Pica 8: Refining dietary reconstruction through amino acid δ^{13} C analysis of tendon collagen and hair keratin

Alice Mora^{a*}, Aryel Pacheco^b, Charlotte Roberts^b & Colin Smith^a

^a Department of Archaeology and History, La Trobe University, Bundoora, 3086, VIC, Australia;

^b Department of Archaeology, Durham University, Durham, DH1 3LE, UK.

*Corresponding author:

A.Mora@latrobe.edu.au

Highlights:

A method for assessing a terrestrial or marine origin for dietary intake is proposed The new method uses δ^{13} C values of phenylalanine, valine, and leucine Tendon collagen is a favourable substitute to bone collagen in dietary reconstructions Tendon is an ideal tissue for characterising the final year of an individual's life 1 Abstract

2 Stable isotope analysis of archaeological human remains is routinely applied to explore dietary habits and

3 mobility patterns. The isotope information pertaining to the period prior to death may help in identifying

4 locals and non-locals, especially when investigating individuals from the same funerary context but believed

5 to have been highly mobile across the landscape.

6 Based on the variety of the funerary goods in graves and what it is believed their diets comprised, it is

7 thought that both local and non-local individuals were buried at the inland funerary site of Pica 8 (northern

8 Chile, Late Intermediate Period, ~1,050-500 BP); however, uncertainties over the dietary intakes and

9 mobility histories of these individuals still persist. The aim of this study is to refine the dietary

10 characterization of a subset of Pica 8 individuals by increasing the temporal resolution of their dietary

11 reconstructions, specifically throughout the last period of their life, and by identifying the multiple sources of

12 food in their overall diets. This is achieved by analysing the amino acid carbon isotope composition of hair

13 keratin and, for the first time, that of tendon collagen.

14 This study proposes a new method for identifying the predominant food source (terrestrial or marine) in a

15 mixed diet using phenylalanine, valine and leucine δ^{13} C values measured in collagenous tissues. Herein,

16 tendon is proven to be an ideal tissue for isotopically characterising the final year of an individual's life. Our

17 results show that individuals previously identified as non-locals, based on long-term food consumption, had

18 in reality abandoned their original dietary habits typical of distant regions many months before death, and

- 19 hence had presumably relocated to the locality of Pica.
- 20
- 21

22 Keywords: Chile; tendon; collagen; amino acid; LC-IRMS; stable isotope; palaeodiet.

- 23
- 24
- 25
- 26
- 27
- 28
- 29
- 30

31 1. Introduction

32 Stable isotope analysis is routinely applied to archaeological human remains for characterising dietary habits 33 and mobility patterns of past populations (Makarewicz and Sealy, 2015). In recent years, attention has 34 focussed on the reconstruction of the life histories of individual identities in past societies (Knapp and van 35 Dommelen, 2008). The reconstruction of an individual's life at different points over their life course is 36 achieved by isotopically analysing multiple tissues (skeletal and non-skeletal) from the same individual, 37 which have differential deposition times and/or turnover rates (Lynnerup, 2007). In particular, the analysis of 38 soft tissues, that have fast remodelling rates, gives information pertaining to the last year/months of life 39 (Lamb, 2015). Moreover, hair retains an unaltered isotope signal locked into the keratins when the tissue was 40 growing (Petzke et al., 2010). This isotope information pertaining to the period prior to death is important 41 when investigating individuals from the same funerary context but believed to have been highly mobile 42 across the landscape, or having had access to long-distance resources. Individuals identified as non-local 43 because of their diet, based on stable isotope compositions averaged over several years (from, for instance, 44 bone collagen) may, in reality, have been consuming locally accessible resources and been part of the local 45 community for a considerable period of time before death.

46 At the inland funerary site of Pica 8 in northern Chile (Late Intermediate Period, ~1,050-500 BP),

47 uncertainties over dietary intakes and mobility histories of the buried individuals still persist, especially

48 pertaining to the last period of their life. Previous archaeological and biomolecular studies (Núñez, 1984,

49 Petruzzelli et al., 2014, Santana-Sagredo et al., 2015a) have suggested that the individuals buried at Pica may

50 have had different geographical origins and/or a high degree of mobility. Núñez (1984), who first excavated

51 the cemetery, identified the presence of non-local items among the funerary inclusions in the graves (e.g.

52 pottery, textiles, bird feathers, foods). These were thought to have been imported from either the eastern

53 Andes, Altiplano, Azapa valley, or from the coast. Stable isotope analyses of bone and tooth enamel have

54 identified three major dietary related groups of people, consisting of: (1) individuals relying mainly on

55 marine resources, complemented by some maize, (2) individuals relying mainly on maize, complemented by

56 marine resources, and (3) individuals consuming predominantly C_3 terrestrial resources (Santana-Sagredo et 57 al., 2015a).

58 Based on the variety of grave goods and diets, it can be argued that the people buried at Pica 8 represent a

59 combination of local and non-local individuals who were involved in inter-regional exchange of foodstuffs

and exotic objects, and/or sedentary individuals who benefited from having access to a broad range of

61 resources and maintained dietary habits distinctive of their place of origin. Despite the paucity of information

62 surrounding the placement of the burials in the cemetery, there is some evidence (i.e. broad distribution of

63 funerary goods and diets) that the Pica 8 cemetery was divided into ten sectors, A to J (Núñez, 1984,

64 Santana-Sagredo et al., 2015a, Zlatar, 1984), where people with common geographic origins, socio-

65 economic status, and/or cultural traits may have been interred.

In light of new radiocarbon dates of paired human and camelid tissues and estimated ¹⁴C offsets, Santana-66 67 Sagredo and colleagues (2017) have reconsidered the original dietary interpretation (Santana-Sagredo et al., 2015a) of the Pica individuals, proposing the consumption of C_4 crops fertilised with seabird guano as a 68 major cause for the high δ^{15} N values (>20‰) measured in bone collagen, rather than the direct consumption 69 70 of marine resources. Andean archaeological and ethnohistoric records (Covey, 2000, Denevan, 2001, Julien, 71 1985, Marcus et al., 1999) report that guano was traditionally mined and marine fish were procured and dried 72 by coastal communities and transported to the highlands via llama caravans. Estimating the proportion of 73 guano-fertilised maize and marine fishes in the diet of the Pica individuals is not straightforward since the practice of fertilising maize with guano increases the plant δ^{15} N values to as much as 20%, but does not 74 affect δ^{13} C values (Szpak et al., 2012a, 2012b). As a result, consumption of high-trophic level marine 75 76 resources (fish and mammals) and guano-fertilised C₄ crops may generate similar ranges of bulk δ^{13} C and 77 δ^{15} N values in human tissues (Szpak et al., 2012b). Single amino acid carbon isotope analysis may help in 78 identifying these diverse sources (marine, terrestrial) of food macronutrients (protein, carbohydrate, lipid) by 79 comparing the δ^{13} C values of essential and non-essential amino acids, given that different metabolic 80 pathways are involved in the processes of assimilation or synthesis of the amino acids making up whole 81 proteins (Newsholme et al., 2011, Reeds, 2000), and that guano is not affecting the carbon stable isotope 82 compositions of plant tissues (Szpak et al., 2012a).

83 The aim of the present study is to refine the dietary characterization of a subset of Pica 8 individuals by 84 increasing the temporal resolution of palaeodietary reconstructions, specifically throughout the last period of 85 their life, and by identifying the different sources of food (marine and terrestrial) that likely comprised their 86 mixed dietary intakes. This is achieved by analysing the amino acid carbon isotope composition of hair 87 keratin and, for the first time, that of tendon collagen. Tendon has the potential to be an ideal substrate for 88 characterising the period leading to/close to the time of death, considering that (1) collagen is significantly 89 more abundant in tendons (~677+57 nmol/g wet tissue) than in the dermis (~335+64 nmol/g wt), bone 90 (~307+71 nmol/g wt) or skeletal muscle (~59+17 nmol/g wt) (Kjaer et al., 2005), and that (2) the rate of 91 collagen turnover in tendon (Babraj et al., 2005, Miller et al., 2005) is more rapid than in bone (Hedges et al., 92 2007).

93

94 2. The cemetery of Pica 8

95 Pica 8 is located approximately 80 km inland, at circa 1,300 m of elevation, on the plain of the Pampa del

96 Tamarugal (northern Chile, Fig. 1) (Jayne et al., 2016). This funerary site comprises 254 burials (Núñez,

97 1984) and dates to the Late Intermediate Period (~1,050-500 BP), based on ceramic seriation and

98 radiocarbon dating (Núñez, 1976, Santana-Sagredo et al., 2017, Uribe et al., 2007). The Pica oasis was part

99 of a complex system of communities situated in the Tarapacá region between the Río Camiña in the north

and the Río Loa in the south, and covered an altitudinal transect from the coast to the Precordillera (~2,500

101 masl) (Uribe et al., 2007).

102 [Figure 1. Map of northern Chile showing the location of the Pica 8 site.]

103 During this period, the Pica-Tarapacá oases were connected to each other by a complex network of routes

and campsites, which allowed long-distance trade of resources and exotic objects between the coast and the

105 highlands, via llama caravans (Briones et al., 2005, Pomeroy, 2013). In this arid region, inter-regional

106 redistribution of surpluses between ecologically different zones was crucial, especially in times of shortage

107 of staple resources (Núñez and Dillehay, 1979, Zori and Brant, 2012). Competition over the control of this

108 trade network likely generated disputes between the local elites. The fact that artefacts related to conflict

109 (found as grave goods) were not associated with violence-related bone injuries, notwithstanding that soft

110 tissue injuries may have occurred, suggests that underlying tension was sublimated in symbolic celebrations

111 of power as a means to reinforce the leadership of certain elites (Pacheco and Retamal, 2017).

112

113 3. Materials

114 The Pica 8 collection is curated at the Department of Anthropology, University of Chile, Santiago de Chile

115 (Lemp et al., 2008), and comprises approximately 150 naturally mummified individuals. Six adult

116 individuals with varying ages, four females and two males, were selected for analysis in order to include

bodies deposited in burial sectors that have been suspected to be linked to people with a diversity of diets,

118 geographical origins and cultural identities (Núñez, 1984, Retamal et al., 2012, Santana-Sagredo et al.,

119 2015a). A foot tendon and a bundle of scalp hair were sampled from each individual.

120

121 4. Methods

122 4.1. Tendon collagen stable isotope analysis

123 Tendon was processed following the Finucane method (2007) originally proposed for soft tissues such as

skin and muscle, but increasing the number of washing steps before protein denaturation. In detail, the

125 tendons were cleaned by sonication in Milli-Q water (Merck), immersed in a mixture of

126 chloroform:methanol (2:1 v/v) (Merck), sonicated for ~20 min, and soaked overnight to remove any lipid

- 127 content. Every 24 hours, the chloroform:methanol (2:1 v/v) was replaced and sonicated (20 min) until all
- 128 lipids were removed. The samples were then rinsed six times with Milli-Q water (sonicated each time for 20

129 min). A few drops (~2-3) of 0.5M HCl (Merck) were added to the samples immersed in fresh Milli-Q water

- 130 to obtain a pH 3 solution. Sealed tubes were placed into a heater block for 48 h at 73°C for gelatinization.
- 131 The supernatant was filtered with an Ezee-Filter[™] (Elkay Laboratory Products), and the residue was
- 132 discarded. Each filtered sample was then decanted into Pyrex® glass tubes (Corning), frozen (-20 °C), and
- 133 then freeze-dried (48 h).

134

135 4.1.1. Bulk carbon and nitrogen stable isotope analysis

- 136 The lyophilised collagen was inserted into tin capsules for the analysis of carbon and nitrogen isotope
- 137 compositions, which was performed using a Carlo Erba CE1110 CHN-S analyser coupled to a Fisons
- 138 Isochrom Continuous-flow Isotope Ratio Mass Spectrometer (GV instruments). The δ^{13} C and δ^{15} N values are
- 139 reported relative to international standards: V-PDB and AIR, respectively. Secondary (reference) materials
- 140 and in-house standards (MZ1 Maize, HAR4 Sunflower, ATP12) were used to monitor analytical precision of
- 141 the carbon and nitrogen isotope ratio measurements that were $\pm 0.1\%$ (1 σ) for both elements.
- 142
- 143 4.1.2. Single amino acid carbon isotope analysis
- 144 Approximately 1 mg of tendon collagen was hydrolysed under vacuum in 1 ml of amino acid-free 6M HCl
- 145 (Sigma-Aldrich) at 110 °C for 24 h, after Choy et al. (2010). The hydrolysates were dried in a rotary vacuum
- 146 concentrator and frozen until required for analysis. Prior to analysis, samples were dissolved under
- sonication in Milli-Q water, with the addition of 10 µl of 1mmol solution of 2-aminoisobutyric acid (Sigma-
- 148 Aldrich) as the internal standard (I.S.); this was to obtain a sample stock of $\sim 1.8 \,\mu g/\mu l$. The sample stock was
- 149 further diluted in Milli-Q water to deliver 10.8 to 16.2 µg of protein on the LC column (10 µl partial loop
- 150 injection mode).
- 151 LC-IRMS analysis was undertaken at the La Trobe Institute for Molecular Sciences (LIMS, La Trobe
- 152 University, Melbourne, Australia) using an Accela 600 pump connected to a Delta V Plus Isotope Ratio
- 153 Mass Spectrometer via a Thermo Scientific LC Isolink (Thermo Scientific). The amino acids were separated
- using a mixed-mode Primesep A column (2.1x250 mm, 100 Å, 5 μm, SIELC Technologies) following the
- 155 chromatographic method described in Mora et al. (2017), after Smith et al. (2009). Mobile phases were:
- 156 Phase A = 35 μ l of 1:50 96% H₂SO₄ (Merck) in 1L Milli-Q water, Phase B (1L)= 1 ml 96% H₂SO₄ (Merck)
- and 2.28 g of \geq 98% K₃PO₄ (Sigma-Aldrich) in Milli-Q water, and Phase C (1L)= 3 ml 96% H₂SO₄ (Merck)
- 158 in Milli-Q water. The LC gradient program was similar to that of Mora et al. (2017), but the flow rate of the
- analytical run was decreased to $60 60 \mu$ /min (Table 1), in order to improve the baseline resolution of the
- 160 isoleucine, leucine, lysine, histidine and tyrosine peaks. When needed, the flow rate was increased to 80 or
- 161 100 μ l/min after the tyrosine peak to gain faster sample elution.
- 162 [Table 1. Here]
- 163 Samples were analysed in duplicate. Baseline separation was achieved for all the amino acids in the tendon
- 164 collagen, with the exception of methionine (Fig. 2). Histidine and methionine peaks were too small to
- 165 generate reliable δ^{13} C values. Overall, the amino acid carbon contribution measured by LC-IRMS analysis
- 166 corresponded to 98.4% of the carbon present in human tendon collagen (Schofield et al., 1971).
- 167 [Figure 2. LC-IRMS chromatogram of tendon collagen hydrolysate (from individual SE-T3).]
- 168

169 4.2. Bayesian stable isotope mixing model: FRUITS

- 170 The Bayesian mixing model FRUITS Food Reconstruction Using Isotopic Transferred Signals (Fernandes
- 171 et al., 2014) has been applied to the collagen stable isotope data produced in the current and previous studies
- 172 (Santana-Sagredo et al., 2015a) to achieve qualitative and quantitative estimates of the nutritional intake of
- the Pica individuals. Although other Bayesian stable isotope mixing models have comparable features to
- those of FRUITS (Phillips et al., 2014), we have used FRUITS as it has been commonly preferred by
- 175 researchers for reconstructing human palaeodiets, especially from Chilean contexts (e.g. Andrade et al.,
- 176 2015, King et al., 2018, Mora et al., 2017, Pestle et al., 2016, Pestle et al., 2017) in recent years. Details can
- 177 be found in Appendix A.
- 178
- 179 4.3. Hair keratin amino acid carbon isotope analysis

180 Amino acid δ^{13} C analysis was performed on 1 cm segments of a single hair from each individual, cut

181 longitudinally along the hair fibre, from the root to the first 10 cm. Hair samples were inserted into

- 182 hydrolysis tubes, and left to soak in methanol for 12-24 h in order to remove organic residues. Once dried,
- 183 the hair segment was hydrolysed in 6 M hydrochloric acid at 110°C until the hair fibre was completely
- 184 dissolved. The amount of 6M HCl used for hydrolysis was increased to $\sim 0.175 \,\mu g/\mu l$, compared to Mora et
- al. (2017). Hydrolysed samples were then dried overnight at 30°C in a rotary vacuum concentrator and
- 186 frozen until required for analysis. For LC-IRMS injection, samples were dissolved under sonication in Milli-
- 187 Q water, supplemented by an internal standard (2-aminoisobutyric acid), to obtain keratin hydrolysates with
- 188 a concentration of approximately $0.8 \,\mu\text{g/}\mu\text{l}$. Samples were analysed in duplicate. The LC-IRMS analysis is
- 189 detailed in section 4.1.2.
- 190
- 191 5. Results and Discussion
- 192 5.1. Elemental and molecular composition of tissues
- 193 5.1.1. Assessment of the preservation of tendon collagen
- 194 The calculated C/N atomic ratios (3.2 to 3.4, Table 2) fall within the range of 2.9-3.6, proposed by DeNiro
- 195 (1985) for well-preserved bone collagen. This range is likely applicable to tendon collagen as the most
- abundant protein in both tissues is type I collagen (Kannus, 2000, Wang, 2006), and the C/N ratios measured
- by Ambrose (1990) in bovine tendon were comparable to those measured in bone collagen. The content by
- 198 weight (%) of carbon ($45.8\pm0.7\%$) and nitrogen ($16.0\pm0.2\%$) of the tendons analysed herein is comparable to
- that measured in bovine tendon collagen (47.6±1.5 %C and 16.0±0.6 %N) by Ambrose (1990). Based on
- 200 these criteria, the tendon collagens submitted for analysis were well preserved.
- 201 [Table 2. here]

203 5.1.2. Assessment of the molecular preservation of tendon collagen

- 204 To assess the molecular preservation of tendon collagen samples, the amino acid Area All [V] values (i.e.
- sums of the peak areas for the ion currents at m/z 44, 45, 46), generated by LC-IRMS, were converted to
- 206 fractions of the total (%) and compared to (1) those derived from archaeological human bone collagen (n=12,
- 207 excluding tyrosine) analysed under similar chromatographic conditions, and to (2) the amino acid carbon
- 208 weights (%) of human tendon collagen estimated from the amino acid residues published by Schofield et al.
- 209 (1971). The residues were firstly multiplied by the number of carbon atoms per residue to determine the
- amino acid carbon contribution, and the fraction of the total was calculated to make the residues comparable
- 211 to the LC-IRMS output. Based on this comparison, it was possible to show that the tendon collagen samples
- had preserved their amino acid composition. Amino acid profiles (mean $\pm 1\sigma$) are shown in Fig. 3.

[Figure 3. Fractions of the total (%) of amino acid peak areas measured in archaeological tendon collagen

(this study) and bone collagen (mean $\pm 1\sigma$), and of amino acid carbon weights (%) of human tendon collagen

- 215 derived from Schofield et al. (1971).]
- 216

217 5.1.3. Assessment of the molecular preservation of hair keratin

218 To assess the molecular preservation of the hair keratins, the amino acid peak areas were converted to

219 fractions of the total (%) and compared to (1) those of modern human hair prepared following the same

220 procedure as for the archaeological hair, but preceded by methanol:chloroform (2:1 v/v) washings to remove

any residue of detergents and lipids (O'Connell et al., 2001), and to (2) the amino acid carbon weights (%) of

human hair derived from Wolfram (2003) and Robbins and Kelly (1970). Based on this comparison, it was

- 223 possible to assess that the hair keratin samples had preserved their amino acid composition. Amino acid
- 224 profiles (mean $\pm 1\sigma$) are shown in Fig. 4.
- [Figure 4. Fractions of the total (%) of amino acid peak areas measured in archaeological and modern hair
- (mean $\pm 1\sigma$), and of amino acid carbon weights (%) measured in human hair (Robbins and Kelly, 1970,
- 227 Wolfram, 2003).]
- 228

229 5.2. Bulk stable isotope compositions and FRUITS diet estimates

230 The δ^{13} C and δ^{15} N isotope values measured in this study from tendon collagen were compared to those

231 measured by Santana-Sagredo et al. (2015a) in bone collagen from the same individuals (Table 2). Due to the

232 differential turnover rates, rib collagen isotope composition represents an average of the dietary intake over

the last three to five years of life (Jørkov et al., 2009), while tendon collagen reflects the food consumed

during the last few months of life (Babraj et al., 2005). Previous carbon and nitrogen isotope studies (Basha

202

235 et al., 2016, Finucane, 2007, Iacumin et al., 1998, White and Schwarcz, 1994) on bone and soft tissues 236 (tendon, muscle, skin) from archaeological human remains have reported differences in the isotope values 237 between tissues from the same individual. Multiple and concurrent factors may be responsible, including: (1) 238 changes in diet and/or in physical condition experienced by an individual at different points in his/her life 239 course (Neuberger et al., 2013, Reitsema, 2013, Warinner and Tuross, 2010), (2) different protein (and 240 amino acid) composition of the tissues (e.g. type I collagen vs. actin and myosin) (Eastoe, 1955, Schofield et 241 al., 1971), and (3) diverse fractionation processes that may be connected to the remodelling of the various 242 proteins and/or to their specific metabolism (Ambrose and Norr, 1993, Hare et al., 1991, Tieszen et al., 243 1983). If it is assumed that different fractionation processes exist for bone and tendon collagen, connected to 244 their specific metabolism and remodelling, these effects are likely to be much smaller than the changes in 245 diet experienced by individuals at different points in their life course, which are differentially recorded in 246 tendon and bone, depending on their turnover rates. Having taken the above into account, the diet-to-

collagen offset for tendon collagen is assumed herein to be comparable to that of bone.

