistan N 64 C 2382C-1 - Great Britain - N 64 C 2382C-1 - Great Britain - - N 64 C 2382C-1 - Great Britain - </th <th></th> <th></th> <th></th> <th></th> <th></th>					
N55 c.2321C>G - Great Britain Patients with econic and intronic changes with ancestain pathogenicity - Pakistan N57 N53 C.155A>G - Restantion (2) N57 N53 C.155A>G - Great Britain (2) N53 N64 e.2656C>A - Restantian N64 e.2327C>T - Great Britain N63 e.36484pt Correst Britain - N64 e.2327C>T - Great Britain N63 e.36498A>C - Great Britain N64 e.364940pT - Great Britain N66 e.1119135C>A e.118148E>A Germany N64 e.364940pT - Africa and Great Britain N68 e.1119135C>A e.118148E>A Germany N70-728 e.364-840pT - Africa and Great Britain N70-728 e.364-840pT - Africa and Great Britain N17-728 e.364-840pT - Interview of the ATC Natastation infinition codon in the refreence sequencongrin op journal guiddines (tww hysocorgonutom					
N55 c.2321C>G - Great Britain Patients with econic and intronic changes with ancestain pathogenicity - Pakistan N57 N53 C.155A>G - Restantion (2) N57 N53 C.155A>G - Great Britain (2) N53 N64 e.2656C>A - Restantian N64 e.2327C>T - Great Britain N63 e.36484pt Correst Britain - N64 e.2327C>T - Great Britain N63 e.36498A>C - Great Britain N64 e.364940pT - Great Britain N66 e.1119135C>A e.118148E>A Germany N64 e.364940pT - Africa and Great Britain N68 e.1119135C>A e.118148E>A Germany N70-728 e.364-840pT - Africa and Great Britain N70-728 e.364-840pT - Africa and Great Britain N17-728 e.364-840pT - Interview of the ATC Natastation infinition codon in the refreence sequencongrin op journal guiddines (tww hysocorgonutom					
N.56 c.2593A/C - Pakistan Patients with econic and intronic changes with uncertain pathogenicity Great-Britain (2) N.59 c.756A/C - Great-Britain (2) N.60 c.966C/C - Bakistan N.61 c.2256C/A - Great-Britain N.64 c.2256C/A - Great-Britain N.64 c.2256C/A - Great-Britain N.64 c.2256C/A - Great-Britain N.64 c.2360/A/C - Great-Britain N.64 c.2360/A/C - Great-Britain N.64 c.2360/A/C - Great-Britain N.66 c.1119-350/A c.11181487/A Gremany N.64 0.6129.59dupT - n.ar. N.07.78 c.364-8dupT - n.dritain Note: M.035 gene. Gene Back accession number NM, 213599.2 Nucleotide numbering reflecters DNA numbering the torresponding to the A of the ATG r translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutmome). The initiation codon is codon 1. L Legend: "patient N 05 and N.13 also with variant c.364-67T	N.54	c.2417A>G	-	Great Britain	
Particular value and intronic changes with uncertain pathogenicity N57N3 Citation N50 0.4042-0 N40 0.20550-3 N40 0.2137-51 N40 0.2137-51 N40 0.2137-51 N40 0.2137-51 N40 0.2137-52 N404 0.2137-52 N404 0.2137-52 N404 0.2137-52 N404 0.2137-52 N404 0.1191-556-3 0.217 0.2164-54 0.217 0.2164-54 0.217 0.2164-54 0.217 0.2164-54 0.217 0.21592. Nuclearing to interval 0.21592. Nuclearing to interval 0.21592. Nuclearing to interval 0.21592. Nuclearing to interval 0.21592. <t< td=""><td>N.55</td><td>c.2521C>G</td><td>-</td><td>Great Britain</td><td></td></t<>	N.55	c.2521C>G	-	Great Britain	
N57-N-58 e-155A-G - Great Britain (2) N:60 e-0848C-6 - Real Britain N:61 e-2256C-A - Real Britain N:62 e-2387C-7 - Great Britain N:63 e-208A-C- - Germany N:64 e-194350-A - Germany N:64 e-194350-A - Germany N:64 e-1109-350-A e-110141487-A Germany N:66 e-1019-350-3940pT - ne: N:70-78 e-364-840pT - ne: N:70-78 e-364-840pT - ne: N:70-78 e-364-840pT - ne: N:70-78 e-364-840pT - ne: scar. AVO5 genc. Genc Bank accession number NM 213599-2. Nucleotide numbering reflects CDNA numbering with +1 corresponding to the A of the ATG - egend: *patient N 05 and N 13 also with variant c 364-67T>C with uncertain pathogenicity. egend: *patient N 05 and N 13 also with variant c 364-67T>C with uncertain pathogenicity. - -	N.56	c.2593A>C	-	Pakistan	-