248 Given these premises, the Bayesian mixing model Food Reconstruction Using Isotopic Transferred Signals 249 (FRUITS) (Fernandes et al., 2014) was applied to the tendon collagen isotope data produced in this study and 250 to the bone collagen isotope data published by Santana-Sagredo et al. (2015a). The food groups used in the mixing model include the widest array of foods that were available through trade from the Pacific coast, 251 252 coastal valleys, mid-altitude valleys and highlands (Aufderheide et al., 1994, Burress et al., 2013, DeNiro 253 and Hastorf, 1985, DeNiro, 1988, Falabella et al., 2007, Finucane et al., 2006, Gil et al., 2011, Hoeinghaus et 254 al., 2011, Hückstädt et al., 2007, Szpak et al., 2013, Szpak et al., 2014, Szpak et al., 2015, Thornton et al., 255 2011, Tieszen and Chapman, 1992, van Der Merwe et al., 1993). The edible portion of flora and fauna (e.g. 256 animal muscle, plant fruit, tuber or grains, etc.) was calculated by applying suitable offsets (depending on the 257 type of tissue and taxonomic rank) (Codron et al., 2005, Hobson and Clark, 1992, Hobson et al., 1996, Kelly, 258 2000, Mateo et al., 2008, Sealy et al., 1987, Sholto-Douglas et al., 1991, Vogel, 1978, Warinner and Tuross, 259 2010, Yoneyama and Ohtani, 1983). A correction of 1.5‰ was applied to the δ^{13} C values of modern samples 260 to account for the Suess effect (Marino and McElroy, 1991, Schloesser et al., 2009). In Fig. 5, the tendon 261 collagen isotope data produced in this study and the bone collagen isotope data published by Santana-262 Sagredo et al. (2015a) are compared against the edible portion of South American flora and fauna.

[Figure 5. Plot of δ^{15} N values versus δ^{13} C values of tendon collagen (*, this study) and bone collagen (†, Santana-Sagredo et al., 2015a) from Pica 8 individuals and of the edible portion of South American flora and fauna.]

266 The food groups used in the FRUITS model consist of: (1) C₄ plants (cultivated and wild), (2) marine

267 animals (fish, sea lions, shellfish), (3) C_3 plants (fruits, vegetables, legumes), and (4) terrestrial animals

268 (camelids, rodents and birds) consuming either C_3 , C_4 or mixed C_3 - C_4 plants (i.e. wild and domesticated

animals living at different elevations). To account for the use of guano (seabird dung) in inland maize

270 cultivation, a second model was run, substituting the 'C₄ plants' food group with 'maize manured with

- 271 guano'. Guano was commonly imported from the coast to increase the productivity of high-altitude
- cultivation (Ajata López, 2013). The diet estimates generated by the FRUITS models are provided in Tables
- 273 3-4. The proportion of animal meat in reconstructed diets changes significantly between the two models. The
- dietary estimates of the second model (Table 4), compared to the first (Table 3), indicate a greater
- 275 contribution of terrestrial animal meat and a lower contribution of marine foodstuffs. In the second model
- (Table 4), the proportion of marine resources is lower than that originally proposed by Santana-Sagredo et al.
- 277 (2015a).
- 278 [Tables 3 and 4. Here]
- 279 The tendons of four individuals (SD-T24, SI-T32, SI-T3, SF-T4) recorded similar bulk isotope values ($\delta^{13}C$ =
- 280 -7.8‰ to -8.7‰, $\delta^{15}N$ = +15.5‰ to +17.5‰, Table 2 and Fig. 5), which, according to FRUITS estimates,
- reflect a dietary intake based mostly on C₄ plant products (56-72% of the food intake), complemented by
- animal meat of terrestrial (9-34%) and/or, to a lesser extent, marine (3-15%) origin (Tables 3-4). FRUITS
- dietary estimates (Tables 3-4), based on tendon collagen isotope data ($\delta^{13}C = -18.3\%$, $\delta^{15}N = +12.2\%$, Table
- 284 2 and Fig. 5), indicate that the young woman SE-T3 consumed predominantly C_3 plant products (77-79%)
- and some meat of terrestrial animals (~11%) during the final months of her life, whereas the young woman
- SI-T74 (δ^{13} C= -10.8‰, δ^{15} N= +25.3‰, Table 2 and Fig. 5) had a significant caloric contribution from
- 287 marine proteins (71-81%, Tables 3-4).
- 288
- 289 5.3. Amino acid carbon stable isotope compositions

290 The tendon collagen amino acid (AA) δ^{13} C values are reported in Appendix B. The δ^{13} C mass balance 291 values, calculated based on tendon amino acid composition reported by Schofield et al. (1971), differ from 292 the measured bulk δ^{13} C values by 0.55±0.11‰. The full dataset of hair keratin amino acid δ^{13} C values is 293 provided in Appendix C.

294 The study of stable carbon isotope compositions of human proteins at the amino acid level makes it possible 295 to track the various sources of the dietary macronutrients because of the different carbon isotope 296 fractionations associated with the processes of assimilation and biosynthesis of different amino acids into 297 human proteinaceous tissues. The extent of the carbon isotope fractionation between diet and bone collagen 298 has been investigated through stable isotope analysis of tissues taken from animals raised on controlled diets 299 (Copley et al., 2004, Hare et al., 1991, Howland et al., 2003, Jim et al., 2006, Webb et al., 2017). Essential 300 amino acids present minimal isotope fractionation since they are assimilated directly from dietary proteins 301 into the body tissues (Newsholme et al., 2011, Reeds, 2000). In particular, bone collagen δ^{13} C values of some essential amino acids such as leucine and phenylalanine were found to closely reflect the δ^{13} C values of the 302 respective dietary amino acids, thus being useful for identifying the source of the protein component of the 303 diet (Copley et al., 2004, Howland et al., 2003, Jim et al., 2006). At the base of the food chain, amino acid 304 305 δ^{13} C values of C₄ plant species are enriched in the ¹³C isotope compared with those of the C₃ plants (Fogel

- and Tuross, 2003), and amino acid δ^{13} C values of organic matter from marine ecosystems are enriched
- 307 relative to freshwater-derived amino acids (Keil and Fogel, 2001). Non-essential amino acids can be
- 308 synthesised *de novo*, in addition to direct assimilation. This means that the human body may synthesise these
- amino acids through metabolic pathways involving all the three macronutrients (proteins, carbohydrates,
- 310 lipids) (Newsholme et al., 2011, Reeds, 2000). Among non-essential amino acids, alanine is preferentially
- 311 synthesized by the body, rather than routed, even under high-protein intake (Fernandes et al., 2012, Jim et
- al., 2006), making this non-essential amino acid useful for identifying the non-protein portion of the diet.
- A number of dietary proxies for human palaeodietary reconstruction based on bone collagen amino acid δ^{13} C
- values have been proposed (e.g. Choy et al., 2010, Corr et al., 2005, Honch et al., 2012, Webb et al., 2016).
- Arguably the greater separation of individuals belonging to four dietary groups (HMP, HFP, C4, C3, i.e.
- 316 consumers of predominantly high-marine proteins, high-freshwater proteins, C4 plants, or C3 plants) was
- 317 achieved using the dietary proxy $\Delta^{13}C_{\text{Val-Phe}}$ and thus by plotting $\delta^{13}C$ phenylalanine vs. $\delta^{13}C$ values
- 318 (Honch et al., 2012). When the bone collagen $\Delta^{13}C_{Val-Phe}$ proxy (Honch et al., 2012) is applied to our dataset,
- the Pica individuals all fall within the range of terrestrial consumers (Fig. 6). This is difficult to reconcile
- 320 with the results of the FRUITS models, as the $\Delta^{13}C_{Val-Phe}$ dietary indicator appears to underestimate the
- 321 marine protein intake, especially in the case of SI-T74, who has been identified through the FRUITS models
- 322 to have a diet containing a significant marine component (41% to 51%).
- 323 [Figure 6. Plot of $\angle 1^{13}$ C _{Val-Phe} values for tendon collagen (this study) and bone collagen (Honch et al., 2012).
- 324 HMP=high-marine protein consumer; HFP=high-freshwater protein consumer; C4=C4 plant consumers;
- 325 C3=C3 plant consumers; Peru=mixed-diet group, Peru, Huari AD 500-900.]
- 326 The bi-plot δ^{13} C Phe vs. δ^{13} C Val values, proposed by Honch et al. (2012) for bone collagen, appears
- 327 effective in tracking the source of the terrestrial resources in the diet of the Pica individuals, being C₃ for SE-
- 328 T3 and C₄ for SD-T24, SI-T32, SF-T4 and SI-T3 (Fig. 7). However, the identification of SI-T74 as a marine
- 329 resource consumer is not straightforward.
- [Figure 7. Plot of δ^{13} C phenylalanine values vs. δ^{13} C values for tendon collagen (this study) and bone collagen (Honch et al., 2012).]
- Previous studies (Choy et al., 2010, Webb et al., 2015) have shown that by using the δ^{13} C values of three
- amino acids the extent of separation of diverse dietary groups may be increased. In our opinion, by including
- a third essential amino acid, leucine, to the previous model of Honch et al. (2012) based on valine and
- 335 phenylalanine δ^{13} C values, the identification of the predominant food source can be facilitated even in a
- 336 mixed marine-terrestrial diet. This is because the source of carbon in the essential (bodily) amino acids
- 337 phenylalanine and leucine is that of dietary proteins, even under conditions of high-lipid and low-protein
- diets. As a result of direct assimilation, δ^{13} C values of phenylalanine and leucine in proteinaceous tissues
- reflect those of the protein component of the diet with minimal isotopic fractionation (Howland et al., 2003,
- 340 Newsome et al., 2011, Newsome et al., 2014). This is not true for the essential amino acid valine. Controlled

- 341 feeding experiments on animals (Newsome et al., 2011, Newsome et al., 2014) have shown that carbon in
- valine may be sourced from the non-protein portion of the diet under conditions of low protein intake. It is
- 343 suggested that gut microflora may synthesize *de novo* essential amino acids such as valine sourcing carbon
- from dietary carbohydrates and/or lipids (Newsome et al., 2011, Newsome et al., 2014). This implies that the
- 345 δ^{13} C value of proteinaceous tissues may be more similar to the δ^{13} C value of the bulk diet rather than
- that of dietary proteins (Newsome et al., 2011), or be strongly correlated with whole diet δ^{13} C value
- 347 (McMahon et al., 2010), or have a non-significant correlation with dietary amino acids (Howland et al.,
- 348 2003), depending on the chosen diet. As a result of the possible processes of assimilation and/or synthesis of
- valine from diet to body tissues, connected to a variety of dietary types, we might expect a wide range of
- valine δ^{13} C values. Given these premises, we hypothesise that by using these three essential amino acids
- 351 $(\delta^{13}C \text{ phenylalanine}, \delta^{13}C \text{ valine}, \delta^{13}C \text{ leucine})$, the reconstruction of palaeodiets of individuals having a
- mixed dietary intake will be more straightforward. The δ^{13} C values of phenylalanine and leucine may be
- useful in tracking and thus separating different dietary groups based on the protein component of their diet,
- and values δ^{13} C values might be useful in tracking the non-protein portion of diet (carbohydrates and/or
- lipids), especially under the condition of protein deficiency.
- [Figure 8. Proposed model for assessing the origin of the predominant dietary intake (terrestrial vs. marine), based on tendon collagen amino acid δ^{13} C values.]
- As shown in Fig. 8, the δ^{13} C phenylalanine values are more negative than the leucine values for consumers
- of mostly marine resources (e.g. SI-T74). Conversely, the leucine δ^{13} C values are more negative than the
- 360 phenylalanine values for consumers of mostly terrestrial resources (e.g. SD-T24, SI-T32, SE-T3). The range
- of these essential amino acid δ^{13} C values is significantly more negative for C₃ terrestrial resource consumers
- 362 (e.g. SE-T3) than for the C₄ terrestrial resource consumers (e.g. SI-T32), reflecting the differential amino
- acid δ^{13} C values of the two plant groups (Fogel and Tuross, 2003). A similar pattern was highlighted by
- Honch et al. (2012) but for collagen δ^{13} C phenylalanine and values. However, value presents more
- variable δ^{13} C values (at least in our dataset), also likely reflecting the effect of the non-protein component of the diet.
- 367 Unfortunately, the most extensive and complete dataset of human bone collagen δ^{13} C values published so far 368 (Honch et al., 2012) does not include leucine δ^{13} C values, making it impossible to test our hypothesis further. 369 The applicability of this Phe/Val/Leu method to various human proteinaceous tissues will need to be tested 370 in future studies.
- 371
- 372 5.4. Dietary reconstruction of selected Pica 8 individuals
- 373 The following interpretation is based on the assumption that, even though a wide spectrum of exotic
- resources was available at Pica (Núñez, 1984, Uribe, 2006), it is unlikely that local individuals consumed
- 375 imported foods exclusively and in a consistent manner. Consumers of exotic foodstuffs (e.g. dried fish)

would be most likely to show a dietary intake made up of local and non-local products, with the proportion of the latter varying through time depending on the nature and outcome of the commerce. Based on this premise, the mobility and migratory histories of the individuals analysed are discussed, although it should be noted that these interpretations are speculative, given the possible mobility of exotic foodstuffs.

Altogether, tendon collagen amino acid δ^{13} C values and FRUITS diet estimates (Fig. 9), based on tendon 380 381 collagen isotope data (δ^{13} C= -7.8‰ to -8.7‰, δ^{15} N= +15.5‰ to +17.5‰), concur in identifying that four 382 individuals (SD-T24, SI-T32, SI-T3, SF-T4) of both sexes and different ages were consuming a terrestrial-383 based diet made of mostly C_4 crops (56-72% of the food intake) and animal meat (9-34%), despite being 384 buried in different sectors (D, F, I) of the Pica 8 cemetery. According to the ecological zone in which the site 385 is geographically located (inland, at mid-altitude, in an arid environment), it may be assumed that this was 386 the 'local' diet consumed at Pica or in nearby communities. This is only partially in line with what has been 387 discussed in previous dietary reconstructions (Santana-Sagredo et al., 2015a, Santana-Sagredo et al., 2015b) 388 undertaken on human remains from the Chilean inland oases of Pica and Quillagua (Late Intermediate 389 Period). Both previous studies have proposed a 'local' dietary intake made up of mixed terrestrial resources, 390 being predominantly composed of maize, and marine foodstuffs. Archaeo-botanical and -faunal remains 391 recovered from Pica-Tarapacá sites of the Late Intermediate Period include C_4 crops such as maize (Zea 392 mays), rodents (Chinchilla sp., Cavia porcellus, Lagidium viscacia), and camelids. The marine resources had 393 to be imported from the Pacific coast and included fish (Trachurus symmetricus, Cilus gilberti), seabirds, marine mammals, molluscs and crustaceans (Mulinia sp., Oliva peruviana, Choromytilus chorus, Argopecten 394 395 purpuratus) (García and Uribe, 2012, Uribe, 2006).

396 Although C_3 cultigens (e.g. beans, potato, squashes, quinoa, pepper) were cultivated and C_3 fruits were 397 gathered (Prosopis spp.) in Pica-Tarapacá communities of the Late Intermediate Period (García and Uribe, 398 2012), these plant products were only consumed by a small number of individuals buried at Pica (4 out of 399 35) (Santana-Sagredo et al., 2015a, Santana-Sagredo et al., 2017), at least based on the skeletons investigated 400 so far. This is surprising as the legume tree Prosopis spp. is a nitrogen-fixing plant that grows well in the arid 401 climate and saline soils of the *Pampa del Tamarugal*, being able to rely solely on groundwater (Fritz et al., 1981, Mooney et al., 1980). Based on the paucity of consumption of C_3 plants at Pica 8, it could be implied 402 403 that these cultigens were less frequently cultivated in this oasis (being more easily cultivated at higher 404 altitude in the Precordillera and Altiplano, where conditions were wetter and colder), or that they were 405 accessed only by a selected group of people within the community, or alternatively that their cultivation was 406 for the purpose of trade (or any of these factors to a greater or lesser extent).

407 [Figure 9. For each individual, from top to bottom: Calorie contribution of each food group to the diet, 408 estimated via Bayesian stable isotope mixing model FRUITS, based on bulk δ^{13} C and δ^{15} N values of tendon 409 collagen (this study) and bone collagen (Santana-Sagredo et al., 2015a). (Asterisk indicates FRUITS model

410 with 'maize fertilised with guano' replacing 'C₄ plants' food group); Amino acid δ^{13} C values and calculated

411 δ^{13} C mass balance (MB) values for tendon collagen; Keratin δ^{13} C values of non-essential and essential amino 412 acids, and calculated δ^{13} C mass balance (MB) values, along the hair fibre.]

413 Individuals SI-T3 and SI-T32 present a comparable set of tendon collagen isotope values (respectively, δ^{13} C = -8.7% and -8.5%, $\delta^{15}N = +15.5\%$ and +16.1%, $\Delta^{13}C_{Val-Phe} = -1.7$ and -0.9, $\delta^{13}C$ Phe = -16.4% and -1.5%414 17.1‰, δ^{13} C Val= -18.1‰ and -18.0‰, δ^{13} C Leu= -18.1‰ and -18.5‰, δ^{13} C mass balance= -9.1‰ and -415 416 9.0‰, Table 2 and Fig. 9) that, in accordance with FRUITS diet estimates, indicate the predominant 417 consumption of C_4 plant products (56-69%), complemented with meat from terrestrial (14-34%) and marine 418 (3-9%) animals (Fig. 9). The less negative δ^{13} C values of both essential and non-essential amino acids (Fig. 419 9), compared to those of marine (SI-T74) and C₃ (SE-T3) resource consumers, show that the adults SI-T3 420 and SI-T32 gleaned the majority of their proteins, carbohydrates and lipids from crops and terrestrial animal 421 meat of C₄ origin. The keratin amino acid δ^{13} C values measured in the hair of SI-T3 and SI-T32 are also very similar (respectively, δ^{13} C Phe= -15.1% to -17.0% and -14.6% to -16.6%, δ^{13} C Leu= -16.3% to -18.5% 422 and -16.0% to -17.7%, δ^{13} C Val= -13.6% to -16.3% and -13.8% to -15.9%, δ^{13} C mass balance= -8.6% 423 to -11.4% and -8.6% to -10.1%, Appendix C and Fig. 9). The limited range of both essential ($\leq 3\%$) and 424 425 non-essential (<5‰) amino acid δ^{13} C values confirms that all three macronutrients (proteins, carbohydrates, 426 lipids) were predominantly retrieved from the same food source, such as C_4 resources. Given that the male 427 SI-T3 and female SI-T32 had a consistent dietary intake typical for mid-altitude (1,300 masl) inland populations (based on local ecology), it may be speculated that these individuals were resident in the locality 428 of Pica, or moved across similar eco-zones, throughout approximately the last year of their lives. However, 429 430 FRUITS diet estimates (Tables 3-4 and Fig. 9), based on rib collagen isotope values ($\delta^{13}C = -10.4\%$, $\delta^{15}N =$ +21.0‰) reported by Santana-Sagredo et al. (2015a), show that, during the previous 3 to 5 years, SI-T3 431 432 relied significantly on marine resources (22-38%), which contributed to 41-65% of his protein intake, and C₄ 433 crops (40-48%). This dissimilarity between bone and tendon isotope values suggests that SI-T3 might have 434 recently entered the inland community after having lived in a location with full access to marine resources 435 (e.g. the Pacific coast and coastal valleys) or, less likely, that exotic foods were available at Pica (or 436 preferred by this individual) only during this period of time.

The tendon collagen isotope values of the adults SD-T24 and SF-T4 (respectively, $\delta^{13}C = -8.2\%$ and -7.8%, 437 δ^{15} N=+16.2‰ and +17.5‰, Δ^{13} C_{Val-Phe}=-0.8 and 0.0, δ^{13} C Phe=-17.2‰ and -16.8‰, δ^{13} C Val=-18.0‰ 438 and -16.8%, δ^{13} C Leu= -18.5% and -18.5%, δ^{13} C mass balance= -8.9% and -8.5%, Table 2 and Fig. 9) 439 440 are broadly similar to those of the two aforementioned C₄ terrestrial resource consumers (SI-T3, SI-T32). 441 However, the keratin amino acid δ^{13} C values (Appendix C and Fig. 9) hint at a more diverse and complex dietary intake. The range of amino acid δ^{13} C values measured in the hair of SF-T4 and SD-T24 is extensive, 442 443 being as high as ~4-5‰ for phenylalanine and leucine, and ~8-9‰ for alanine, proline, aspartic acid, glutamic acid and arginine (for SD-T24). This, combined with the fact that along the hair fibres of SD-T24 444 445 and SF-T4 the δ^{13} C values of essential and non-essential amino acids change in a broadly synchronous 446 pattern (Fig. 9), suggests that all the macronutrients (proteins, lipids, carbohydrates) were sourced from a 447 variety of different foods that changed in proportion and quality over time. For instance, it may be inferred

- that several months prior to death (4-5 cm hair segment), the young male SD-T24 had a significant intake of
- 449 C₃ foods, temporarily ceasing or lowering his consumption of C₄ resources, since the serine δ^{13} C value drops
- 450 to -8.7%, alanine to -15.5%, proline to -15.9%, arginine to -15.6%, aspartic acid to -13.5%, and glutamic
- 451 acid to -11.6‰ (Fig. 9). FRUITS dietary estimates (Fig. 9), calculated based upon rib collagen isotope
- 452 compositions published by Santana-Sagredo et al. (2015a), indicate that SD-T24 had a diet made up of
- 453 predominantly C_4 crops (50-65%) complemented by terrestrial animal meat (18-38%) and C_3 plant products
- 454 (10-13%) for at least several years before his death.
- Although the adults SF-T4 and SD-T24 maintained a high intake of C₄ resources during approximately the
- last 10 months and 3 to 5 years of life, respectively, they consumed additional and diverse foods (e.g. C₃ and,
- to a lesser extent, marine resources) concentrated especially, but not only, during certain months. A potential
- 458 explanation is that these individuals, due to their social, economic or political status had access to a vast
- 459 array of resources or had control over the commercial traffic in the community. An alternative explanation is
- that SF-T4 (female) and SD-T24 (male) had a high degree of mobility across different eco-zones from which
- they derived their foods.
- 462 According to tendon collagen isotope data (δ^{13} C = -18.3‰, δ^{15} N= +12.2‰, Δ^{13} C_{Val-Phe}= -1.7, δ^{13} C Phe= -
- 463 25.9‰, δ^{13} C Val= -27.6‰, δ^{13} C Leu= -28.3‰, Table 2 and Fig. 9), the young woman SE-T3 consumed
- 464 predominantly C₃ terrestrial resources during the final months of her life. The very negative δ^{13} C values of
- both essential and non-essential amino acids (Fig. 9) indicate that she derived proteins, as well as
- 466 carbohydrates and lipids, from C₃ foods. FRUITS dietary estimates (Fig. 9) suggest that this young adult was
- 467 retrieving these three macronutrients mostly from plant products (77-79%) and also from some animal meat
- 468 (~11%). Based on the comparison of tendon and rib collagen isotope data (Table 2), her dietary habits were
- 469 broadly consistent at least during the last 3 to 5 years of life, although the increase in the δ^{15} N value, from
- +10.9% in bone to +12.2% in tendon collagen, may suggest a greater consumption of meat from terrestrial
- 471 animals or, more likely, that she suffered an injury several months before death (D'Ortenzio et al., 2015,
- 472 Fuller et al., 2005, Reitsema, 2013), since instances of bone fractures have been identified.
- 473 The extensive ranges of keratin δ^{13} C values (Appendix C and Fig. 9), measured in both essential (e.g. 6.5%)
- for valine and 5.8‰ for phenylalanine and leucine) and non-essential (e.g. 8‰ for serine, 7.2‰ for proline,
- 475 7‰ for arginine) amino acids, suggest that the SE-T3 female retrieved dietary proteins, carbohydrates and
- 476 lipids from different food sources: C_3 and C_4 . However, the variations among the essential and non-essential
- 477 amino acid δ^{13} C values are broadly synchronous implying that one food source was swapped for the other
- 478 through time and that, if meat was consumed by SE-T3, these animals were fed on the same plant type (C_3 or
- 479 C₄) as that of the plant products directly consumed by this woman. Following the changes in amino acid δ^{13} C
- 480 values along her hair fibre (Fig. 9), it can be inferred that SE-T3's diet was made up of mostly C_3 resources
- 481 about 10 months before death (δ^{13} C Phe= -21.2‰, δ^{13} C Leu= -23.1‰, δ^{13} C Val= -22.2‰), but the
- 482 proportion of these foods declined gradually through time, being replaced by C₄ resources. However, during
- 483 a specific month (4-5 cm hair segment) the source of the carbohydrates was predominantly of C₄ origin (i.e.

- less negative alanine, proline and glutamic acid δ^{13} C values). Closer to the time of death, her diet was mostly composed of C₄ foods (δ^{13} C Phe= -15.8‰, δ^{13} C Leu= -17.5‰, δ^{13} C Val= -15.7‰). The onset of a new diet (i.e. rapid and drastic change) would have likely generated a sharp change in δ^{13} C values, followed by a subsequent gradual adjustment to the new diet, based on previous studies (Ayliffe et al., 2004, O'Connell and Hedges, 1999). However, this is not observed here, since the signal in SE-T3's hair is most likely the result of a gradual change in diet. Nevertheless, the fact that only 10 cm of hair has been analysed makes it possible that the change in diet happened before the time period investigated, and that the gradual change visible here
- 491 (Fig. 9) is, in reality, the isotope equilibration from a C_3 -based diet to the new C_4 -based diet.
- 492 It remains to be ascertained whether the female SE-T3 was experiencing a gradual transition towards C₄ 493 resources as a result of moving from a place where C₃ foods were usually produced/collected and consumed, 494 such as the Precordillera and Altiplano, to the locality of Pica, where maize was more commonly grown and 495 eaten (Santana-Sagredo et al., 2015a, Uribe, 2006). The fact that tendon and rib collagens present C₃ isotope 496 signals might suggest that this young woman lived in another community, plausibly at a higher altitude, 497 which she left several months before death. The highland origin for this individual has already been proposed by Santana-Sagredo et al. (2015a) based on the enamel δ^{18} O value (-11.3‰) measured in the 3rd molar of 498 499 SE-T3. Furthermore, it cannot be excluded that this mobility pattern, spanning from the altiplano to 500 intermediate elevations, might be linked to camelid herding practices or long-distance trade, as women were involved in these activities (Pomeroy, 2013). An alternative explanation, though less likely, is that SE-T3 501 502 was a local member of the Pica communities that had access to imported C_3 foods from the highlands, until 503 approximately a year before death but, during the final year of life, she was only able to access C₄ resources 504 such as maize, as a result of a change in her socio-economic status.
- Given that the hair keratin recorded a transition from a C_3 to a C_4 food source, while the tendon tissues reflect a completely C_3 dietary intake likely occurring during a previous period of time, it appears that the tendon collagen is slower in turning over and adjusting to the new isotope signal than previously hypothesised (Babraj et al., 2005). A recent study by Heinemeier et al. (2013) found that the renewal of tendon tissues is quite variable within individuals, and that the turnover of the tendon core is very limited/slow. With respect to the present study, it may be speculated that the tendon of SE-T3 had poor regenerative capacity, or that only the core of the tendon has survived.
- Based on tendon collagen isotope values ($\delta^{15}N$ =+25.3‰, $\delta^{13}C$ = -10.8‰, $\Delta^{13}C$ _{Val-Phe}=2.0, $\delta^{13}C$ Phe=-512 20.6‰, δ^{13} C Val= -18.6‰, δ^{13} C Leu= -19.9‰, Table 2 and Fig. 9), it appears that towards the end of her 513 life the young woman SI-T74 relied predominantly on marine foodstuffs, which represented 41-51% of the 514 food she consumed and contributed to 71-81% of her protein intake (Tables 3-4). The value δ^{13} C value of 515 516 this individual is in line with those of high-marine protein consumers reported by Honch et al. (2012), but the 517 δ^{13} C value of phenylalanine is slightly less negative, thus suggesting an additional intake of C₄ resources. 518 Dietary estimates generated by FRUITS models confirm the concurrent consumption of some C_4 crops (36-519 42%), such as maize. According to tendon and rib collagen isotope compositions (Table 2), the dependence

- 520 on marine resources was likely consistent over the course of the last 3 to 5 years of SI-T74's life. The
- slightly higher (+2.4‰) tendon δ^{15} N value (compared to that of bone) may have been induced by the
- 522 consumption of manured maize and/or high trophic level marine animals, such as sea lions, or alternatively
- 523 by the occurrence of a disease. Bone lesions have been identified on SI-T74's rib, which could have been
- 524 generated by tuberculosis infection, although there are many other potential causes (Roberts et al., 1994;
- 525 Santos and Roberts 2006).
- 526 The range of δ^{13} C values, measured along SI-T74's hair fibre, is greater than 4‰ for some essential amino 527 acids (phenylalanine, isoleucine, lysine) and greater than 6‰ for some non-essential amino acids (alanine, 528 glutamic acid, proline) (Appendix C and Fig. 9). This suggests that this young woman was deriving her 529 dietary proteins from a sole source of food, such as marine resources (based on the range of essential amino 530 acids), and that carbohydrates and lipids were also derived from another source, such as C_4 (based on the range of non-essential amino acids). The variation in amino acid δ^{13} C values along the hair fibre (Fig. 9) 531 532 shows that the proportion of marine and C₄ foods changed in the diet of SI-T74 through time. Assuming a 533 growth rate of approximately 1 cm per month (Saitoh et al., 1969) for scalp hair in the anagen phase, 534 between about 10 to 8 months before her death this woman consumed predominantly marine resources $(\Delta^{13}C_{\text{Val-Phe}} = 3.4 \text{ to } 4.3, \delta^{13}C \text{ Phe} = -21.3\% \text{ to } -19.6\%, \delta^{13}C \text{ Val} = -17.3\% \text{ to } -16.2\%, \delta^{13}C \text{ Leu} = -20.1\% \text{ to } -10.2\%$ 535 -19.1%), while during the subsequent month (6-7 cm hair segment) she notably increased her intake of C₄ 536 crops ($\Delta^{13}C_{Val-Phe} = 1.7$). This generated a shift of several per mill towards less negative $\delta^{13}C$ values in both 537 essential (δ^{13} C Phe= from -21.0‰ to -17.4‰, δ^{13} C Val= from -17.3‰ to -15.7‰, δ^{13} C Leu= from -19.5‰ 538 539 to -17.6‰, δ^{13} C Lys= from -13.7‰ to -11.8‰, δ^{13} C IIe= from -13.2‰ to -11.2‰) and non-essential amino acids (δ^{13} C Ala= from -14.1‰ to -8.0‰, δ^{13} C Pro= from -14.6‰ to -8.4‰, δ^{13} C Glx= from -8.7‰ to -540 541 3.8%). Subsequently, SI-T74 decreased her intake of C4 resources, relying mostly on marine resources 542 $(\Delta^{13}C_{Val-Phe} = 3.0)$ for a couple of months (4 to 6 cm hair segments). In the final period of her life, she then 543 returned to a diet rich in C₄ resources ($\Delta^{13}C_{Val-Phe} = 1.0$ to 1.8) and the $\delta^{13}C$ values of the aforementioned 544 amino acids again became less negative (Fig. 9). Considering that the human body breaks down and recycles 545 old proteins, as well as synthesizes new ones (O'Connell and Hedges, 1999), and that maize (the most likely 546 consumed C_4 resource) is low in protein (although this may be slightly increased by the use of fertilisers; Keeney, 1970), it is reasonable to assume that her marine resource intake could be in reality lower than 547 548 previously discussed. In other words, the marine isotope signal recorded by the keratin amino acids may 549 derive from recycled proteins formed during a previous dietary phase characterized by high marine protein 550 consumption, and not from a recent intake of marine resources.
- 551 Given that Pica is situated ~80 km from the sea, it is difficult to explain this consistent and significant
- 552 consumption of marine resources throughout the years. Although some dried fish, fish bones and molluscs
- 553 have been found at the cemetery (Núñez, 1984), it is unclear how available they would have been for regular
- 554 consumption at such a distance from their source. The exchange network would need to have been developed
- 555 enough such that marine resources were available routinely and in high quantities for a few selected
- individuals at Pica; it is therefore possible that SI-T74 had access to these foods, perhaps due to political,

cultural or socio-economic reasons. An alternative explanation is that she was resident on the coast or in the coastal valleys of the Atacama Desert where marine foods and maize were both local, and only recently before death she migrated to the locality of Pica where she was eventually buried, or was travelling inland passing nearby these oases.

561

562 5. Conclusions

563 The individuals buried at Pica 8 present heterogeneous nutritional histories, both individually and collectively. At the time of their deaths, all six adults (SD-T24, SI-T32, SF-T4, SI-T3, SE-T3, SI-T74) were 564 565 characterised by a terrestrial-C₄ diet, which is in line with what may have been the most easily accessible foodstuffs in the locality of Pica: maize and terrestrial animal meat. Among these individuals, SI-T3 and SE-566 567 T3 experienced a shift in their dietary intake several months to a year before death, possibly as a result of 568 relocation to mid-altitude communities. The original dietary habits of SE-T3 and SI-T3 were respectively 569 characteristic of the highlands (C_3 plants) and the coast and coastal valleys (marine resources). Only one 570 individual (SI-T74) might have been resident on the coast or in the coastal valleys of the Atacama Desert at 571 least over the course of the last 3 to 5 years of her life, based on a consistent intake of marine resources. 572 However, shortly before her death SI-T74 shifted to a more terrestrial diet when she possibly migrated to, or 573 was travelling close to, the locality of Pica.

574 Based on the dietary and mobility reconstruction of this subset of individuals, during the Late Intermediate 575 Period (~1,050-500 BP), the oasis of Pica appears to have been an economically and commercially dynamic environment, which attracted people from distant regions. The individuals buried at Pica 8 may have been 576 577 actively involved in the trade of exotic objects and foodstuffs, acting as traders along the caravan routes, or 578 have belonged to the elite group who managed exchanges (Briones et al., 2005, Núñez, 1984, Pacheco and 579 Retamal, 2017, Pomeroy, 2013). In particular, the individuals from Sector I (SI-T3, SI-T74) may have been 580 relocated from, or linked to, communities located on the Pacific coast in order to support mutual 581 redistribution of resources between different eco-zones (Santana-Sagredo et al., 2015a, Uribe, 2006).

To our knowledge, this is the first study that has analysed collagen amino acid δ^{13} C values in tendon samples from archaeological human remains, and this research shows that tendon may be a favourable substitute for bone in palaeodietary reconstructions, providing it is preserved in the archaeological context. Given that collagen is more abundant in tendon than in bone and its rate of turnover is faster (Babraj et al., 2005, Kjaer et al., 2005), the dietary information reconstructed based on tendon stable isotope compositions presents a higher temporal resolution and may be more fine-grained than that of bone collagen, since the original isotope signal is averaged over a shorter period of time, i.e. several months/a year instead of years/decades.

6. Acknowledgements

We would like to express our sincere gratitude to the Department of Anthropology of the Universidad de Chile for granting access to the Pica 8 collection.

A.M. was supported by La Trobe University PhD scholarships. A.P. was supported by the Advanced Human Capital Program of the National Commission for Scientific and Technological Research (CONICYT) of Chile. This work was supported by La Trobe University HUSS-IRGS (#2015-2-HDR-0021) and Australian Research Council Future Fellowship FT0992258.

References

Ajata López, R.C., 2013. Distribucion y emplazamiento de sitios arqueologicos en Pica-Tarapacá, norte de Chile. Un acercamiento a través de los Sistemas de Información Geográfica., El uso de Sistemas de Información Geográfica (SIG) en arqueología sudamericana BAR International Series 2497, 139-155. Ambrose, S.H., 1990. Preparation and characterization of bone and tooth collagen for isotopic analysis, Journal of Archaeological Science 17, 431-451.

Ambrose, S.H., Norr, L., 1993. Experimental evidence for the relationship of the carbon isotope ratios of whole diet and dietary protein to those of bone collagen and carbonate, in: Lambert, J.B., Grupe, G. (Eds.), Prehistoric human bone, Springer-Verlag, Berlin.

Andrade, P., Fernandes, R., Codjambassis, K., Urrea, J., Olguin, L., Rebolledo, S., Lira, F., Aravena, C., Berríos, M., 2015. Subsistence continuity linked to consumption of marine protein in the Formative Period in the interfluvic coast of northern Chile: Re-assessing contacts with agropastoral groups from highlands, Radiocarbon 57, 679-688.

Aufderheide, A.C., Kelley, M.A., Rivera, M., Gray, L., Tieszen, L.L., Iversen, E., Krouse, H.R., Carevic, A., 1994. Contributions of chemical dietary reconstruction to the assessment of adaptation by ancient highland immigrants (Alto Ramirez) to coastal conditions at Pisagua, north Chile, Journal of Archaeological Science 21, 515-524.

Ayliffe, L.K., Cerling, T.E., Robinson, T., West, A.G., Sponheimer, M., Passey, B.H., Hammer, J., Roeder, B., Dearing, M.D., Ehleringer, J., 2004. Turnover of carbon isotopes in tail hair and breath CO₂ of horses fed an isotopically varied diet, Oecologia 139, 11-22.

Babraj, J.A., Cuthbertson, D.J., Smith, K., Langberg, H., Miller, B., Krogsgaard, M.R., Kjaer, M., Rennie,M.J., 2005. Collagen synthesis in human musculoskeletal tissues and skin, American Journal of Physiology-Endocrinology and Metabolism 289, E864-E869.

Basha, W.A., Chamberlain, A.T., Zaki, M.E., Kandeel, W.A., Fares, N.H., 2016. Diet reconstruction through stable isotope analysis of ancient mummified soft tissues from Kulubnarti (Sudanese Nubia), Journal of Archaeological Science: Reports 5, 71-79.

Briones, L., Núñez, L., Standen, V.G., 2005. Geoglifos y tráfico prehispánico de caravanas de llamas en el desierto de Atacama (norte de Chile), Chungara Revista de Antropología Chilena 37, 195-223.

Burress, E.D., Duarte, A., Gangloff, M.M., Siefferman, L., 2013. Isotopic trophic guild structure of a diverse subtropical South American fish community, Ecology of Freshwater Fish 22, 66-72.

Choy, K., Smith, C.I., Fuller, B.T., Richards, M.P., 2010. Investigation of amino acid δ^{13} C signatures in bone collagen to reconstruct human palaeodiets using liquid chromatography–isotope ratio mass spectrometry, Geochimica et Cosmochimica Acta 74, 6093-6111.

Codron, J., Codron, D., Lee-Thorp, J.A., Sponheimer, M., Bond, W.J., de Ruiter, D., Grant, R., 2005. Taxonomic, anatomical, and spatio-temporal variations in the stable carbon and nitrogen isotopic compositions of plants from an African savanna, Journal of Archaeological Science 32, 1757-1772.

Copley, M.S., Jim, S., Jones, V., Rose, P., Clapham, A., Edwards, D.N., Horton, M., Rowley-Conwy, P., Evershed, R.P., 2004. Short-and long-term foraging and foddering strategies of domesticated animals from Qasr Ibrim, Egypt, Journal of Archaeological Science 31, 1273-1286.

Corr, L.T., Sealy, J.C., Horton, M.C., Evershed, R.P., 2005. A novel marine dietary indicator utilising compound-specific bone collagen amino acid δ^{13} C values of ancient humans, Journal of Archaeological Science 32, 321-330.

Covey, R.A., 2000. Inka administration of the far south coast of Peru, Latin American Antiquity 11, 119-138.

D'Ortenzio, L., Brickley, M., Schwarcz, H., Prowse, T., 2015. You are not what you eat during physiological stress: Isotopic evaluation of human hair, American Journal of Physical Anthropology 157, 374-388.