N.57.N.58 e.155A-G - Great Britain (2) N.69 e.746C-G - Great Britain N.60 e.966C-G - Palistan N.61 e.236G>A - Great Britain N.62 e.2387C>T - Great Britain N.63 e.2698A-C - Germany N.64 e.236G>A - Germany N.64 e.2387C>T - Germany N.64 e.2387C>T - Germany N.64 e.24484upT Africe and Great Britain - N.68 e.1119+35G>A e.1414 18T>A Germany N.69 e.130-9594upT - n= N.70-78 e.364 84upT - n= N.70-78 e.364 84upT - Africe and Great Britain Note: ANO5 gene. Gene Bank accession number NM 213599-2. Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG - ranslation initiation codon is codon 1. Legend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity.					
N-59 e-746C>G - Great Britain N-60 e-266C>A - Relistant N-61 e-2256C>A - Great Britain N-62 e-2387C>T - Great Britain N-63 e-2668A>C - Great Britain N-64 e-2287C>T - Great Britain N-64 e-2387C>T - Great Britain N-64 e-2468AvC - Great Britain N-66 e-1119-25G>A e-1181-8T>A Germany N-66 e-1190-25G>A e-1181-8T>A Germany N-69 e-139-59dupT - Africe and Great Britain N:02: M-02 gene. Great Bark accession number NM 213599.2. Nucleotide numbering reflects cDNA numbering reflects down have accession anumber NM 213599.2. Nucleotide numbering reflects cDNA numbering reflects down have accession anumber NL 213599.2. Nucleotide numbering reflects down have accession anumber NL 213599.2. Nucleotide numbering reflects down have accession anumber NL 213599.2. Nucleotide numbering reflects down have accession anumber NL 213599.2. Nucleotide numbering reflects down have accession anumber NL 21359.2. Nucleotide numbering reflects down have accession anumber NL 21359.2. Nucleotide numbering reflects down have accession anumber NL 21359.2. Nucleotide number accession anumber NL 21359.2. Nucleotide number acceston anumber NL 21359.2. Nu				Creat Britain (2)	
N40 e-9882~6 - Pakisum N-64 e-22966~A - Great Ditain N-62 e-2387C~T - Great Ditain N-63 e-2698A~C. - Great Ditain N-64 e-2387C~T - Great Ditain N-63 e-2698A~C. - Great Ditain N-64 e-364-84upT e-364-84upT Africe and Creat Britain N-66 e-119-396G~A e-1181-48T>A Germany N-69 e-129-594upT - n-at N-70-78 e-364-84upT - M-Africe and Creat Britain Note: ANO5 gene. Gene Bank accession number NM 213599.2. Nucleotide numbering reflects eDNA numbering with +1 corresponding to the A of the ATG - ranslation initiation codon in the reference sequence, according to journal guidelines (www.lgvs.org/mutnumen). The initiation codon is codon 1. - .egend.*patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. - 2 - - - .egend.*patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. -					
N-61 e-2256G>A - Great Britain N-62 e-2367C>T - Gremany N-63 e-2668A>C - Gremany N-64 e-344-8dupT Africe and Great Britain N-64 e-1119+35G>A e-1181-48T>A Germany N-69 e-139-59dupT - n.a. N-70-78 e-364-8dupT - Intra-not draw for the ATG Note: AV03 gene. Gene Bank accession number NM 213599.2. Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG - Itarialition initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/nutnomen). The initiation codon is codon 1. - Legend. * patient N 05 and N 13 also with variant c 364-67T-C with uncertain pathogenicity. - -					
N-62 e-2387C>T - Great Britain N-63 e-2698A-C: - Germany N-64-67 e-36448dupT Africe and Great Britain N-68 e-1110+25G>A e-1181-48T>A Germany N-69 e-130-59dupT - n:n: N-69 e-130-59dupT - Africe and Great Britain N-69 e-130-59dupT - Africe and Great Britain Note: ANO5 gene. Gene Bank accession number NM 213599.2. Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG - Instalation infinition codon in the reference sequence, according to journal guidelines (www hervs org/muttomen). The initiation codon is codon 1. Legend.*patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. 2 Africe and N.13 also with variant c.364-67T>C with uncertain pathogenicity. -					
N:63 e:2698A>C: - Germany N:64 e:364-8&upT e:364-8&upT Africe and Great Britain N:68 e:1119+35G>A e:1181-48T>A Germany N:69 e:304-9&upT - n:n N:70-78 e:364-8&upT - n:n Note: AVOS gene, Gene Bank accession number NM 213599.2. Nucleotide numbering reflects EDNA numbering with +1 corresponding to the A of the ATG - Iterastation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/minnomen). The initiation codon is codon 1. - Legend: *patient N:05 and N:13 also with variant c:364-67T>C with uncertain pathogenicity. - 2 - - - 8 - - - 9 - - - 9 -					
N.64-67 e.364-8dupT Africa and Great Britain N.68 e.119+35G>A e.1181-48T>A Germany N.69 e.139-S9dupT - n: N.70-78 e.364-8dupT - - Not: AN70-78 e.364-8dupT - - Not: AN70-78 e.364-8dupT - - Not: AN70-78 e.364-8dupT - - - Not: AN05 gene, Gene Bank accession number NM 213599.2, Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG - translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. - Legend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. - - 2 - - - - 8 - - - - 9 - - - - 1 - - - - 1 - - - - 1 - - - - 1					
N.66 e.1119+35G-A e.1181-48T-A Germany N.60 e.139-59dupT - n:tr N.70-78 e.364-8dupT - Africe and Great Britain Note: ANO3 gene. Gene Bank accession number NM 213599.2. Nucleotide numbering crelects DNA numbering with +1 corresponding to the A of the ATG * Note: ANO3 gene. Gene Bank accession number NM 213599.2. Nucleotide numbering crelects DNA numbering with +1 corresponding to the A of the ATG * Instation infinition codon in the reference sequence, according to journal guidelines (www.hgvs.org/muthomen). The initiation codon is codon 1. * Legend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. *			<u>c.364-8dunT</u>	· · · · · · · · · · · · · · · · · · ·	
N-69 e.139-59dupT - n.t. N.70-78 e.364-8dupT - Africe and Great Britain Note: ANOS gene, Gene Bank accession number NM 213599.2. Nucleotide numbering refets cDNA numbering with +1 corresponding to the A of the ATG + Instalation initiation codon in the reference sequence, according to journal guidelines (www.hgys.org/mutnomen). The initiation codon is codon 1. - Legend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. - - 2 - - - -		*	· · · · · · · · · · · · · · · · · · ·		
N.70-78 e-364-8dupT - Africa and Great Britain Note: AV05 gene. Gene Bank accession number NM_213599.2. Nucleotide numbering reflects cDNA numbering with ±1 corresponding to the A of the ATG + translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. - Legend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity. - - 2 - - - - Africa and Great Britain - - - - 2 - - - - - - 2 - - - - - - - - 2 - <t< td=""><td></td><td></td><td></td><td></td><td></td></t<>					
vote: ANO5 gene. Gene Bank accession number NM_213599.2. Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG ranslation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/muthomen). The initiation codon is codon I. eegend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity.		_			
ranslation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. .egend: *patient N.05 and N.13 also with variant c.364-67T>C with uncertain pathogenicity.			213599.2 Nucleotide numberin		corresponding to the Δ of the ΔTG
2					
2					
2					
John Wiley & Sons, Inc.					2
John Wiley & Sons, Inc.					
John Wiley & Sons, Inc.					
John Wiley & Sons, Inc.					
John Wiley & Sons, Inc.					
John Wiley & Sons, Inc.					
			J	ohn Wiley & Sons, Inc.	
				-	