Denevan, M.W., 2001. Crops, tools, and soft technologies, Cultivated landscapes of Native Amazonia and the Andes, Oxford University Press, New York, pp. 27-49.

DeNiro, M.J., 1985. Postmortem preservation and alteration of in vivo bone collagen isotope ratios in relation to palaeodietary reconstruction, Nature 317, 806-809.

DeNiro, M.J., Hastorf, C.A., 1985. Alteration of ¹⁵N/¹⁴N and ¹³C/¹²C ratios of plant matter during the initial stages of diagenesis: Studies utilizing archaeological specimens from Peru, Geochimica et Cosmochimica Acta 49, 97-115.

DeNiro, M.J., 1988. Marine food sources for prehistoric coastal Peruvian camelids: Isotopic evidence and implications, in: Wing, E.S., Wheeler, J.C. (Eds.), Economic Prehistory of the Central Andes, BAR International Series, Oxford, pp. 119-129.

Eastoe, J.E., 1955. The amino acid composition of mammalian collagen and gelatin, Biochemical Journal 61, 589-600.

Falabella, F., Planella, M.T., Aspillaga, E., Sanhueza, L., Tykot, R.H., 2007. Dieta en sociedades alfareras de Chile central: Aporte de análisis de isótopos estables, Chungara Revista de Antropología Chilena 39, 5-27. Fernandes, R., Nadeau, M.-J., Grootes, P.M., 2012. Macronutrient-based model for dietary carbon routing in bone collagen and bioapatite, Archaeological and Anthropological Sciences 4, 291-301.

Fernandes, R., Millard, A.R., Brabec, M., Nadeau, M.-J., Grootes, P., 2014. Food reconstruction using isotopic transferred signals (FRUITS): A Bayesian model for diet reconstruction, PLoS ONE 9, e87436. Fernandes, R., 2015. A simple(r) model to predict the source of dietary carbon in individual consumers, Archaeometry 58, 500-512.

Fernandes, R., Grootes, P., Nadeau, M.-J., Nehlich, O., 2015. Quantitative diet reconstruction of a Neolithic population using a Bayesian mixing model (FRUITS): The case study of Ostorf (Germany), American Journal of Physical Anthropology 158, 325-340.

Finucane, B., Agurto, P.M., Isbell, W.H., 2006. Human and animal diet at Conchopata, Peru: Stable isotope evidence for maize agriculture and animal management practices during the Middle Horizon, Journal of Archaeological Science 33, 1766-1776.

Finucane, B.C., 2007. Mummies, maize, and manure: Multi-tissue stable isotope analysis of late prehistoric human remains from the Ayacucho Valley, Perú, Journal of Archaeological Science 34, 2115-2124.Fogel, M.L., Tuross, N., 2003. Extending the limits of paleodietary studies of humans with compound

specific carbon isotope analysis of amino acids, Journal of Archaeological Science 30, 535-545.

Fritz, P., Suzuki, O., Silva, C., Salati, E., 1981. Isotope hydrology of groundwaters in the Pampa del Tamarugal, Chile, Journal of Hydrology 53, 161-184.

Fuller, B.T., Fuller, J.L., Sage, N.E., Harris, D.A., O'Connell, T.C., Hedges, R.E.M., 2005. Nitrogen balance and δ^{15} N: Why you're not what you eat during nutritional stress, Rapid Communications in Mass Spectrometry 19, 2497-2506.

García, M., Uribe, M., 2012. Contextos de uso de las plantas vinculadas al Complejo Pica Tarapacá, Andes Centro-Sur: Arqueobotánica y agricultura en el período Intermedio Tardío (ca. 1250-1450 DC), Estudios Atacameños 44, 107-122.

Gil, A.F., Neme, G.A., Tykot, R.H., 2011. Stable isotopes and human diet in central western Argentina, Journal of Archaeological Science 38, 1395-1404.

Hare, E.P., Fogel, M.L., Stafford, T.W., Mitchell, A.D., Hoering, T.C., 1991. The isotopic composition of carbon and nitrogen in individual amino acids isolated from modern and fossil proteins, Journal of Archaeological Science 18, 277-292.

Hedges, R.E.M., Clement, J.G., Thomas, C.D.L., O'Connell, T.C., 2007. Collagen turnover in the adult femoral mid-shaft: Modeled from anthropogenic radiocarbon tracer measurements, American Journal of Physical Anthropology 133, 808-816.

Heinemeier, K.M., Schjerling, P., Heinemeier, J., Magnusson, S.P., Kjaer, M., 2013. Lack of tissue renewal in human adult Achilles tendon is revealed by nuclear bomb ¹⁴C, The FASEB Journal 27, 2074-2079. Hobson, K.A., Clark, R.G., 1992. Assessing avian diets using stable isotopes II: Factors influencing diet-tissue fractionation, The Condor 94, 189-197.

Hobson, K.A., Schell, D.M., Renouf, D., Noseworthy, E., 1996. Stable carbon and nitrogen isotopic fractionation between diet and tissues of captive seals: Implications for dietary reconstructions involving marine mammals, Canadian Journal of Fisheries and Aquatic Sciences 53, 528-533.

Hoeinghaus, D.J., Vieira, J.P., Costa, C.S., Bemvenuti, C.E., Winemiller, K.O., Garcia, A.M., 2011. Estuary hydrogeomorphology affects carbon sources supporting aquatic consumers within and among ecological guilds, Hydrobiologia 673, 79-92.

Honch, N.V., McCullagh, J.S.O., Hedges, R.E.M., 2012. Variation of bone collagen amino acid δ^{13} C values in archaeological humans and fauna with different dietary regimes: Developing frameworks of dietary discrimination, American Journal of Physical Anthropology 148, 495-511.

Howland, M.R., Corr, L.T., Young, S.M.M., Jones, V., Jim, S., Van Der Merwe, N.J., Mitchell, A.D., Evershed, R.P., 2003. Expression of the dietary isotope signal in the compound-specific δ^{13} C values of pig bone lipids and amino acids, International Journal of Osteoarchaeology 13, 54-65.

Hückstädt, L.A., Rojas, C.P., Antezana, T., 2007. Stable isotope analysis reveals pelagic foraging by the Southern sea lion in central Chile, Journal of Experimental Marine Biology and Ecology 347, 123-133. Iacumin, P., Bocherens, H., Chaix, L., Marioth, A., 1998. Stable carbon and nitrogen isotopes as dietary indicators of ancient Nubian populations (Northern Sudan), Journal of Archaeological Science 25, 293-301. Jayne, R.S., Pollyea, R.M., Dodd, J.P., Olson, E.J., Swanson, S.K., 2016. Spatial and temporal constraints on regional-scale groundwater flow in the Pampa del Tamarugal Basin, Atacama Desert, Chile, Hydrogeology Journal 24, 1921-1937.

Jim, S., Jones, V., Ambrose, S.H., Evershed, R.P., 2006. Quantifying dietary macronutrient sources of carbon for bone collagen biosynthesis using natural abundance stable carbon isotope analysis, British Journal of Nutrition 95, 1055-1062.

Jørkov, M.L.S., Heinemeier, J., Lynnerup, N., 2009. The petrous bone-A new sampling site for identifying early dietary patterns in stable isotopic studies, American Journal of Physical Anthropology 138, 199-209. Julien, C.J., 1985. Guano and resource control in sixteenth-century Arequipa, in: Masuda, S., Shimada, I., Morris, C. (Eds.), Andean ecology and civilization: An interdisciplinary perspective on Andean ecological complementarity, University of Tokyo Press, Tokyo, pp. 185-231.

Kannus, P., 2000. Structure of the tendon connective tissue, Scandinavian Journal of Medicine & Science in Sports 10, 312-320.

Keeney, D.R., 1970. Protein and amino acid composition of maize grain as influenced by variety and fertility, Journal of the Science of Food and Agriculture 21, 182-184.

Keil, R.G., Fogel, M.L., 2001. Reworking of amino acid in marine sediments: Stable carbon isotopic composition of amino acids in sediments along the Washington coast, Limnology and Oceanography 46, 14-23.

Kelly, J.F., 2000. Stable isotopes of carbon and nitrogen in the study of avian and mammalian trophic ecology, Canadian Journal of Zoology 78, 1-27.

King, C.L., Millard, A.R., Gröcke, D.R., Standen, V.G., Arriaza, B.T., Halcrow, S.E., 2018. Marine resource reliance in the human populations of the Atacama Desert, northern Chile – A view from prehistory, Quaternary Science Reviews 182, 163-174.

Kjaer, M., Langberg, H., Miller, B.F., Boushel, R., Crameri, R., Koskinen, S., Heinemeier, K., Olesen, J.L., Døssing, S., Hansen, M., Pedersen, S.G., Rennie, M.J., Magnusson, P., 2005. Metabolic activity and collagen turnover in human tendon in response to physical activity, Journal of Musculoskeletal and Neuronal Interactions 5, 41-52.

Knapp, A.B., van Dommelen, P., 2008. Past practices: Rethinking individuals and agents in archaeology, Cambridge Archaeological Journal 18, 15-34.

Lamb, A.L., 2015. Stable isotope analysis of soft tissues from mummified human remains, Environmental Archaeology 21, 271-284.

Lemp, C., Rodríguez, M., Retamal, R., Aspillaga, E., 2008. Arqueología del depósito: Manejo integral de las colecciones bioantropológicas en el Departamento de Antropología de la Universidad de Chile, Conserva 12, 69-96.

Lynnerup, N., 2007. Mummies, American Journal of Physical Anthropology 134, 162-190.

Makarewicz, C.A., Sealy, J., 2015. Dietary reconstruction, mobility, and the analysis of ancient skeletal tissues: Expanding the prospects of stable isotope research in archaeology, Journal of Archaeological Science 56, 146-158.

Marcus, J., Sommer, J.D., Glew, C.P., 1999. Fish and mammals in the economy of an ancient Peruvian kingdom, Proceedings of the National Academy of Sciences 96, 6564-6570.

Marino, B.D., McElroy, M.B., 1991. Isotopic composition of atmospheric CO₂ inferred from carbon in C4 plant cellulose, Nature 349, 127-131.

Mateo, M.A., Serrano, O., Serrano, L., Michener, R.H., 2008. Effects of sample preparation on stable isotope ratios of carbon and nitrogen in marine invertebrates: Implications for food web studies using stable isotopes, Oecologia 157, 105-115.

McMahon, K.W., Fogel, M.L., Elsdon, T.S., Thorrold, S.R., 2010. Carbon isotope fractionation of amino acids in fish muscle reflects biosynthesis and isotopic routing from dietary protein, Journal of Animal Ecology 79, 1132–1141.

Miller, B.F., Olesen, J.L., Hansen, M., Døssing, S., Crameri, R.M., Welling, R.J., Langberg, H., Flyvbjerg, A., Kjaer, M., Babraj, J.A., Smith, K., Rennie, M.J., 2005. Coordinated collagen and muscle protein synthesis in human patella tendon and quadriceps muscle after exercise, The Journal of Physiology 567, 1021-1033.

Mooney, H.A., Gulmon, S.L., Rundel, P.W., Ehleringer, J., 1980. Further observations on the water relations of *Prosopis tamarugo* of the northern Atacama desert, Oecologia 44, 177-180.

Mora, A., Arriaza, B.T., Standen, V.G., Valdiosera, C., Salim, A., Smith, C., 2017. High-resolution palaeodietary reconstruction: Amino acid δ^{13} C analysis of keratin from single hairs of mummified human individuals, Quaternary International 436, 96-113.

Neuberger, F.M., Jopp, E., Graw, M., Püschel, K., Grupe, G., 2013. Signs of malnutrition and starvation-Reconstruction of nutritional life histories by serial isotopic analyses of hair, Forensic Science International 226, 22-32.

Newsholme, P., Stenson, L., Sulvucci, M., Sumayao, R., Krause, M., 2011. Amino Acid Metabolism, in: Moo-Young, M. (Ed.), Comprehensive Biotechnology, 2 ed., Elsevier, Amsterdam, pp. 3-14.

Newsome, S.D., Fogel, M.L., Kelly, L., del Rio, C.M., 2011. Contributions of direct incorporation from diet and microbial amino acids to protein synthesis in Nile tilapia, Functional Ecology 25, 1051-1062.

Newsome, S.D., Wolf, N., Peters, J., Fogel, M.L., 2014. Amino acid δ^{13} C analysis shows flexibility in the routing of dietary protein and lipids to the tissue of an omnivore, Integrative and Comparative Biology 54, 890-902.

Núñez, L., 1976. Registro regional de fechas radiocarbónicas del norte de Chile, Estudios Atacameños 4, 69-111.

Núñez, L., Dillehay, T.D., 1979. Movilidad giratoria, armonía social y desarrollo en los Andes meridionales: Patrones de tráfico e interacción económica, Universidad Católica del Norte, Antofagasta.

Núñez, L., 1984. Tráfico de Complementariedad de Recursos entre las Tierras Altas y el Pacífico en el área Centro sur Andina, University of Tokyo, Tokyo.

O'Connell, T.C., Hedges, R.E.M., 1999. Investigations into the effect of diet on modern human hair isotopic values, American Journal of Physical Anthropology 108, 409-425.

O'Connell, T.C., Hedges, R.E.M., Healey, M.A., Simpson, A.H.R.W., 2001. Isotopic comparison of hair, nail and bone: Modern analyses, Journal of Archaeological Science 28, 1247-1255.

Pacheco, A., Retamal, R., 2017. Avoiding war in Tarapacá (Northern Chile) during the Andean Late

Intermediate Period (AD 1000-1450), International Journal of Osteoarchaeology 27, 35-44.

Pestle, W.J., Torres-Rouff, C., Hubbe, M., 2016. Modeling diet in times of change: The case of Quitor, San Pedro de Atacama, Chile, Journal of Archaeological Science: Reports 7, 82-93.

Pestle, W.J., Torres-Rouff, C., Hubbe, M., Smith, E.K., 2017. Eating out or dining in: Modeling diverse dietary strategies in the Middle Period, San Pedro de Atacama, Chile, Archaeological and Anthropological Sciences 9, 1363-1377.

Petruzzelli, B., Roberts, A., Pate, D.F., Santoro, C.M., Maddern, T., Carter, C., Westaway, M.C., 2014. Stable carbon and nitrogen isotopic analysis of skeletal remains from Azapa 71 and Pica-8, Northern Chile: An assessment of human diet and landscape use in the Late Holocene, Journal of the Anthropological Society of South Australia 35, 52-80.

Petzke, K.J., Fuller, B.T., Metges, C.C., 2010. Advances in natural stable isotope ratio analysis of human hair to determine nutritional and metabolic status, Current Opinion in Clinical Nutrition and Metabolic Care 13, 532-540.

Phillips, D.L., Inger, R., Bearhop, S., Jackson, A.L., Moore, J.W., Parnell, A.C., Semmens, B.X., Ward, E.J., 2014. Best practices for use of stable isotope mixing models in food-web studies, Canadian Journal of Zoology 92, 823-835.

Pomeroy, E., 2013. Biomechanical insights into activity and long distance trade in the south-central Andes (AD 500-1450), Journal of Archaeological Science 40, 3129-3140.

Reeds, P.J., 2000. Dispensable and indispensable amino acids for humans, The Journal of Nutrition 130, 1835S-1840S.

Reitsema, L.J., 2013. Beyond diet reconstruction: Stable isotope applications to human physiology, health, and nutrition, American Journal of Human Biology 25, 445-456.

Retamal, R., Pacheco, A., Uribe, M., 2012. Dimorfismo sexual, distribución etaria y longevidad del cementerio Pica 8 (período Intermedio Tardío, 950-1450 DC, Norte Grande de Chile), Estudios Atacameños, 89-106.

Robbins, C.R., Kelly, C.H., 1970. Amino acid composition of human hair, Textile Research Journal 40, 891-896.

Roberts, C., Lucy, D., Manchester, K., 1994. Inflammatory lesions of ribs: An analysis of the Terry Collection, American Journal of Physical Anthropology 95, 169-182.

Saitoh, M., Uzuka, M., Sakamoto, M., Kobori, T., 1969. Rate of hair growth, in: Montagna, W., Dobson, R.L. (Eds.), Advances in Biology of Skin. Volume IX-'Hair Growth', Pergamon Press, Oxford, pp. 183-201. Santana-Sagredo, F., Lee-Thorp, J.A., Schulting, R., Uribe, M., 2015a. Isotopic evidence for divergent diets and mobility patterns in the Atacama Desert, northern Chile, during the Late Intermediate Period (AD 900-1450), American Journal of Physical Anthropology 156, 374-387.

Santana-Sagredo, F., Hubbe, M., Uribe, M., 2015b. Isotopic evidence for marine consumption and mobility in the Atacama Desert (Quillagua, Northern Chile), International Journal of Osteoarchaeology 26, 476–489. Santana-Sagredo, F., Schulting, R., Lee-Thorp, J., Agüero, C., Uribe, M., Lemp, C., 2017. Paired radiocarbon dating on human samples and camelid fibers and textiles from northern Chile: The case of Pica 8 (Tarapacá), Radiocarbon 59, 1195-1213.

Santos, A.L., Roberts, C.A., 2006. Anatomy of a serial killer: Differential diagnosis of tuberculosis based on rib lesions of adult individuals from the Coimbra Identified Skeletal Collection, Portugal, American Journal of Physical Anthropology 130, 38-49.

Schloesser, R.W., Rooker, J.R., Louchuoarn, P., Neilson, J.D., Secord, D.H., 2009. Interdecadal variation in seawater δ^{13} C and δ^{18} O recorded in fish otoliths, Limnology and Oceanography 54, 1665-1668.

Schofield, J.D., Freeman, I.L., Jackson, D.S., 1971. The isolation, and amino acid and carbohydrate composition, of polymeric collagens prepared from various human tissues, Biochemical Journal 124, 467-473.

Sealy, J.C., van der Merwe, N.J., Thorp, J.A.L., Lanham, J.L., 1987. Nitrogen isotopic ecology in southern Africa: Implications for environmental and dietary tracing, Geochimica et Cosmochimica Acta 51, 2707-2717.

Sholto-Douglas, A.D., Field, J.G., James, A.G., van der Merwe, N.J., 1991. ¹³C/¹²C and ¹⁵N/¹⁴N isotope ratios in the Southern Benguela Ecosystem: indicators of food web relationships among different size-classes of

plankton and pelagic fish; differences between fish muscle and bone collagen tissues, Mar. Ecol. Prog. Ser. 78, 23-31.

Smith, C.I., Fuller, B.T., Choy, K., Richards, M.P., 2009. A three-phase liquid chromatographic method for δ^{13} C analysis of amino acids from biological protein hydrolysates using liquid chromatography–isotope ratio mass spectrometry, Analytical Biochemistry 390, 165-172.

Szpak, P., Longstaffe, F.J., Millaire, J.-F., White, C.D., 2012a. Stable isotope biogeochemistry of seabird guano fertilization: Results from growth chamber studies with Maize (*Zea mays*), PLoS ONE 7, e33741. Szpak, P., Millaire, J.-F., White, C.D., Longstaffe, F.J., 2012b. Influence of seabird guano and camelid dung fertilization on the nitrogen isotopic composition of field-grown maize (Zea mays), Journal of Archaeological Science 39, 3721-3740.

Szpak, P., White, C.D., Longstaffe, F.J., Millaire, J.-F., Vásquez Sánchez, V.F., 2013. Carbon and nitrogen isotopic survey of northern Peruvian plants: Baselines for paleodietary and paleoecological studies, PLoS ONE 8, e53763.

Szpak, P., Millaire, J.-F., White, C.D., Longstaffe, F.J., 2014. Small scale camelid husbandry on the north coast of Peru (Virú Valley): Insight from stable isotope analysis, Journal of Anthropological Archaeology 36, 110-129.

Szpak, P., Chicoine, D., Millaire, J.-F., White, C.D., Parry, R., Longstaffe, F.J., 2015. Early Horizon camelid management practices in the Nepeña Valley, north-central coast of Peru, Environmental Archaeology 21, 230-245.

Thornton, E.K., Defrance, S.D., Krigbaum, J., Williams, P.R., 2011. Isotopic evidence for Middle Horizon to 16th century camelid herding in the Osmore Valley, Peru, International Journal of Osteoarchaeology 21, 544-567.

Tieszen, L.L., Boutton, T.W., Tesdahl, K.G., Slade, N.A., 1983. Fractionation and turnover of stable carbon isotopes in animal tissues: Implications for δ^{13} C analysis of diet, Oecologia 57, 32-37.

Tieszen, L.L., Chapman, M., 1992. Carbon and nitrogen isotopic status of the major marine and terrestrial resources in the Atacama Desert of northern Chile, Proceedings of the First World Congress on Mummy Studies, Museo Arqueológico y Etnográfico Tenerife, Santa Cruz de Tenerife, pp. 409–425.