0 Bage 21 of 27

Page 23 of 27

0 PM

0 BM

Human Mutation

 Table 2: List of detected ANO5 gene changes

Nucleotidic change	Amino acidic change	Exon Intron	Protein domain	Zygosity	Reported	Interpretation?			
c.139-59dupT		IVS3	CTD	1 HzPt	novel	benign in silico			
c.155A>G	p.Asn52Ser	4	CTD	3 CHzPt + 2 HzPt	reported	benign in silico			
c.191dupA		5	CTD	16 HomoZpt+ 19 CHzPt + 5 HzPt	reported	frameshift mutation			
c.242A>G	p.Asp81Gly	5	CTD	1 CHzPt	reported	pathogenic missense chang			
c.364-8delT		IVS7	CTD	5 HomoZpt+ 8 HzPt	novel reported	unconfirmed pathogenicit			
c.364-67T>C		IVS7	CTD	2 HzPt	novel	unconfirmed pathogenicit			
c.400C>T	p.His134Tyr	7	CTD	1 CHzPt	novel	pathogenic in silico			
c.401A>G	p.His134Arg	7	CTD	1 CHzPt	novel	pathogenic in silico			
c.692G>T	p.Gly231Val	8	CTD	5 CHzPt+ 3 HzPt	reported	pathogenic missense chang			
c.746C>G	Ala249Gly	8	CTD	1 HzPt	novel	benign in silico			
c.762+1G>A		IVS8	CTD	1 CHzPt+1 HzPt	novel	putative splice mutation			
c.878+78delT			CTD	1 HzPt	novel	benign in silico unconfirmed pathogenicit			
c.879-41A>T			CTD	1 HzPt	novel				
c.968C>G	p.Ala323Gly	10	CTD	1 HzPt	novel	unconfirmed pathogenici			
c.989dupT		10	CTD	1 CHzPt	novel	frameshift mutation			
c.1097A>G	p.Asn366Ser	11	ETD 1	ETD 1	ETD 1	ETD 1	1CHzPt	novel reported	pathogenic in silico
c.1119+35G>A		IVS11	ETD 1	1 CHzPt	novel	unconfirmed pathogenicit			
c.1120-24A>T		IVS12	ETD1	2 HzPt	novel	unconfirmed pathogenicit			
c.1181-21T>A		IVS13	TMD2	1 HzPt	novel	unconfirmed pathogenicit			
c.1181-48T>A		IVS13	TMD2	1 CHzPt +8 HzPt	novel	unconfirmed pathogenicit			
c.1391C>A	Ala464Asp	14	TMD3	1 CHzPt	novel	pathogenic in silico			
c.1407+5G>T		IVS14	TMD3	1 CHzPt	reported	putative splice mutation			
c.1627dupA		15	CTD3	2 CHzPt	reported	frameshift mutation			
c.1639C>T	p.Arg547STOP	16	CTD3	1 HomoZpt	novel	stop mutation			
c.1640G>A	p.Arg547Gln	16	CTD3	1 HzPt	novel	pathogenic in silico			
c.1643C>T	pThr548Ile	16	CTD3	1 CHzPt	reported	pathogenic in silico			
c.1733T>C	Phe578Ser	16	CTD3	1 CHzPt	reported	pathogenic missense chang			

c.1898+1G>A		IVS17	ETD3	4 CHzPt	reported	putative splice mutation	
c.1925G>T	Arg642Leu	18	ETD3	1 HzPt	novel	pathogenic in silico	
c.2018A>G	Tyr673Cys	18	ETD3	1 CHzPtt	reported	pathogenic missense chang	
c.2030-1G>T		IVS18		1 HomoZpt	novel	putative splice mutation	
c.2235+1G>A		IVS19	TMD7	1 CHzPt	novel	putative splice mutation	
c.2236-13_2236- 4delinsATTCTTCTGGC		20	TMD7	1 HomoZpt	novel	frameshift mutation	
c.2256G>A		20	TMD7	1 HzPt	novel	unconfirmed pathogenicit	
c.2272C>T	Arg758Cys	20	TMD7	1 CHzPt	reported	pathogenic missense chang	
c.2387C>T	p.Ser796Leu	20	TMD7	1 HzPt	reported	benign in silico	
c.2395C>T	p.Arg799STOP	20		1 CHzPt	novel	stop mutation	
c.2417A>G	p.Tyr806Cys	21	ETD4	1 CHzPt+ 1 HzPt	novel	pathogenic in silico	
c.2521-13A>G		IVS21	TMD8	1 HzPt	novel reported	unconfirmed pathogenicit	
c.2521C>G	p.His841Asp	22	TMD8	1 HzPt	novel	pathogenic in silico	
c.2593G>T	p.Ile865Leu	22	TMD8	1 HzPt	novel	pathogenic in silico	
c.2698A>C	p.Met900Leu	22	CTD5	1 HzPt	novel	benign in silico	