Uribe, M., 2006. Acerca de complejidad, desigualdad social y el complejo cultural Pica-Tarapacá en los Andes Centro-Sur (1000-1450 DC), Estudios Atacameños 31, 91-114.

Uribe, M., Sanhueza, L., Bahamondes, F., 2007. La cerámica prehispánica tardía de Tarapaca, sus valles interiores y costa desértica, Norte de Chile (ca. 900-1.450 D.C.): Una propuesta tipológica y cronológica, Chungara Revista de Antropología Chilena 39, 143-170.

van Der Merwe, N.J., Lee-Thorp, J.A., Raymond, J.S., 1993. Light, stable isotopes and the subsistence base of Formative cultures at Valdivia, Ecuador, in: Lambert, J.B., Grupe, G. (Eds.), Prehistoric human bone: Archaeology at the molecular level Springer-Verlag, Berlin, pp. 63-97.

Vogel, J.C., 1978. Isotopic assessment of the dietary habits of ungulates, South African Journal of Science 74, 298-301.

Wang, J.H.-C., 2006. Mechanobiology of tendon, Journal of Biomechanics 39, 1563-1582.

Warinner, C., Tuross, N., 2010. Brief communication: Tissue isotopic enrichment associated with growth depression in a pig: Implications for archaeology and ecology, American Journal of Physical Anthropology 141, 486-493.

Webb, E.C., Honch, N.V., Dunn, P.J.H., Eriksson, G., Lidén, K., Evershed, R.P., 2015. Compound-specific amino acid isotopic proxies for detecting freshwater resource consumption, Journal of Archaeological Science 63, 104-114.

Webb, E.C., Honch, N.V., Dunn, P.J.H., Linderholm, A., Eriksson, G., Lidén, K., Evershed, R.P., 2016. Compound-specific amino acid isotopic proxies for distinguishing between terrestrial and aquatic resource consumption, Archaeological and Anthropological Sciences, 1-18.

Webb, E.C., Lewis, J., Shain, A., Kastrisianaki-Guyton, E., Honch, N.V., Stewart, A., Miller, B., Tarlton, J., Evershed, R.P., 2017. The influence of varying proportions of terrestrial and marine dietary protein on the stable carbon-isotope compositions of pig tissues from a controlled feeding experiment, STAR: Science & Technology of Archaeological Research 3, 36-52.

White, C.D., Schwarcz, H.P., 1994. Temporal trends in stable isotopes for Nubian mummy tissues, American Journal of Physical Anthropology 93, 165-187.

Wolfram, L.J., 2003. Human hair: A unique physicochemical composite, Journal of the American Academy of Dermatology 48, S106-S114.

Yoneyama, T., Ohtani, T., 1983. Variations of natural ¹³C abundances in leguminous plants, Plant and Cell Physiology 24, 971-977.

Zlatar, V., 1984. Cementerio prehispánico Pica 8, Facultad de Educación y Ciencias Humanas, Universidad de Antofagasta.

Zori, C., Brant, E., 2012. Managing the risk of climatic variability in late prehistoric northern Chile, Journal of Anthropological Archaeology 31, 403-421.

Time (min)	Mobile	phases (%)	Flow rate (µl/min)
Conditioning run	А	В	С	
0	0	92	8	110
35	0	92	8	110
36	100	0	0	110
55	100	0	0	110
Analytical run	А	В	С	
0	100	0	0	60
45	100	0	0	60
65	60	40	0	60
75	40	25	35	60
150	0	0	100	60
180	0	0	100	60

Table 1. LC gradient program for Primesep A column (2.1x250 mm, 100 Å, 5 $\mu m).$

Individua	.1		Tendo	on colla	gen (th	nis study)		Rib collag Sagredo et	en (Santana- t al., 2015a)	\triangle Rib co tendon c	llagen- ollagen
Burial [*]	Sex	Age	%N	%C	C/N	δ^{15} N/‰	δ^{13} C/‰	δ^{15} N/‰	δ^{13} C/‰	δ^{15} N/‰	δ^{13} C/‰
SI-T74	F	20-35 yrs	15.9	46.1	3.4	$+25.3 \pm 0.0$	-10.8 ± 0.0	+22.9	-10.5	-2.4	0.3
			16.1	46.0	3.3						
SD-T24	М	20-35 yrs	16.1	45.7	3.3	$+16.2 \pm 0.1$	-8.2 ± 0.0	+14.2	-9.6	-2.0	-1.4
			16.1	45.6	3.3						
SI-T32	F	35-50 yrs	15.8	45.9	3.4	$+16.1\pm0.0$	-8.5 ± 0.0				
			15.7	46.1	3.4						
SE-T3	F	20-35 yrs	16.1	46.6	3.4	$+12.2\pm0.1$	-18.3 ± 0.0	+10.9	-18.6	-1.3	-0.3
			15.9	46.0	3.4						
SF-T4	F	35-50 yrs	16.5	45.6	3.2	$+17.5 \pm 0.0$	-7.8 ± 0.0				
			16.0	44.2	3.2						
SI-T3	М	35-50 yrs	16.0	46.6	3.4	$+15.5 \pm 0.0$	-8.7 ± 0.1	+21.0	-10.4	5.5	-1.7
			15.5	45.1	3.4						

Table 2. Bulk carbon and nitrogen isotope compositions of collagens from Pica 8 individuals.

*S indicates the burial sector; T indicates the grave number.

Pica	SD-T2	4	SD-T24	Ļ	SE-T3		SE-T3		SF-T4	
	Tendor	n	Bone		Tendon		Bone		Tendon	
Food (%)	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C3 plants	7	5	13	7	77	7	81	6	6	4
C4 plants	72	8	65	9	8	6	9	6	70	9
Terrestrial animals	11	8	18	11	11	8	7	6	9	7
Marine animals	10	4	4	3	4	3	3	2	15	5
Food fractions (%)										
Protein	27	3	29	4	30	2	27	2	28	3
Energy	73	3	71	3	70	2	73	2	72	3
Dietary proxies										
(Food)(%)										
$\delta^{13}C_{col}$ (C3 plants)	7	5	12	8	71	9	77	7	5	4
$\delta^{13}C_{col}$ (C4 plants)	61	10	54	11	7	5	8	5	59	11
$\delta^{13}C_{col}$ (Terr. animals)	18	11	28	15	17	12	11	8	14	9
$\delta^{13}C_{col}$ (Mar. animals)	14	6	6	4	5	4	4	3	22	7
$\delta^{15}N_{col}$ (C3 plants)	7	5	11	8	60	12	70	10	5	4
$\delta^{15}N_{col}$ (C4 plants)	39	12	34	12	4	3	5	4	37	11
δ^{15} N _{col} (Terr. animals)	30	17	45	20	27	16	19	13	23	14
$\delta^{15}N_{col}$ (Mar. animals)	24	9	10	7	9	6	6	5	35	9
D '	CT T2		CT T2		0T T22		01 T74		QI T74	
Pica	SI-T3	_	SI-T3		SI-T32		SI-T74		SI-T74	
Pica	SI-T3 Tendor	1	SI-T3 Bone	CD	SI-T32 Tendon	CD	SI-T74 Tendon	(D	SI-T74 Bone	<u>CD</u>
Pica Food (%)	SI-T3 Tendor Mean	n SD	SI-T3 Bone Mean	SD	SI-T32 Tendon Mean	SD	SI-T74 Tendon Mean	SD	SI-T74 Bone Mean	SD
Pica Food (%) C3 plants	SI-T3 Tendor Mean 9	n SD 6	SI-T3 Bone Mean 16	SD 8	SI-T32 Tendon Mean 8	SD 6	SI-T74 Tendon Mean 12	SD 7	SI-T74 Bone Mean 15	SD 8
Pica Food (%) C3 plants C4 plants	SI-T3 Tendor Mean 9 69	n SD 6 9	SI-T3 Bone Mean 16 40	SD 8 9	SI-T32 Tendon Mean 8 69	SD 6 9	SI-T74 Tendon Mean 12 36	SD 7 7	SI-T74 Bone Mean 15 36	SD 8 8
Pica Food (%) C3 plants C4 plants Terrestrial animals	SI-T3 Tendor Mean 9 69 15	n SD 6 9 9	SI-T3 Bone Mean 16 40 6	SD 8 9 5	SI-T32 Tendon Mean 8 69 14	SD 6 9 9	SI-T74 Tendon Mean 12 36 1	SD 7 7 2	SI-T74 Bone Mean 15 36 3	SD 8 8 3
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals	SI-T3 Tendon 9 69 15 7	n SD 6 9 9 4	SI-T3 Bone Mean 16 40 6 38	SD 8 9 5 7	SI-T32 Tendon Mean 8 69 14 9	SD 6 9 9 4	SI-T74 Tendon Mean 12 36 1 51	SD 7 7 2 4	SI-T74 Bone Mean 15 36 3 46	SD 8 8 3 5
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%)	SI-T3 Tendor Mean 9 69 15 7	n SD 6 9 9 4	SI-T3 Bone Mean 16 40 6 38	SD 8 9 5 7	SI-T32 Tendon Mean 8 69 14 9	SD 6 9 9 4	SI-T74 Tendon Mean 12 36 1 51	SD 7 7 2 4	SI-T74 Bone Mean 15 36 3 46	SD 8 8 3 5
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein	SI-T3 Tendor Mean 9 69 15 7 28 72	n SD 6 9 9 4 3	SI-T3 Bone Mean 16 40 6 38 40 60	SD 8 9 5 7	SI-T32 Tendon Mean 8 69 14 9 29 71	SD 6 9 9 4	SI-T74 Tendon Mean 12 36 1 51 43	SD 7 7 2 4	SI-T74 Bone Mean 15 36 3 46 42	SD 8 8 3 5
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy	SI-T3 Tendor Mean 9 69 15 7 28 72	n SD 6 9 9 4 3 3	SI-T3 Bone Mean 16 40 6 38 40 60	SD 8 9 5 7 2 2	SI-T32 Tendon Mean 8 69 14 9 29 71	SD 6 9 9 4 3 3	SI-T74 Tendon Mean 12 36 1 51 43 57	SD 7 7 2 4 1 1	SI-T74 Bone Mean 15 36 3 46 42 58	SD 8 8 3 5 1 1
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Terrest)(%)	SI-T3 Tendor Mean 9 69 15 7 28 72	n SD 6 9 9 4 3 3	SI-T3 Bone Mean 16 40 6 38 40 60	SD 8 9 5 7 2 2	SI-T32 Tendon Mean 8 69 14 9 29 71	SD 6 9 9 4 3 3	SI-T74 Tendon Mean 12 36 1 51 43 57	SD 7 7 2 4 1 1	SI-T74 Bone Mean 15 36 3 46 42 58	SD 8 8 3 5 1 1
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%)	SI-T3 Tendor Mean 9 69 15 7 28 72	n SD 6 9 9 4 3 3	SI-T3 Bone Mean 16 40 6 38 40 60	SD 8 9 5 7 2 2	SI-T32 Tendon Mean 8 69 14 9 29 71	SD 6 9 4 3 3	SI-T74 Tendon Mean 12 36 1 51 43 57	SD 7 7 2 4 1 1	SI-T74 Bone Mean 15 36 3 46 42 58	SD 8 8 3 5 1 1
PicaFood (%)C3 plantsC4 plantsTerrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 50	n SD 6 9 4 3 3 6	SI-T3 Bone Mean 16 40 6 38 40 60	SD 8 9 5 7 2 2 6	SI-T32 Tendon Mean 8 69 14 9 29 71 7	SD 6 9 4 3 3	SI-T74 Tendon Mean 12 36 1 51 43 57 9	SD 7 7 2 4 1 1 5 5	SI-T74 Bone Mean 15 36 3 46 42 58	SD 8 3 5 1 1 1
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (C4 plants)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 58 22	n SD 6 9 9 4 3 3 6 11 12	SI-T3 Bone Mean 16 40 6 38 40 60 13 29 0	SD 8 9 5 7 2 2 6 7 6	SI-T32 Tendon Mean 8 69 14 9 29 71 7 58 21	SD 6 9 4 3 3 6 11	SI-T74 Tendon Mean 12 36 1 51 43 57 9 24 2	SD 7 7 2 4 1 1 5 5 2	SI-T74 Bone Mean 15 36 3 46 42 58 12 25 5	SD 8 3 5 1 1 1 6 6
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (C4 plants) $\delta^{13}C_{col}$ (Terr. animals)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 58 22	n SD 6 9 9 4 3 3 6 11 13 5	SI-T3 Bone Mean 16 40 6 38 40 60 13 29 9 40	SD 8 9 5 7 2 2 2 6 7 6 7 6	SI-T32 Tendon Mean 8 69 14 9 29 71 7 58 21	SD 6 9 4 3 3 6 11 13	SI-T74 Tendon Mean 12 36 1 51 43 57 9 24 3 6	SD 7 7 2 4 1 1 5 5 2	SI-T74 Bone Mean 15 36 3 46 42 58 12 25 5 5	SD 8 8 3 5 1 1 1 6 6 4
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (C4 plants) $\delta^{13}C_{col}$ (Terr. animals) $\delta^{13}C_{col}$ (Mar. animals)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 58 22 11	n SD 6 9 4 3 3 6 11 13 5 6	SI-T3 Bone Mean 16 40 6 38 40 60 8 13 29 9 49 0	SD 8 9 5 7 2 2 2 6 7 6 7 5	SI-T32 Tendon Mean 8 69 14 9 29 71 7 58 21 14 7	SD 6 9 4 3 3 6 11 13 6 6	SI-T74 Tendon Mean 12 36 1 51 43 57 9 24 3 64 64	SD 7 7 2 4 1 1 5 5 2 4 2	SI-T74 Bone Mean 15 36 3 46 42 58 12 25 5 5 5 8	SD 8 3 5 1 1 1 6 6 4 6
PicaFood (%)C3 plantsC4 plantsTerrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (C4 plants) $\delta^{13}C_{col}$ (Terr. animals) $\delta^{13}C_{col}$ (Mar. animals) $\delta^{15}N_{col}$ (C3 plants)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 58 22 11 8 27	n SD 6 9 9 4 3 3 6 11 13 5 6 12	SI-T3 Bone Mean 16 40 6 38 40 60 8 40 60 13 29 9 49 9 14	SD 8 9 5 7 2 2 2 6 7 6 7 5 5 5	SI-T32 Tendon Mean 8 69 14 9 29 71 7 58 21 14 7 26	SD 6 9 4 3 3 6 11 13 6 6	SI-T74 Tendon Mean 12 36 1 51 43 57 9 24 3 64 6 10	SD 7 7 2 4 1 1 5 5 2 4 3 2	SI-T74 Bone Mean 15 36 3 46 42 58 12 25 5 5 8 8 12	SD 8 8 3 5 1 1 1 6 6 4 6 4
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (C4 plants) $\delta^{13}C_{col}$ (Mar. animals) $\delta^{15}N_{col}$ (C3 plants) $\delta^{15}N_{col}$ (C4 plants) $\delta^{15}N_{col}$ (C4 plants)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 58 22 11 8 37 27	n SD 6 9 9 4 3 3 6 11 13 5 6 12	SI-T3 Bone Mean 16 40 6 38 40 60 60 13 29 9 49 9 14 12	SD 8 9 5 7 2 2 2 6 7 6 7 5 5 5	SI-T32 Tendon Mean 8 69 14 9 29 71 7 58 21 14 7 36 24	SD 6 9 4 3 3 6 11 13 6 6 12	SI-T74 Tendon Mean 12 36 1 51 43 57 9 24 3 64 6 10 2	SD 7 7 2 4 1 1 1 5 5 5 2 4 3 3 2	SI-T74 Bone Mean 15 36 3 46 42 58 12 25 5 5 8 8 12 6	SD 8 8 3 5 1 1 1 6 6 4 6 4 4 5
Pica Food (%) C3 plants C4 plants Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (C4 plants) $\delta^{13}C_{col}$ (Mar. animals) $\delta^{15}N_{col}$ (C3 plants) $\delta^{15}N_{col}$ (C4 plants) $\delta^{15}N_{col}$ (C4 plants) $\delta^{15}N_{col}$ (C4 plants) $\delta^{15}N_{col}$ (C4 plants)	SI-T3 Tendor Mean 9 69 15 7 28 72 9 58 22 11 8 37 37 18	n SD 6 9 9 4 3 3 6 11 13 5 6 12 19 0	SI-T3 Bone Mean 16 40 6 38 40 60 13 29 9 49 9 49 9 14 12 65	SD 8 9 5 7 2 2 2 6 7 6 7 5 5 9 8	SI-T32 Tendon Mean 8 69 14 9 29 71 7 58 21 14 7 36 34 22	SD 6 9 4 3 3 6 11 13 6 6 12 18 0	SI-T74 Tendon Mean 12 36 1 51 43 57 9 24 3 64 6 10 3 81	SD 7 7 2 4 1 1 5 5 2 4 3 3 3 4	SI-T74 Bone Mean 15 36 3 46 42 58 12 25 5 5 8 8 12 6 74	SD 8 8 3 5 1 1 1 6 6 4 6 4 4 5 6

Table 3. FRUITS model diet estimates based on the collagen isotope data from the Pica 8 individuals.

Energy combines the contribution of lipids and carbohydrates. Food (%) represents the calorie contribution of each food group; Fraction (%) represents the calorie contribution of each food fraction; Dietary proxy (Food)(%) represents the calorie contribution of each food group to the dietary proxies. Estimates are normalised to 100%, and uncertainty is 1-sigma (Fernandes et al., 2014, Fernandes, 2015, Fernandes et al., 2015).

Pica	SD-T2	4	SD-T2	4	SE-T3		SE-T3		SF-T4	
	Tendor	n	Bone		Tendon		Bone		Tendon	
Food (%)	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C3 plants	6	5	10	7	79	7	84	6	5	5
Maize manured (guano)	59	7	50	6	7	5	5	4	64	7
Terrestrial animals	31	8	38	7	11	8	9	7	27	7
Marine animals	4	3	2	2	3	3	2	2	4	4
Food fractions (%)										
Protein	36	3	39	2	30	2	29	2	33	3
Energy	64	3	61	2	70	2	71	2	67	3
Dietary proxies										
(Food)(%)										
$\delta^{13}C_{col}$ (C3 plants)	5	5	8	6	73	9	79	8	5	4
δ^{13} C _{col} (Maize manured)	44	7	36	6	6	4	5	4	49	8
$\delta^{13}C_{col}$ (Terr. animals)	46	9	53	8	16	11	13	10	40	8
$\delta^{13}C_{col}$ (Mar. animals)	5	4	3	3	5	4	3	3	6	5
δ^{15} N _{col} (C3 plants)	4	4	6	5	63	12	70	12	4	4
δ^{15} N _{col} (Maize manured)	21	5	15	4	4	3	3	2	25	6
δ^{15} N _{col} (Terr. animals)	68	9	75	7	25	15	22	14	62	9
δ^{15} N _{col} (Mar. animals)	7	6	4	4	8	6	5	4	9	7
Pica	SI T3		SI T3		SI T32		SI T74		SI T74	
Pica	SI-T3	n	SI-T3 Bone		SI-T32 Tendon		SI-T74		SI-T74 Bone	
Pica	SI-T3 Tendor Mean	n SD	SI-T3 Bone	SD	SI-T32 Tendon	SD	SI-T74 Tendon	SD	SI-T74 Bone Mean	<u>SD</u>
Pica Food (%)	SI-T3 Tendor Mean	n SD	SI-T3 Bone Mean	SD 9	SI-T32 Tendon Mean	SD	SI-T74 Tendon Mean	SD 7	SI-T74 Bone Mean	SD 7
Pica Food (%) C3 plants Maize manured (muano)	SI-T3 Tendor Mean 7 56	n SD 5 7	SI-T3 Bone Mean 18 48	SD 9	SI-T32 Tendon Mean 6 57	SD 5 7	SI-T74 Tendon Mean 14 42	SD 7	SI-T74 Bone Mean 18 45	SD 7 12
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals	SI-T3 Tendor Mean 7 56 34	n SD 5 7 7	SI-T3 Bone Mean 18 48 12	SD 9 11 7	SI-T32 Tendon Mean 6 57 33	SD 5 7 7	SI-T74 Tendon Mean 14 42 3	SD 7 10 2	SI-T74 Bone Mean 18 45 6	SD 7 12 5
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals	SI-T3 Tendor Mean 7 56 34 3	n SD 5 7 7 3	SI-T3 Bone Mean 18 48 12 22	SD 9 11 7 10	SI-T32 Tendon Mean 6 57 33 4	SD 5 7 7 3	SI-T74 Tendon Mean 14 42 3 41	SD 7 10 2 9	SI-T74 Bone Mean 18 45 6 31	SD 7 12 5
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals Food fractions (%)	SI-T3 Tendor Mean 7 56 34 3	n SD 5 7 7 3	SI-T3 Bone Mean 18 48 12 22	SD 9 11 7 10	SI-T32 Tendon Mean 6 57 33 4	SD 5 7 7 3	SI-T74 Tendon Mean 14 42 3 41	SD 7 10 2 9	SI-T74 Bone Mean 18 45 6 31	SD 7 12 5 11
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals Food fractions (%) Protein	SI-T3 Tendor Mean 7 56 34 3 3	n SD 5 7 7 3	SI-T3 Bone Mean 18 48 12 22	SD 9 11 7 10	SI-T32 Tendon Mean 6 57 33 4 37	SD 5 7 7 3	SI-T74 Tendon Mean 14 42 3 41	SD 7 10 2 9	SI-T74 Bone Mean 18 45 6 31 36	SD 7 12 5 11
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals Food fractions (%) Protein Energy	SI-T3 Tendor Mean 7 56 34 3 3 38 62	n SD 5 7 7 3 3	SI-T3 Bone Mean 18 48 12 22 35 65	SD 9 11 7 10 3 3	SI-T32 Tendon Mean 6 57 33 4 37 63	SD 5 7 3 3	SI-T74 Tendon Mean 14 42 3 41 39 61	SD 7 10 2 9 2 2	SI-T74 Bone Mean 18 45 6 31 36 64	SD 7 12 5 11 3 3
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary provies	SI-T3 Tendor Mean 7 56 34 3 3 38 62	n SD 5 7 7 3 3 2	SI-T3 Bone Mean 18 48 12 22 35 65	SD 9 11 7 10 3 3	SI-T32 Tendon Mean 6 57 33 4 37 63	SD 5 7 3 3 2	SI-T74 Tendon Mean 14 42 3 41 39 61	SD 7 10 2 9 2 2	SI-T74 Bone Mean 18 45 6 31 36 64	SD 7 12 5 11 3 3
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%)	SI-T3 Tendor Mean 7 56 34 3 3 8 62	n SD 5 7 7 3 3 2	SI-T3 Bone Mean 18 48 12 22 35 65	SD 9 11 7 10 3 3	SI-T32 Tendon Mean 6 57 33 4 37 63	SD 5 7 3 3 2	SI-T74 Tendon Mean 14 42 3 41 39 61	SD 7 10 2 9 2 2	SI-T74 Bone Mean 18 45 6 31 36 64	SD 7 12 5 11 3 3
PicaFood (%)C3 plantsMaize manured (guano)Terrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{rel}$ (C3 plants)	SI-T3 Tendor Mean 7 56 34 3 3 8 62	n SD 5 7 7 3 3 2	SI-T3 Bone Mean 18 48 12 22 35 65	SD 9 11 7 10 3 3	SI-T32 Tendon Mean 6 57 33 4 37 63	SD 5 7 3 3 2	SI-T74 Tendon Mean 14 42 3 41 39 61	SD 7 10 2 9 2 2	SI-T74 Bone Mean 18 45 6 31 36 64	SD 7 12 5 11 3 3
PicaFood (%)C3 plantsMaize manured (guano)Terrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (Maize manured)	SI-T3 Tendor Mean 7 56 34 3 3 38 62 6 41	n SD 5 7 7 3 3 2 5 7	SI-T3 Bone Mean 18 48 12 22 35 65 16 38	SD 9 11 7 10 3 3 8 12	SI-T32 Tendon Mean 6 57 33 4 37 63 5 42	SD 5 7 3 3 2 4 7	SI-T74 Tendon Mean 14 42 3 41 39 61 11 31	SD 7 10 2 9 2 2 2 5 10	SI-T74 Bone Mean 18 45 6 31 36 64 15 35	SD 7 12 5 11 3 3 6 12
PicaFood (%)C3 plantsMaize manured (guano)Terrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (Terr animals)	SI-T3 Tendon 7 56 34 3 3 8 62 6 41 49	n 5 7 7 3 3 2 5 7 8	SI-T3 Bone Mean 18 48 12 22 35 65 16 38 16	SD 9 11 7 10 3 3 8 12 10	SI-T32 Tendon Mean 6 57 33 4 37 63 5 42 48	SD 5 7 3 3 2 4 7 8	SI-T74 Tendon 14 42 3 41 39 61 11 31 4	SD 7 10 2 9 2 2 2 5 10 3	SI-T74 Bone Mean 18 45 6 31 36 64 15 35 8	SD 7 12 5 11 3 3 6 12 6
Pica Food (%) C3 plants Maize manured (guano) Terrestrial animals Marine animals Food fractions (%) Protein Energy Dietary proxies (Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (Maize manured) $\delta^{13}C_{col}$ (Terr. animals) $\delta^{13}C_{col}$ (Mar. animals)	SI-T3 Tendon 7 56 34 3 3 8 62 6 41 49 4	n 5 7 7 3 3 2 5 7 8 4	SI-T3 Bone Mean 18 48 12 22 35 65 16 38 16 30	SD 9 11 7 10 3 3 8 12 10 13	SI-T32 Tendon Mean 6 57 33 4 37 63 5 42 48 5	SD 5 7 3 3 2 4 7 8 4	SI-T74 Tendon Mean 14 42 3 41 39 61 11 31 4 54	SD 7 10 2 9 2 2 2 5 10 3 10	SI-T74 Bone Mean 18 45 6 31 36 64 15 35 8 42	SD 7 12 5 11 3 3 6 12 6 12 6 13
PicaFood (%)C3 plantsMaize manured (guano)Terrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (Maize manured) $\delta^{13}C_{col}$ (Mar. animals) $\delta^{15}N_{col}$ (C3 plants)	SI-T3 Tendon 7 56 34 3 38 62 6 41 49 4 4	n 5 7 7 3 3 2 5 7 8 4 4	SI-T3 Bone Mean 18 48 12 22 35 65 16 38 16 30 13	SD 9 11 7 10 3 3 8 12 10 13 8	SI-T32 Tendon Mean 6 57 33 4 37 63 5 42 48 5 4	SD 5 7 3 3 2 4 7 8 4 4 4	SI-T74 Tendon Mean 14 42 3 41 39 61 11 31 4 54 8	SD 7 10 2 9 2 2 5 10 3 10 4	SI-T74 Bone Mean 18 45 6 31 36 64 15 35 8 42 12	SD 7 12 5 11 3 3 3 6 12 6 13 6
PicaFood (%)C3 plantsMaize manured (guano)Terrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (Maize manured) $\delta^{13}C_{col}$ (Mar. animals) $\delta^{15}N_{col}$ (C3 plants) $\delta^{15}N_{col}$ (Maize manured)	SI-T3 Tendor Mean 7 56 34 3 3 8 62 6 41 49 4 4 19	n SD 5 7 7 3 2 5 7 8 4 4 5	SI-T3 Bone Mean 18 48 12 22 35 65 16 38 16 30 13 21	SD 9 11 7 10 3 3 3 8 12 10 13 8 10	SI-T32 Tendon Mean 6 57 33 4 37 63 5 42 48 5 42 48 5 4 20	SD 5 7 3 3 2 4 7 8 4 4 5	SI-T74 Tendon Mean 14 42 3 41 39 61 11 31 4 54 8 16	SD 7 10 2 9 2 2 2 5 10 3 10 4 8	SI-T74 Bone Mean 18 45 6 31 36 64 15 35 8 42 12 12 19	SD 7 12 5 11 3 3 6 12 6 13 6 10
PicaFood (%)C3 plantsMaize manured (guano)Terrestrial animalsMarine animalsFood fractions (%)ProteinEnergyDietary proxies(Food)(%) $\delta^{13}C_{col}$ (C3 plants) $\delta^{13}C_{col}$ (Maize manured) $\delta^{13}C_{col}$ (Mar. animals) $\delta^{15}N_{col}$ (C3 plants) $\delta^{15}N_{col}$ (Maize manured) $\delta^{15}N_{col}$ (Maize manured) $\delta^{15}N_{col}$ (Maize manured)	SI-T3 Tendon 7 56 34 3 3 6 2 6 41 49 4 4 19 71	n SD 5 7 7 3 2 5 7 8 4 4 5 8	SI-T3 Bone Mean 18 48 12 22 35 65 16 38 16 30 13 21 25	SD 9 11 7 10 3 3 3 8 12 10 13 8 10 13	SI-T32 Tendon Mean 6 57 33 4 37 63 5 42 48 5 42 48 5 4 20 70	SD 5 7 3 3 2 4 7 8 4 4 5 8	SI-T74 Tendon Mean 14 42 3 41 39 61 11 31 4 54 8 16 5	SD 7 10 2 9 2 2 2 5 10 3 10 4 8 5	SI-T74 Bone Mean 18 45 6 31 36 64 15 35 8 42 12 19 12	SD 7 12 5 11 3 3 6 12 6 13 6 10 9

Table 4. FRUITS model diet estimates based on the collagen isotope data from the Pica 8 individuals (Model with 'maize fertilized with guano').



Fig 1. Map of northern Chile showing the location of the Pica 8 site.



Fig 2. LC-IRMS chromatogram of tendon collagen hydrolysate (from individual SE-T3).



Fig 3. Fractions of the total (%) of amino acid peak areas measured in archaeological tendon collagen (this study) and bone collagen (mean $\pm 1\sigma$), and of amino acid carbon weights (%) of human tendon collagen derived from Schofield et al. (1971).



Fig 4. Fractions of the total (%) of amino acid peak areas measured in archaeological and modern hair (mean $\pm 1\sigma$), and of amino acid carbon weights (%) measured in human hair (Robbins and Kelly, 1970; Wolfram, 2003).



Fig 5. Plot of δ^{15} N values versus δ^{13} C values of tendon collagen (*, this study) and bone collagen (†, Santana-Sagredo et al., 2015a) from Pica 8 individuals and of the edible portion of South American flora and fauna.



Fig 6. Plot of Δ^{13} C _{Val-Phe} values for tendon collagen (this study) and bone collagen (Honch et al., 2012). HMP = high-marine protein consumer; HFP = high-freshwater protein consumer; C4=C4 plant consumers; C3=C3 plant consumers; Peru = mixed-diet group, Peru, Huari AD 500–900.



Fig 7. Plot of δ^{13} C phenylalanine values vs. δ^{13} C values for tendon collagen (this study) and bone collagen (Honch et al., 2012).



Fig 8. Proposed model for assessing the origin of the predominant dietary intake (terrestrial vs. marine), based on tendon collagen amino acid δ^{13} C values.











Fig 9. For each individual, from top to bottom: Calorie contribution of each food group to the diet, estimated via Bayesian stable isotope mixing model FRUITS, based on bulk δ^{13} C and δ^{15} N values of tendon collagen (this study) and bone collagen (Santana-Sagredo et al., 2015a). (Asterisk indicates FRUITS model with 'maize fertilised with guano' replacing 'C4 plants' food group); Amino acid δ^{13} C values and calculated δ^{13} C mass balance (MB) values for tendon collagen; Keratin δ^{13} C values of non-essential and essential amino acids, and calculated δ^{13} C mass balance (MB) values, along the hair fibre.

Appendix A. Supplementary data

Bayesian stable isotope mixing model: FRUITS

The Bayesian mixing model Food Reconstruction Using Isotopic Transferred Signals (FRUITS) (Fernandes et al., 2014) has been applied to the tendon collagen isotope data produced in this study and to the bone collagen isotope data published by Santana-Sagredo et al. (2015) in order to achieve an estimation of the qualitative and quantitative nutritional intake of the Pica individuals.

The dietary proxies used in the FRUITS models are the tendon and rib collagen δ^{15} N and δ^{13} C values with an associated uncertainty of 0.5‰ to account for instrumental analytical error. The food groups used include the widest array of foods available through trade from the Pacific coast, coastal valleys, mid-altitude valleys and highlands. The food groups consist of: (1) C4 plants (cultivated and wild), (2) marine animals (fish, sea lions, shellfish), (3) C3 plants (fruits, vegetables, legumes), and (4) terrestrial animals (camelids, rodents and birds) consuming either C3, C4 or mixed C3-C4 plants (i.e. wild and domesticated animals living at different elevations) (Mora et al., 2017). To account for the use of guano (seabird dung) for maize cultivation, a second model was run, substituting the 'C4 plants' food group with 'maize manured with guano' (Table A.1).

Food group	Food fraction	δ ¹³ C/‰	$\delta^{15}N/\%$	Concentration (%)
C3 plants	Protein	-26.7 ± 1	$+4.1\pm1$	23±2.5
_	Energy [†]	-24.2 ± 1		77±2.5
C4 plants [*]	Protein	-12.3 ± 1	$+8.1\pm1$	14±2.5
	Energy [†]	$-9.8{\pm}1$		86±2.5
Terrestrial animals	Protein	-17.7 ± 1	$+8.9\pm1$	78±2.5
	Energy [†]	-23.7 ± 1		22±2.5
Marine animals	Protein	-13.5 ± 1	$+19.2\pm1$	68±2.5
	Energy [†]	-19.5 ± 1		32±2.5
*Substituting food group:				
Maize fertilised with guano	Protein	-11.8 ± 1	$+22.8\pm1$	11±2.5
	Energy [†]	-9.3 ± 1		89±2.5
*E 11 /1 /1		1 1 /		

Table A.1. Isotope values and concentrations of food fractions for each food group used in the FRUITS models.

[†] Energy combines the contribution of lipids and carbohydrates

Details on calculations of isotope values and concentrations of protein, lipid and carbohydrate for each food group were reported in Mora et al. (2017) and were based on Fernandes (2015) and Newsome et al. (2004). As in Mora et al. (2017), the carbon contribution from dietary proteins has been set below 45% by applying an a priori constraint to the FRUITS model. The diet-to-tissue offset for human bone collagen has been estimated by Fernandes et al. (2012) via regression analysis performed on isotope data measured in controlled feeding experiments on animals. The statistical analysis showed that carbon in collagen is routed by about $74\pm4\%$ from dietary proteins and by 26% from the energetic macronutrients (lipids and

carbohydrates), while nitrogen in collagen is derived from only dietary proteins (100%) (Fernandes et al., 2012, Fernandes, 2015). The resulting diet-to-collagen offsets, which have been proposed by Fernandes et al. (2012, 2015) and used by several authors (Andrade et al., 2015, Fernandes et al., 2015), are: $4.8\pm0.5\%$ for δ^{13} C and $5.5\pm0.5\%$ for δ^{15} N values. Given that the contribution of the energetic macronutrients to the (bone) collagen carbon is from the three glycolytic amino acids (serine, glycine, alanine) (Fernandes et al., 2012) and that the amino acid composition of bone and tendon collagen is similar (if not identical, being both composed of type I collagen) (Eastoe, 1955), the same diet-to-collagen offsets and macronutrients' contributions will be used in the present model for the collagens extracted from bones and tendons.

It is acknowledged that it is not possible to take into account the isotope fractionation that might exist associated with the different protein metabolism and remodelling between bone and tendon. Furthermore, these FRUITS models do not account for possible alterations of the healthy metabolism induced by malnutrition or disease.

References

Andrade, P., Fernandes, R., Codjambassis, K., Urrea, J., Olguin, L., Rebolledo, S., Lira, F., Aravena, C., Berríos, M., 2015. Subsistence continuity linked to consumption of marine protein in the Formative Period in the interfluvic coast of northern Chile: Re-assessing contacts with agropastoral groups from highlands, Radiocarbon 57, 679-688.

Eastoe, J.E., 1955. The amino acid composition of mammalian collagen and gelatin, Biochemical Journal 61, 589-600.

Fernandes, R., Nadeau, M.-J., Grootes, P.M., 2012. Macronutrient-based model for dietary carbon routing in bone collagen and bioapatite, Archaeological and Anthropological Sciences 4, 291-301.

Fernandes, R., Millard, A.R., Brabec, M., Nadeau, M.-J., Grootes, P., 2014. Food reconstruction using isotopic transferred signals (FRUITS): A Bayesian model for diet reconstruction, PLoS ONE 9, e87436.

Fernandes, R., 2015. A simple(r) model to predict the source of dietary carbon in individual consumers, Archaeometry 58, 500-512.

Fernandes, R., Grootes, P., Nadeau, M.-J., Nehlich, O., 2015. Quantitative diet reconstruction of a Neolithic population using a Bayesian mixing model (FRUITS): The case study of Ostorf (Germany), American Journal of Physical Anthropology 158, 325-340.

Mora, A., Arriaza, B.T., Standen, V.G., Valdiosera, C., Salim, A., Smith, C., 2017. High-resolution palaeodietary reconstruction: Amino acid δ 13C analysis of keratin from single hairs of mummified human individuals, Quaternary International 436, 96-113.

Newsome, S.D., Phillips, D.L., Culleton, B.J., Guilderson, T.P., Koch, P.L., 2004. Dietary reconstruction of an early to middle Holocene human population from the central California coast: Insights from advanced stable isotope mixing models, Journal of Archaeological Science 31, 1101-1115.

Santana-Sagredo, F., Lee-Thorp, J.A., Schulting, R., Uribe, M., 2015. Isotopic evidence for divergent diets and mobility patterns in the Atacama Desert, northern Chile, during the Late Intermediate Period (AD 900-1450), American Journal of Physical Anthropology 156, 374-387.

Appendix B. Supplementary data

	Asx	Нур	Ser	Glx	Thr	Gly	Ala	Pro	Val	Hyl	Ile	Leu	Lys	Tyr	Arg	Phe	MB	Bulk
SI-T74	-11.0	-10.5	-7.8	-9.9	-7.6	-4.9	-13.0	-12.0	-18.9	-14.1	-13.6	-19.8	-13.7	-17.7	-13.8	-20.6		
SI-T74	-10.5	-10.3	-8.0	-9.4	-8.4	-4.7	-13.1	-11.7	-18.3	-14.4	-13.9	-20.0	-14.0	-18.6	-14.2	-20.5		
mean	-10.8	-10.4	-7.9	-9.7	-8.0	-4.8	-13.0	-11.8	-18.6	-14.2	-13.7	-19.9	-13.9	-18.1	-14.0	-20.6	-11.3	-10.8
SD	0.3	0.2	0.1	0.3	0.6	0.1	0.0	0.2	0.4	0.2	0.2	0.2	0.2	0.6	0.3	0.1		
SD-T24	-8.3	-7.2	-6.5	-6.5	-7.9	-3.3	-9.1	-8.8	-18.2	-13.6	-13.3	-18.4	-12.2	-15.0	-10.5	-17.2		
SD-T24	-8.7	-7.7	-6.3	-7.0	-8.3	-4.0	-9.4	-9.0	-17.7	-13.6	-12.9	-18.5	-11.9	-15.2	-10.5	-17.2		
mean	-8.5	-7.5	-6.4	-6.7	-8.1	-3.6	-9.2	-8.9	-18.0	-13.6	-13.1	-18.5	-12.1	-15.1	-10.5	-17.2	-8.9	-8.2
SD	0.3	0.3	0.2	0.4	0.2	0.5	0.2	0.2	0.4	0.0	0.3	0.1	0.2	0.2	0.0	0.0		
SI-T32	-8.2	-8.0	-5.2	-7.0	-8.2	-3.7	-10.6	-8.5	-18.1	-13.7	-13.4	-18.3	-12.0		-10.2	-17.0		
SI-T32	-7.9	-7.6	-5.5	-6.5	-7.6	-4.0	-10.6	-8.4	-17.9	-13.1	-14.2	-18.7	-12.5	-14.4	-10.3	-17.3		
mean	-8.1	-7.8	-5.3	-6.7	-7.9	-3.8	-10.6	-8.5	-18.0	-13.4	-13.8	-18.5	-12.3	-14.4	-10.2	-17.1	-9.0	-8.5
SD	0.2	0.3	0.2	0.4	0.4	0.3	0.0	0.1	0.1	0.4	0.5	0.3	0.4		0.0	0.2		
SE-T3	-19.6	-17.1	-17.7	-18.4	-15.6	-13.7	-20.5	-17.7	-27.8	-19.6	-21.8	-28.4	-19.7	-23.5	-20.3	-25.7		
SE-T3	-19.2	-16.8	-17.3	-18.8	-16.1	-14.5	-20.7	-18.0	-27.4	-20.0	-22.1	-28.2	-19.6	-24.9	-20.3	-26.0		
mean	-19.4	-16.9	-17.5	-18.6	-15.9	-14.1	-20.6	-17.9	-27.6	-19.8	-21.9	-28.3	-19.7	-24.2	-20.3	-25.9	-18.8	-18.3
SD	0.3	0.2	0.2	0.3	0.4	0.6	0.2	0.2	0.3	0.3	0.2	0.1	0.1	1.0	0.0	0.3		
SF-T4	-8.3	-7.7	-7.3	-6.6	-8.2	-3.7	-8.0	-8.0	-17.1	-12.0	-12.8	-18.7	-11.9	-15.6	-10.5	-17.1		
SF-T4	-7.9	-7.6	-7.4	-6.6	-7.4	-3.2	-8.4	-7.7	-16.6	-11.5	-13.1	-18.4	-11.6		-10.2	-16.5		
mean	-8.1	-7.6	-7.3	-6.6	-7.8	-3.4	-8.2	-7.8	-16.8	-11.7	-13.0	-18.5	-11.7	-15.6	-10.4	-16.8	-8.5	-7.8
SD	0.3	0.1	0.1	0.0	0.6	0.3	0.3	0.2	0.3	0.3	0.2	0.3	0.2		0.2	0.4		
SI-T3	-9.6	-8.6	-6.9	-7.0	-10.4	-3.7	-9.9	-9.4	-18.3	-12.2	-14.1	-18.2	-11.8	-14.8	-9.4	-16.6		
SI-T3	-9.2	-8.3	-6.6	-6.8	-9.7	-4.2	-10.0	-9.2	-18.0	-12.4	-14.0	-18.0	-11.3	-14.1	-8.9	-16.2		
mean	-9.4	-8.5	-6.8	-6.9	-10.0	-3.9	-10.0	-9.3	-18.1	-12.3	-14.1	-18.1	-11.6	-14.4	-9.1	-16.4	-9.1	-8.7
SD	0.3	0.2	0.2	0.1	0.4	0.3	0.1	0.1	0.2	0.2	0.1	0.2	0.3	0.5	0.3	0.3		

Amino acid δ^{13} C (‰) values (in order of LC elution), calculated δ^{13} C (‰) Mass Balance (MB) values and bulk δ^{13} C (‰) values for tendon collagen.