Note: *ANO5* gene, Gene Bank accession number NM_213599.2. <u>Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1. Legend: CTD: cytoplasmic topological domain; ETD: extracellular topological domain; TMD: transmembrane domain; CHzPt: compound heterozygous patient; HomoZpt: homozygous patient; IVS: intron</u>

				Onset	CI I	Pattern of Muscle involvement at last assessment											
Patient	Patient Age (years)			Age (years) Sex	Age	Symptoms	CK (IU/L)	ambulant	UL prox	LL Prox	LL Distal	Walk on toes	Walk on heels	Muscle atrophy	scapular winging	AS	other features
N.01	50	m	38	thigh weakness	241-2603	yes	-	+++	-	diff.	diff.	thighs	-	+	-		
N.02	68	f	64	proximal LL weakness, thigh wasting	n.a.	yes	+/-	+++	-	diff	diff	thighs and calves (AS)	-	+			
N.03	20	f	-	-	3300-8800	yes	-	-	-	able	able	-	-	-	-		
N.04	63	m	50s	walking difficulties	2122	yes	+/-	+++	+	n.a.	n.a.	-	-	-	calf hypertrophy, KH		
N.05	44	m	20s	Thigh and calf wasting (AS)	4060	yes	-	+++	+/-	able	able	calves, thighs (AS)	-	+	Calf hypertrophy (as child), TA contractures, ptosis		
N.06	35	m	20s	calf swelling, back pain, leg weakness	1642-3080	yes	+/-	++	++ (AS)	unable	unable	-	-	+	muscle twitching, TA contractures		
N.07	56	m	40s	LL weakness and wasting	1517	yes	++	+++	+	unable	unable	n.a.	-				
N.09	40	m	20s	reduces sport fitness and muscle wasting	4000	yes	+/-	++	-	able	able	biceps/pectoral. quads (AS)	-	+			
N.10	64	m	46	Raised CK and myalgia	2200-3300	yes	+/-	+		able	able	quads	-	-	Sleep apnoea, CTS		
N.11	43	m	20s	n.a.	5400	yes	no	-	+	diff	diff	calves (AS)	-	+	thigh hypertrophy		
N.12	16	m	16	myalgia, raised CK	2000-30,000	yes	+/-	-	-	able	able	-	-	-	neck flexion weakness		
N.13	70	m	38	Difficulty running and calf asymmetry	1800	yes	+/-	+ (AS)	+	unable	unable	biceps and calves (AS)	-	+	Calf hypetrophy (AS)		
N.14	42	f	30s	prox LL weakness	2500-3000	yes	-	+	-			vasti	+/-	n.a.	calf hypertrophy, CTS		
N.15	67	m	50s	raised CK and myalgia	2100	yes	-	-	-	able	able	-	-	n.a.	calf hypertrophy		
N.16	44	m	40	Myalgia	1500-2500	yes	+/-	-	-	able	able	-	+	-			
N.17	47	m	20s	myalgia	1600-6000	yes	-	+++	-	able	unable	thighs, hamstrings especially, calf (AS)	-	+	thigh hypertrophy, CTS		
N.18	65	m	30s	walking difficulties	3661	no 60yrs	-	++++	-	unable	unable	quads	-	-	calf hypertrophy		