Appendix C. Supplementary data

Table C.1. Amino acid δ^{13} C values in order of LC elution and calculated δ^{13} C Mass Balance (MB) values for hair keratin (1cm segment sequentially cut along the hair fibre starting at the root).

Individual	cm	Asx	Ser	Glx	Thr	Gly	Ala	Pro	Val	Ile	Leu	Lys	Tyr	Arg	Phe	MB
SI-T74	1	-9.2	-3.9	-6.8	-2.5	-9.8	-12.5	-9.6	-15.9	-10.4	-17.5	-11.8	-16.4	-11.3	-17.2	
SI-T74	1	-9.5	-3.5	-6.8	-2.0	-8.8	-12.3	-9.3	-16.1	-10.3	-17.0	-11.4	-17.0	-10.8	-16.8	
SI-T74	mean	-9.4	-3.7	-6.8	-2.3	-9.3	-12.4	-9.5	-16.0	-10.3	-17.2	-11.6	-16.7	-11.0	-17.0	-10.3
	SD	0.2	0.3	0.0	0.4	0.7	0.2	0.2	0.2	0.0	0.3	0.3	0.4	0.4	0.3	
SI-T74	2	-9.5	-3.7	-8.0	-3.3	-8.9	-11.8	-12.0	-17.2	-12.9	-18.8	-12.7	-18.0	-12.6	-18.9	
SI-T74	2	-9.1	-3.7	-7.6	-2.7	-9.6	-11.5	-12.4	-17.4	-13.3	-18.7	-12.3	-18.0	-12.6	-18.6	
SI-T74	mean	-9.3	-3.7	-7.8	-3.0	-9.2	-11.6	-12.2	-17.3	-13.1	-18.8	-12.5	-18.0	-12.6	-18.7	-11.4
	SD	0.3	0.1	0.3	0.4	0.6	0.2	0.3	0.2	0.3	0.1	0.3	0.0	0.0	0.2	
SI-T74	3	-9.8	-4.9	-7.3	-3.6	-12.5	-12.7	-14.2	-18.8	-14.3	-20.6	-15.7	-19.6	-14.0	-20.3	
SI-T74	3	-9.3	-5.0	-7.4	-3.1	-11.6	-12.3	-13.6	-18.2	-14.7	-20.2	-16.3	-20.2	-14.0	-20.3	
SI-T74	mean	-9.6	-5.0	-7.3	-3.4	-12.0	-12.5	-13.9	-18.5	-14.5	-20.4	-16.0	-19.9	-14.0	-20.3	-12.5
	SD	0.3	0.1	0.1	0.3	0.6	0.3	0.4	0.4	0.3	0.3	0.4	0.4	0.0	0.0	
SI-T74	4	-10.4	-5.0	-8.7	-3.7	-11.5	-12.4	-15.3	-17.0	-12.4	-19.0	-13.5	-18.8	-13.9	-18.4	
SI-T74	4	-10.5	-5.0	-9.0	-2.9	-12.1	-12.6	-15.1	-16.9	-12.8	-18.6	-13.1		-13.4	-18.1	
SI-T74	mean	-10.4	-5.0	-8.8	-3.3	-11.8	-12.5	-15.2	-16.9	-12.6	-18.8	-13.3	-18.8	-13.6	-18.3	-12.3
	SD	0.1	0.0	0.2	0.6	0.4	0.1	0.1	0.1	0.3	0.3	0.3		0.3	0.2	
SI-T74	5	-7.7	-2.4	-6.3	-3.6	-12.8	-12.0	-12.0	-16.0	-11.9	-18.7	-14.2	-18.1	-11.9	-19.0	
SI-T74	5	-7.8	-2.7	-6.1	-4.5	-13.1	-12.3	-12.7	-16.6	-12.7	-19.3	-14.8		-12.3	-19.6	
SI-T74	mean	-7.8	-2.5	-6.2	-4.0	-12.9	-12.2	-12.3	-16.3	-12.3	-19.0	-14.5	-18.1	-12.1	-19.3	-11.2
	SD	0.1	0.2	0.1	0.6	0.2	0.2	0.5	0.5	0.5	0.4	0.4		0.3	0.4	
SI-T74	6	-9.1	-2.3	-5.6	-3.8	-10.0	-10.8	-12.0	-16.9	-14.3	-20.2	-15.0	-18.5	-13.5	-20.5	
SI-T74	6	-9.3	-2.4	-5.0	-4.0	-9.7	-10.8	-12.4	-17.4	-13.9	-20.0	-15.4	-19.1	-13.4	-19.9	
SI-T74	mean	-9.2	-2.3	-5.3	-3.9	-9.8	-10.8	-12.2	-17.2	-14.1	-20.1	-15.2	-18.8	-13.4	-20.2	-11.4
	SD	0.2	0.0	0.4	0.2	0.2	0.0	0.3	0.4	0.2	0.1	0.2	0.4	0.1	0.4	
SI-T74	7	-6.6	-0.5	-3.8	-2.6	-9.6	-7.8	-8.2	-15.5	-11.0	-17.4	-11.5	-17.1	-10.3	-17.2	
SI-T74	7	-6.8	-0.9	-3.7	-3.5	-10.3	-8.2	-8.7	-16.0	-11.4	-17.8	-12.1		-10.7	-17.6	
SI-T74	mean	-6.7	-0.7	-3.8	-3.1	-10.0	-8.0	-8.4	-15.7	-11.2	-17.6	-11.8	-17.1	-10.5	-17.4	-9.2
	SD	0.2	0.3	0.1	0.6	0.5	0.3	0.3	0.4	0.3	0.2	0.4		0.3	0.3	
SI-T74	8	-9.2	-1.8	-9.0	-4.8	-11.0	-14.5	-14.8								
SI-T74	8	-9.5	-1.4	-8.4	-3.9	-11.9	-13.7	-14.3	-17.3	-13.2	-19.5	-13.7	-17.8	-13.3	-21.0	

SI-T74	mean	-9.3	-1.6	-8.7	-4.3	-11.5	-14.1	-14.6	-17.3	-13.2	-19.5	-13.7	-17.8	-13.3	-21.0	-12.2
	SD	0.2	0.3	0.4	0.6	0.7	0.6	0.3								
SI-T74	9	-10.2	-1.0	-9.9	-2.7	-9.1	-12.7	-12.1	-17.0	-13.2	-20.1	-13.8	-18.8	-14.7	-21.4	
SI-T74	9	-10.6	-0.8	-10.2	-3.3	-8.1	-12.8	-11.9	-17.0	-13.6	-20.2	-14.2	-18.4	-14.8	-21.3	
SI-T74	mean	-10.4	-0.9	-10.1	-3.0	-8.6	-12.8	-12.0	-17.0	-13.4	-20.1	-14.0	-18.6	-14.7	-21.3	-12.1
	SD	0.2	0.2	0.2	0.5	0.7	0.1	0.2	0.0	0.3	0.0	0.3	0.2	0.1	0.1	
SI-T74	10	-9.6	-0.9	-7.6	-5.5	-9.5	-11.6	-10.9	-16.3	-12.6	-19.3	-13.5	-16.8	-13.3	-19.7	
SI-T74	10	-9.4	-0.5	-7.5	-5.5	-8.7	-11.2	-10.3	-16.0	-12.0	-19.0	-13.0	-17.4	-12.9	-19.6	
SI-T74	mean	-9.5	-0.7	-7.6	-5.5	-9.1	-11.4	-10.6	-16.2	-12.3	-19.1	-13.2	-17.1	-13.1	-19.6	-11.1
	SD	0.1	0.2	0.1	0.0	0.6	0.3	0.4	0.2	0.4	0.2	0.4	0.4	0.2	0.1	
SD-T24	1	-7.7	-2.2	-6.7	-0.8	-12.3	-9.1	-8.7	-15.7	-12.4	-18.5	-11.8	-16.0	-10.0	-16.1	
SD-T24	1	-8.1	-2.0	-6.3	-0.9	-13.0	-9.4	-8.4	-15.7	-11.9	-17.9	-11.3	-15.8	-9.4	-16.0	
SD-T24	mean	-7.9	-2.1	-6.5	-0.9	-12.6	-9.2	-8.5	-15.7	-12.1	-18.2	-11.6	-15.9	-9.7	-16.0	-9.7
	SD	0.2	0.1	0.3	0.1	0.5	0.2	0.3	0.0	0.3	0.4	0.3	0.1	0.4	0.1	
SD-T24	2	-7.5	-2.6	-6.1	-2.4	-9.9	-9.8	-9.0	-15.7	-11.9	-18.0	-12.0	-15.3	-9.9	-16.4	
SD-T24	2	-7.9	-3.0	-6.7	-2.9	-9.3	-10.4	-9.2	-16.3	-12.4	-18.6	-12.2	-14.7	-10.1	-16.9	
SD-T24	mean	-7.7	-2.8	-6.4	-2.6	-9.6	-10.1	-9.1	-16.0	-12.1	-18.3	-12.1	-15.0	-10.0	-16.6	-9.9
	SD	0.3	0.3	0.4	0.4	0.4	0.4	0.2	0.4	0.4	0.4	0.2	0.4	0.1	0.4	
SD-T24	3	-7.8	-3.1	-6.7	-4.7	-10.8	-7.7	-8.1	-14.0	-9.2	-15.5	-9.1	-14.1	-7.6	-13.7	
SD-T24	3	-8.0	-2.5	-6.3	-4.3	-10.5	-7.1	-7.5	-13.4	-8.6	-15.3	-8.5	-14.2	-7.0	-13.7	
SD-T24	mean	-7.9	-2.8	-6.5	-4.5	-10.7	-7.4	-7.8	-13.7	-8.9	-15.4	-8.8	-14.2	-7.3	-13.7	-8.7
	SD	0.2	0.4	0.3	0.3	0.3	0.4	0.4	0.4	0.4	0.1	0.4	0.1	0.4	0.0	
SD-T24	4	-8.3	-3.4	-7.0	-5.2	-10.6	-11.7	-11.8	-15.3	-10.1	-17.1	-10.2	-13.4	-10.4	-15.7	
SD-T24	4	-8.1	-2.9	-6.5	-4.6	-11.4	-11.7	-11.3	-14.8	-10.6	-16.5	-10.3	-13.9	-10.1	-15.2	
SD-T24	mean	-8.2	-3.2	-6.7	-4.9	-11.0	-11.7	-11.6	-15.0	-10.3	-16.8	-10.2	-13.6	-10.3	-15.4	-10.1
	SD	0.1	0.4	0.4	0.4	0.5	0.0	0.4	0.4	0.3	0.4	0.1	0.4	0.2	0.3	
SD-T24	5	-13.4	-8.7	-12.0	-5.0	-13.2	-15.3	-16.1	-17.5	-13.9	-20.0	-13.0	-17.2	-15.5	-18.9	
SD-T24	5	-13.6	-8.6	-11.3	-4.6	-12.6	-15.7	-15.6	-17.7	-14.3	-19.5	-12.8	-17.8	-15.7	-19.2	
SD-T24	mean	-13.5	-8.7	-11.6	-4.8	-12.9	-15.5	-15.9	-17.6	-14.1	-19.8	-12.9	-17.5	-15.6	-19.0	-14.0
	SD	0.2	0.0	0.5	0.3	0.4	0.3	0.4	0.1	0.2	0.3	0.2	0.4	0.1	0.2	
SD-T24	6	-11.2	-4.4	-9.0	-4.4	-13.7	-14.8	-15.5	-16.7	-13.0	-19.2	-12.4	-14.6	-12.8	-17.3	
SD-T24	6	-11.2	-4.7	-8.8	-3.7	-14.7	-14.3	-15.6	-16.6	-12.6	-18.9	-12.3	-15.1	-13.1	-17.1	
SD-T24	mean	-11.2	-4.5	-8.9	-4.0	-14.2	-14.5	-15.5	-16.7	-12.8	-19.1	-12.4	-14.9	-12.9	-17.2	-12.3
	SD	0.0	0.2	0.1	0.5	0.7	0.3	0.1	0.1	0.2	0.2	0.0	0.3	0.2	0.2	
SD-T24	7	-10.4	-3.8	-7.6	-2.0	-11.2	-12.8	-12.1	-16.8	-13.0	-18.7	-13.3	-16.7	-12.6	-18.4	
SD-T24	7	-10.7	-4.3	-7.0	-2.9	-12.1	-12.2	-11.5	-16.2	-12.9	-18.2	-12.7		-12.0	-17.8	
SD-T24	mean	-10.5	-4.0	-7.3	-2.4	-11.6	-12.5	-11.8	-16.5	-12.9	-18.4	-13.0	-16.7	-12.3	-18.1	-11.3

	SD	0.2	0.3	0.4	0.6	0.6	0.4	0.4	0.4	0.1	0.4	0.4		0.4	0.4	
SD-T24	8	-5.1	-1.1	-2.4	-2.0	-9.1	-6.9	-6.4	-15.3	-10.8	-17.0	-11.1	-14.3	-8.0	-14.7	
SD-T24	8	-5.2	-1.4	-3.0	-2.3	-8.8	-6.9	-6.6	-14.8	-11.5	-17.1	-11.1	-14.8	-8.6	-15.3	
SD-T24	mean	-5.1	-1.2	-2.7	-2.2	-8.9	-6.9	-6.5	-15.1	-11.1	-17.1	-11.1	-14.6	-8.3	-15.0	-8.1
	SD	0.1	0.1	0.4	0.2	0.2	0.1	0.1	0.4	0.4	0.1	0.0	0.4	0.4	0.4	
SD-T24	9	-6.9		-3.7	-1.3	-9.7	-7.3	-7.3	-14.1	-10.0	-16.1	-10.8	-13.0	-9.0	-16.0	
SD-T24	9	-7.0	-1.7	-4.2	-1.6	-10.4	-8.0	-6.8	-14.6	-10.3	-16.7	-11.0	-13.7	-9.6	-16.7	
SD-T24	mean	-6.9	-1.7	-3.9	-1.4	-10.0	-7.7	-7.1	-14.3	-10.2	-16.4	-10.9	-13.3	-9.3	-16.3	-8.4
	SD	0.1		0.4	0.2	0.5	0.5	0.4	0.4	0.2	0.4	0.2	0.5	0.4	0.5	
SD-T24	10	-10.4	-4.0	-11.2	-4.0	-12.3	-16.2	-11.1	-16.9	-13.5	-19.3	-13.5	-16.5	-12.3	-18.3	
SD-T24	10	-10.4	-3.6	-10.6	-3.8	-11.7	-15.7	-10.6	-17.1	-13.0	-18.9	-13.0	-16.6	-11.9	-18.2	
SD-T24	mean	-10.4	-3.8	-10.9	-3.9	-12.0	-16.0	-10.8	-17.0	-13.3	-19.1	-13.2	-16.6	-12.1	-18.2	-12.1
	SD	0.0	0.3	0.4	0.1	0.5	0.3	0.4	0.2	0.4	0.3	0.3	0.1	0.3	0.1	
SI-T32	1	-7.0	-1.3	-5.2	-1.9	-12.9	-12.0	-8.7	-14.8	-12.4	-17.7	-12.2		-11.0	-16.5	
SI-T32	1	-7.3	-1.5	-5.5	-2.0	-13.8	-12.4	-9.2	-14.8	-12.4	-17.5	-11.6	-14.3	-10.8	-16.8	
SI-T32	mean	-7.2	-1.4	-5.4	-2.0	-13.3	-12.2	-8.9	-14.8	-12.4	-17.6	-11.9	-14.3	-10.9	-16.6	-9.6
	SD	0.2	0.1	0.2	0.0	0.6	0.3	0.4	0.0	0.0	0.1	0.4		0.2	0.2	
SI-T32	2	-6.7	-2.1	-5.5	-4.2	-14.9	-12.8	-8.8	-14.0	-10.4	-16.5	-10.7	-14.9	-9.0	-15.5	
SI-T32	2	-6.7	-1.4	-5.4	-4.0	-14.1	-12.9	-8.9	-14.3	-10.3	-16.5	-10.2	-15.5	-8.8	-14.8	
SI-T32	mean	-6.7	-1.7	-5.5	-4.1	-14.5	-12.9	-8.8	-14.1	-10.4	-16.5	-10.5	-15.2	-8.9	-15.2	-9.4
	SD	0.0	0.4	0.1	0.2	0.6	0.1	0.1	0.2	0.1	0.0	0.3	0.4	0.1	0.5	
SI-T32	3	-5.4		-4.5	-3.9	-14.4	-9.8	-8.4	-13.9	-9.8	-15.8	-9.6	-14.1	-8.5	-14.9	
SI-T32	3	-5.4	-0.7	-4.0	-4.6	-14.1	-9.8	-8.6	-14.0	-9.0	-16.2	-10.1	-14.2	-7.9	-14.2	
SI-T32	mean	-5.4	-0.7	-4.2	-4.2	-14.3	-9.8	-8.5	-13.9	-9.4	-16.0	-9.8	-14.1	-8.2	-14.6	-8.6
	SD	0.0		0.4	0.5	0.2	0.0	0.2	0.1	0.6	0.3	0.3	0.1	0.4	0.5	
SI-T32	4	-6.8	-2.1	-4.3	-3.2	-13.8	-11.3	-7.6	-13.8	-10.0	-16.7	-9.3	-13.2	-7.8	-14.5	
SI-T32	4	-6.4		-4.4	-3.6	-14.3	-11.1	-7.6	-13.8	-9.4	-16.7	-9.9	-13.7	-8.0	-14.8	
SI-T32	mean	-6.6	-2.1	-4.4	-3.4	-14.0	-11.2	-7.6	-13.8	-9.7	-16.7	-9.6	-13.4	-7.9	-14.7	-8.7
	SD	0.3		0.1	0.3	0.4	0.1	0.0	0.0	0.4	0.0	0.4	0.4	0.1	0.3	
SI-T32	5	-8.1	-1.8	-6.3	-1.4	-14.4	-12.5	-9.0	-15.0	-10.7	-16.5	-11.7	-13.5	-7.2	-14.9	
SI-T32	5	-7.8	-1.5	-5.9	-2.0	-13.5	-12.0	-8.5	-14.4	-10.7	-16.2	-11.4	-13.4	-7.7	-14.7	
SI-T32	mean	-7.9	-1.7	-6.1	-1.7	-13.9	-12.3	-8.7	-14.7	-10.7	-16.3	-11.5	-13.5	-7.5	-14.8	-9.2
	SD	0.2	0.2	0.2	0.4	0.6	0.4	0.3	0.4	0.0	0.3	0.2	0.1	0.4	0.2	
SI-T32	6	-8.2	-3.0	-6.0	-4.1	-10.2	-9.9	-7.2	-15.0	-10.9	-16.1	-10.6	-14.4	-8.5	-15.1	
SI-T32	6	-8.7	-2.3	-5.7	-3.4	-10.5	-10.5	-7.7	-14.9	-11.3	-16.8	-10.1	-14.4	-8.0	-14.3	
SI-T32	mean	-8.4	-2.7	-5.8	-3.7	-10.3	-10.2	-7.5	-15.0	-11.1	-16.5	-10.3	-14.4	-8.2	-14.7	-9.2
	SD	0.4	0.5	0.2	0.5	0.2	0.4	0.3	0.1	0.3	0.5	0.3	0.0	0.4	0.5	