Table 3: Phenotypic analysis of patients with 2 pathogenic ANO5 variants

Human Mutation

N.19	40	m	34	LL weakness	5000	yes	-	+++	+/-	able	able	hamstrings, buttocks, calves	-	-	
N.20	65	m	40s	walking difficulties	2000	yes	-	+	-	n.a.	n.a.	n.a.	-	n.a.	
N.21A	14	f	-	-	1200	Yes	-	-	-	able	able	-	-	-	-
N.21B	17	m	-	-	n.a.	-	-	-	-	able	able	-	-	-	-
N.22A	72	m	50s	walking difficulties	900	yes	+ (AS)	+++ (AS)	n.a.	diff	diff	+	+	+	
N.22B	74	m	40s	UL weakness	900	yes	+	+ (AS)	n.a.	diff	diff	medial gastrocnemius (AS)	+	+	calf, paraspinal muscle hypertrophy
N.23	41	m	38	LL weakness and wasting	209-6301	yes	+/-	+++	-	unable	diff	thighs and calves (AS)	+ (AS)	+	KH (AS)
N.24	55	m	28	weakness in gastrocnemius muscle	2500-4500	yes	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	
N.25	42	m	23	Wasting and weakness left calf	5300-12500	yes	-	-	++	unable	able	calves	-	+	
N.27	41	f	34	hyperCKaemia	2503	yes	-	-	-	able	able	gastrocnemius medialis (AS)	-	+	
N.28A	56	m	40	asymmetric thigh weakness	1500-6100	yes	+/-	++	-	diff	diff.	Thighs (AS)	-	+	
N.28B	53	f	-	-	2280	yes	-	-	-	able	able	-	-	-	
N.29	64	m	30s	difficulties with climbing stairs	829	yes	+/-	+++	+++	unable	unable		-	+	calf hypertroph (AS), IHD Anaemia, FVC 70%
N.30	61	m	30	weakness in LL muscles	5956	yes	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	
N.31	28	m	20	difficulties running	7868	yes	-	+++ (AS)	-	able	able	UL, bamstrings, buttocks, quads	-	+	sensory axonal polyneuropathy
N.32	52	m	20s	hyperCKaemia and calf wasting	>3000	yes	+/- (AS)	+++	-	unable	able	vastus medialis, calf (AS)	+/-	+	TA contractures
N.33	32	m	20s	LGMD type weakness	n.a.	restricted	-	+/-	+	n.a.	n.a.	biceps, medial quads, calf (AS)	n.a.	+	
N.34	32	m	22	calf wasting, unable to stand on toes	10,000	yes	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	
N.35	29	m	23	Calf muscles stiffness when walking	1700-2900	yes	-	-	+/-	diff	unable (AS)	calves (AS)	-	+	
N.36	54	m	32	proximal LL weakness	4029	yes	-	+/-	-	able	able	-	-	-	
N.38	77	m	77	proximal LL weakness and raised CK	4400	yes	n.a.	+ (AS)	-	able	able	LL (AS)	_	+	

N.39	59	m	38	left LL weakness	3000	yes	-	+/-		unable	unable	thighs (AS)	-	+	hypertension, KH (AS)
N.40A	32	m	21	decreased endurance, inability standing on right tiptoes and raised CK	4200-22.000	yes	-	++	+	unable	able	adductors, quadriceps and calves	-	+	КН
N.40B	13	m	-	-	1970	yes	-	-	-	able	able	-	-	-	
N.40C	23	f	21	myalgia	861	yes	-	-	-	able	able	-	-	-	
N.41	56	m	35	difficulties running	1168-3612	yes	-	++	++	unable	able	thighs and calves	-	-	
N.42	44	m	39	distal weakness and calf wasting	2666- 4200	yes	-	+	++	unable	able	UL (distally), thighs, calves	-	-	hand weakness, back twitching

Legend: Pt, patient number; CK, creatine kinase; UL, upper limbs; LL, lower limbs; AS, asymmetry; m, male patient; f, female patient; diff, able with difficulties; n.a, data not available; TA, Achilles tendons; KH, knee hyperextension; IHD, ischaemic heart disease; FVC, forced vital capacity; CTS: carpal tunnel syndrome; +, ++, +++: grade of weakness (from milder to more severe)

2	Tał
3	
4 5	
6	
7	Pa
8 9	
9	
10 11	
12	
13	
14	
15 16	
17	
18	
19	
20	
21 22	Leg
23	AT
24	
25	
26	
27 28	
29	
30	
31	
32	
33 34	
35	
36	
37	
38	
39 40	
40 41	
42	
43	
44	
45	
46	
47 48	
4ŏ	

10

 Table S1. Patients with ANO5 gene mutations

Patient	allele 1	allele 2	Geographic origin					
Patients with exonic and intronic changes with uncertain pathogenicity								
N.57-N.58	c.155A>G	-	Great Britain (2)					
N.59	c.746C>G	-	Great Britain					
N.60	c.968C>G	-	Pakistan					
N.61	c.2256G>A	-	Great Britain					
N.62	c.2387C>T	-	Great Britain					
N.63	c.2698A>C.	-	Germany					
N.64-67	c.364-8dupT	c.364-8dupT	Africa (2) and Great Britain (2)					
N.68	c.1119+35G>A	c.1181-48T>A	Germany					
N.69	c.139-59dupT	-	n.a.					
N.70-78	c.364-8dupT	-	Africa (1) and Great Britain (8)					

Legend: ANO5 gene, Gene Bank accession number NM_213599.2. Nucleotide numbering reflects cDNA numbering with +1 corresponding to the A of the ATG translation initiation codon in the reference sequence, according to journal guidelines (www.hgvs.org/mutnomen). The initiation codon is codon 1.