SI-T32	7	-10.3	-2.8	-6.3	-3.2	-12.6	-12.6	-9.0	-16.0	-9.9	-17.8	-11.6	-14.1	-9.0	-15.4	
SI-T32	7	-10.0	-2.4	-6.1	-3.0	-11.8	-12.9	-8.9	-15.7	-10.7	-17.5	-11.1	-14.0	-9.0	-15.8	
SI-T32	mean	-10.2	-2.6	-6.2	-3.1	-12.2	-12.7	-9.0	-15.8	-10.3	-17.6	-11.3	-14.0	-9.0	-15.6	-9.9
	SD	0.2	0.3	0.2	0.2	0.6	0.2	0.1	0.3	0.5	0.2	0.4	0.1	0.0	0.3	
SI-T32	8	-9.8	-3.0	-6.3	-3.7	-13.0	-11.4	-9.0	-16.1	-11.0	-17.7	-11.9	-14.6	-9.2	-16.1	
SI-T32	8	-9.7	-2.4	-7.0	-3.4	-13.2	-11.8	-8.5	-15.7	-11.2	-17.7	-11.3	-15.4	-9.4	-16.4	
SI-T32	mean	-9.7	-2.7	-6.6	-3.5	-13.1	-11.6	-8.7	-15.9	-11.1	-17.7	-11.6	-15.0	-9.3	-16.2	-10.1
	SD	0.0	0.5	0.5	0.2	0.1	0.3	0.4	0.3	0.2	0.1	0.4	0.6	0.1	0.2	
SI-T32	9	-6.8	-0.2	-5.0	-2.9	-11.6	-9.6	-7.3	-15.0	-10.5	-17.2	-11.0	-15.7	-8.7	-15.7	
SI-T32	9	-6.6	-0.9	-4.7	-3.8	-12.5	-9.1	-7.6	-15.2	-10.0	-16.9	-10.7	-15.7	-8.6	-15.5	
SI-T32	mean	-6.7	-0.5	-4.9	-3.4	-12.1	-9.4	-7.4	-15.1	-10.2	-17.1	-10.8	-15.7	-8.7	-15.6	-8.9
	SD	0.1	0.5	0.2	0.6	0.7	0.3	0.2	0.1	0.4	0.3	0.2	0.0	0.1	0.1	
SI-T32	10	-8.7	-1.2	-5.3	-3.1	-12.9	-10.0	-7.7	-15.3	-11.0	-17.3	-10.9	-15.3	-9.0	-15.5	
SI-T32	10	-8.7	-0.9	-4.7	-2.2	-13.9	-10.6	-8.3	-14.9	-10.4	-17.0	-11.0	-14.9	-9.5	-15.3	
SI-T32	mean	-8.7	-1.0	-5.0	-2.7	-13.4	-10.3	-8.0	-15.1	-10.7	-17.1	-10.9	-15.1	-9.3	-15.4	-9.2
	SD	0.0	0.2	0.4	0.6	0.7	0.4	0.4	0.2	0.4	0.2	0.1	0.3	0.4	0.2	
SE-T3	1	-9.6	-2.7	-6.3	-4.1	-12.4	-12.3	-8.5	-15.8	-11.3	-17.8	-11.7	-14.3	-9.1	-16.0	
SE-T3	1	-9.6	-2.4	-6.2	-3.6	-12.1	-11.9	-7.8	-15.7	-10.8	-17.3	-11.4	-13.8	-8.8	-15.6	
SE-T3	mean	-9.6	-2.6	-6.2	-3.8	-12.2	-12.1	-8.2	-15.7	-11.1	-17.5	-11.6	-14.0	-8.9	-15.8	-9.8
	SD	0.0	0.2	0.1	0.3	0.3	0.2	0.4	0.1	0.4	0.3	0.2	0.3	0.2	0.3	
SE-T3	2	-8.8	-2.7	-6.9	-3.3	-16.4	-12.7	-8.0	-16.0	-11.5	-17.2	-11.1	-13.5	-8.7	-15.5	
SE-T3	2	-9.1	-2.6	-6.4	-3.7	-17.1	-12.3	-8.3	-16.0	-11.6	-17.4	-11.6	-13.9	-8.7	-15.3	
SE-T3	mean	-9.0	-2.6	-6.6	-3.5	-16.7	-12.5	-8.1	-16.0	-11.5	-17.3	-11.4	-13.7	-8.7	-15.4	-9.9
	SD	0.3	0.0	0.3	0.3	0.5	0.3	0.2	0.0	0.1	0.2	0.3	0.3	0.0	0.2	
SE-T3	3	-9.2	0.8	-6.8	-1.4	-15.1	-10.9	-8.9	-15.7	-11.0	-17.5	-12.5	-14.7	-9.2	-16.2	
SE-T3	3	-8.9	0.4	-6.4	-1.8	-15.6	-10.9	-8.7	-15.8	-10.6	-17.2	-12.0	-14.0	-8.9	-16.3	
SE-T3	mean	-9.1	0.6	-6.6	-1.6	-15.4	-10.9	-8.8	-15.7	-10.8	-17.3	-12.2	-14.3	-9.0	-16.3	-9.5
	SD	0.2	0.3	0.3	0.3	0.3	0.0	0.2	0.1	0.3	0.3	0.3	0.5	0.2	0.1	
SE-T3	4	-9.1	-0.1	-7.5	-4.0	-16.5	-13.7	-10.5	-17.3	-11.7	-18.3	-12.8	-13.5	-9.8	-16.7	
SE-T3	4	-9.0	-0.4	-7.5	-3.3	-15.5	-13.2	-9.9	-17.0	-12.1	-18.8	-13.2	-13.9	-10.1	-17.1	
SE-T3	mean	-9.0	-0.2	-7.5	-3.7	-16.0	-13.5	-10.2	-17.1	-11.9	-18.5	-13.0	-13.7	-10.0	-16.9	-10.6
	SD	0.1	0.2	0.0	0.5	0.7	0.4	0.4	0.2	0.3	0.4	0.3	0.3	0.2	0.3	
SE-T3	5	-8.1	0.1	-5.1	-0.6	-9.8	-7.8	-7.0	-16.0	-10.8	-18.0	-13.0	-13.9	-9.0	-16.6	
SE-T3	5	-7.9	0.2	-5.5	-0.9	-10.2	-7.4	-7.1	-16.0	-10.8	-18.0	-12.8	-14.3	-8.8	-16.9	
SE-T3	mean	-8.0	0.2	-5.3	-0.7	-10.0	-7.6	-7.1	-16.0	-10.8	-18.0	-12.9	-14.1	-8.9	-16.8	-8.9
	SD	0.2	0.1	0.3	0.2	0.3	0.3	0.1	0.0	0.0	0.1	0.2	0.2	0.1	0.2	
SE-T3	6		-1.2	-6.2	-3.4	-15.9	-9.2	-9.7	-17.9	-12.6	-19.5	-14.5	-16.4	-10.5	-18.6	

SE-T3	6	-9.3	-1.8	-6.1	-3.9	-15.5	-9.5	-9.0	-17.3	-12.0	-18.9	-14.0	-15.9	-9.8	-18.0	
SE-T3	mean	-9.3	-1.5	-6.1	-3.7	-15.7	-9.4	-9.3	-17.6	-12.3	-19.2	-14.3	-16.2	-10.1	-18.3	-10.6
	SD		0.4	0.0	0.3	0.3	0.2	0.5	0.4	0.4	0.4	0.4	0.4	0.5	0.4	
SE-T3	7	-12.8	-3.7	-9.5	-4.6	-16.6	-14.3	-10.5	-19.7	-14.6	-20.9	-15.7	-17.5	-12.6	-19.3	
SE-T3	7	-12.6	-4.2	-9.2	-5.1	-16.4	-14.0	-10.7	-19.8	-14.9	-21.3	-16.0	-18.3	-12.7	-19.5	
SE-T3	mean	-12.7	-4.0	-9.4	-4.9	-16.5	-14.2	-10.6	-19.7	-14.7	-21.1	-15.9	-17.9	-12.6	-19.4	-12.8
	SD	0.1	0.3	0.2	0.4	0.1	0.2	0.2	0.1	0.2	0.3	0.2	0.5	0.0	0.2	
SE-T3	8	-12.7	-4.7	-10.8	-7.0	-16.0	-13.7	-12.0	-20.8	-16.0	-21.8	-16.0	-19.0	-14.8	-20.0	
SE-T3	8	-12.7	-5.3	-10.3	-7.7	-15.1	-13.3	-12.3	-20.5	-16.5	-21.9	-16.0	-19.0	-14.9	-20.1	
SE-T3	mean	-12.7	-5.0	-10.5	-7.3	-15.6	-13.5	-12.2	-20.7	-16.2	-21.9	-16.0	-19.0	-14.8	-20.1	-13.9
	SD	0.0	0.4	0.3	0.5	0.7	0.3	0.2	0.2	0.3	0.1	0.0	0.0	0.0	0.0	
SE-T3	9	-14.7	-6.0	-9.9	-7.0	-16.5	-14.0	-14.4	-21.4	-16.1	-22.3	-16.4	-19.4	-14.7	-20.8	
SE-T3	9	-14.4	-5.6	-10.5	-7.8	-16.8	-14.2	-14.2	-21.0	-15.7	-22.4	-16.5	-20.0	-14.5	-20.5	
SE-T3	mean	-14.6	-5.8	-10.2	-7.4	-16.7	-14.1	-14.3	-21.2	-15.9	-22.4	-16.5	-19.7	-14.6	-20.6	-14.5
	SD	0.2	0.3	0.4	0.5	0.2	0.1	0.1	0.3	0.3	0.1	0.1	0.4	0.1	0.2	
SE-T3	10	-14.7	-7.3	-10.6	-8.5	-16.7	-14.6	-13.0	-22.0	-16.2	-22.9	-16.2	-19.3	-15.4	-21.0	
SE-T3	10	-15.0	-7.4	-10.9	-9.2		-14.4	-13.0	-22.3	-16.3	-23.3	-16.6	-20.1	-16.0	-21.3	
SE-T3	mean	-14.8	-7.4	-10.7	-8.8	-16.7	-14.5	-13.0	-22.2	-16.3	-23.1	-16.4	-19.7	-15.7	-21.2	-15.0
	SD	0.2	0.1	0.2	0.5		0.1	0.0	0.2	0.0	0.3	0.3	0.5	0.4	0.3	
SF-T4	1	-11.7	-2.9	-8.9	-4.3	-13.0	-14.3	-10.3	-17.4	-12.1	-19.7	-12.8	-17.8	-11.9	-18.0	
SF-T4	1	-11.9	-2.7	-9.0	-4.6	-14.0	-14.3	-10.1	-17.2	-12.6	-19.5	-12.8	-18.0	-11.5	-18.3	
SF-T4	mean	-11.8	-2.8	-9.0	-4.4	-13.5	-14.3	-10.2	-17.3	-12.3	-19.6	-12.8	-17.9	-11.7	-18.1	-11.8
	SD	0.1	0.2	0.1	0.2	0.7	0.0	0.1	0.1	0.3	0.1	0.0	0.2	0.2	0.2	
SF-T4	2	-9.0	-0.8	-6.6	-0.7	-10.3	-9.9	-7.5	-14.0	-10.3	-15.5	-9.2	-12.4	-7.3	-13.1	
SF-T4	2	-8.8	-0.7	-7.0	-1.5	-9.5	-10.2	-7.0	-13.2	-9.9	-15.4	-9.0	-12.3	-7.8	-13.3	
SF-T4	mean	-8.9	-0.8	-6.8	-1.1	-9.9	-10.0	-7.2	-13.6	-10.1	-15.4	-9.1	-12.4	-7.6	-13.2	-8.4
	SD	0.2	0.1	0.3	0.5	0.6	0.3	0.4	0.5	0.3	0.1	0.1	0.1	0.4	0.2	
SF-T4	3	-9.4		-7.0	-2.3	-13.6	-10.3	-9.7	-15.6	-11.8	-17.8	-12.0	-14.6	-10.1	-16.7	
SF-T4	3	-9.7	0.2	-6.9	-1.6	-13.1	-10.5	-10.1	-15.3	-11.1	-17.6	-11.7	-14.5	-9.7	-16.3	
SF-T4	mean	-9.6	0.2	-6.9	-1.9	-13.3	-10.4	-9.9	-15.5	-11.5	-17.7	-11.8	-14.6	-9.9	-16.5	-9.8
	SD	0.2		0.0	0.5	0.4	0.1	0.3	0.2	0.4	0.2	0.2	0.0	0.3	0.3	
SF-T4	4	-9.4	-3.4	-8.1	-2.3	-13.5	-10.0	-10.2	-15.6	-12.2	-18.2	-11.6	-14.1	-10.7	-16.6	
SF-T4	4	-9.7	-3.2	-7.7	-2.0	-14.1	-10.5	-10.5	-15.4	-11.6	-17.7	-11.1	-14.2	-10.7	-16.5	
SF-T4	mean	-9.5	-3.3	-7.9	-2.1	-13.8	-10.2	-10.4	-15.5	-11.9	-17.9	-11.4	-14.2	-10.7	-16.6	-10.5
	SD	0.2	0.1	0.3	0.2	0.4	0.4	0.2	0.2	0.4	0.3	0.3	0.1	0.0	0.1	
SF-T4	5	-10.4	-3.4	-7.8	-3.5	-16.6	-12.0	-11.9	-15.6	-10.8	-17.8	-11.9	-14.1	-11.0	-16.6	
SF-T4	5	-10.8	-3.2	-8.2	-3.7	-15.6	-12.7	-12.2	-16.0	-11.4	-18.1	-12.3	-14.4	-11.5	-17.0	

SF-T4	mean	-10.6	-3.3	-8.0	-3.6	-16.1	-12.3	-12.0	-15.8	-11.1	-18.0	-12.1	-14.3	-11.3	-16.8	-11.1
	SD	0.3	0.1	0.3	0.1	0.7	0.5	0.3	0.3	0.4	0.2	0.3	0.2	0.3	0.3	
SF-T4	6	-7.1	-1.8	-4.1	-1.6	-15.6	-9.1	-8.7	-14.4	-9.1	-17.0	-10.3	-11.7	-7.4	-14.3	
SF-T4	6	-6.7	-2.6	-4.7	-0.7	-14.7	-9.5	-8.0	-14.5	-9.4	-16.8	-10.3	-12.2	-7.6	-14.6	
SF-T4	mean	-6.9	-2.2	-4.4	-1.1	-15.2	-9.3	-8.3	-14.4	-9.3	-16.9	-10.3	-12.0	-7.5	-14.4	-8.5
	SD	0.3	0.5	0.4	0.6	0.7	0.3	0.5	0.1	0.2	0.2	0.0	0.3	0.1	0.2	
SF-T4	7	-10.4	-2.3	-9.3	-2.7	-14.2	-15.0	-10.0	-16.7	-12.0	-18.9	-12.0	-16.5	-11.6	-16.9	
SF-T4	7	-10.0	-2.2	-8.6	-2.0	-14.8	-14.3	-10.3	-16.6	-12.2	-18.5	-11.8	-15.7	-11.7	-17.4	
SF-T4	mean	-10.2	-2.2	-9.0	-2.4	-14.5	-14.7	-10.1	-16.7	-12.1	-18.7	-11.9	-16.1	-11.7	-17.1	-11.2
	SD	0.3	0.1	0.5	0.5	0.4	0.5	0.2	0.1	0.2	0.2	0.1	0.5	0.1	0.3	
SF-T4	8	-6.9		-5.7	-0.3	-10.2	-11.3	-8.1	-14.0	-10.3	-17.0	-10.2	-13.7	-9.1	-15.1	
SF-T4	8	-7.2	0.9	-5.5	-0.5	-9.5	-10.8	-7.6	-14.4	-9.8	-17.0	-9.6	-12.9	-8.7	-15.4	
SF-T4	mean	-7.1	0.9	-5.6	-0.4	-9.8	-11.0	-7.8	-14.2	-10.1	-17.0	-9.9	-13.3	-8.9	-15.3	-8.5
	SD	0.2		0.2	0.1	0.5	0.3	0.3	0.3	0.3	0.0	0.4	0.5	0.3	0.2	
SF-T4	9	-7.7	0.8	-5.2	-0.4	-11.8	-10.7	-7.8	-14.6	-11.8	-17.6	-10.7	-13.8	-9.3	-16.4	
SF-T4	9	-7.1	0.7	-4.9	-1.1	-10.8	-11.0	-7.3	-14.4	-11.4	-17.3	-10.8	-14.0	-9.7	-16.2	
SF-T4	mean	-7.4	0.8	-5.0	-0.8	-11.3	-10.8	-7.6	-14.5	-11.6	-17.5	-10.7	-13.9	-9.5	-16.3	-8.8
	SD	0.4	0.1	0.2	0.4	0.7	0.2	0.3	0.1	0.3	0.2	0.1	0.1	0.3	0.1	
SF-T4	10	-7.0	-1.9	-3.0	-1.7	-9.1	-6.7	-6.2	-13.0	-9.7	-15.9	-11.8	-14.0	-7.9	-14.6	
SF-T4	10	-6.6	-2.3	-3.5	-2.2	-9.8	-7.2	-6.6	-13.6	-9.5	-16.5	-11.4	-13.9	-8.4	-15.1	
SF-T4	mean	-6.8	-2.1	-3.3	-2.0	-9.5	-7.0	-6.4	-13.3	-9.6	-16.2	-11.6	-14.0	-8.2	-14.8	-8.0
	SD	0.3	0.3	0.4	0.3	0.5	0.3	0.2	0.4	0.1	0.4	0.2	0.1	0.3	0.4	
SI-T3	1	-8.4	-0.9	-4.6	-0.6	-11.8	-9.8	-8.2	-13.1	-8.7	-15.9	-10.9	-14.4	-8.5	-14.8	
SI-T3	1	-9.1	-0.7	-4.5	-0.9	-10.8	-10.0	-8.1	-14.0	-9.6	-16.8	-11.2	-15.0	-9.4	-15.4	
SI-T3	mean	-8.7	-0.8	-4.5	-0.8	-11.3	-9.9	-8.1	-13.6	-9.1	-16.3	-11.1	-14.7	-8.9	-15.1	-8.6
	SD	0.5	0.1	0.1	0.2	0.7	0.2	0.1	0.6	0.6	0.6	0.2	0.4	0.6	0.4	
SI-T3	2	-8.9	-1.6	-5.8	-0.9	-11.4	-11.4	-9.1	-14.3	-9.9	-16.7	-11.7	-14.2	-9.5	-15.9	
SI-T3	2	-9.5	-0.9	-5.5	-1.1	-10.7	-10.9	-9.1	-14.7	-10.3	-17.0	-12.0	-14.5	-9.9	-16.5	
SI-T3	mean	-9.2	-1.2	-5.6	-1.0	-11.1	-11.2	-9.1	-14.5	-10.1	-16.9	-11.8	-14.3	-9.7	-16.2	-9.3
	SD	0.4	0.5	0.2	0.1	0.5	0.3	0.0	0.3	0.3	0.2	0.2	0.2	0.3	0.4	
SI-T3	3	-9.0	-0.1	-6.6	-1.7	-12.9	-10.4	-9.1	-15.0	-11.2	-17.5	-11.9	-14.7	-10.0	-16.5	
SI-T3	3	-9.4	-0.2	-6.3	-1.6	-11.8	-10.5	-9.1	-15.3	-11.4	-17.8	-12.3	-14.7	-9.9	-16.8	
SI-T3	mean	-9.2	-0.2	-6.5	-1.6	-12.4	-10.5	-9.1	-15.2	-11.3	-17.6	-12.1	-14.7	-10.0	-16.7	-9.6
	SD	0.3	0.1	0.2	0.1	0.7	0.1	0.0	0.2	0.1	0.2	0.3	0.0	0.1	0.2	
SI-T3	4	-8.6	-0.4	-5.2	-0.8	-12.6	-11.8	-8.7	-14.7	-11.4	-17.4	-11.7	-15.5	-9.5	-15.9	
SI-T3	4	-8.8	-0.1	-5.2	-1.5	-11.6	-11.5	-8.7	-15.1	-10.9	-17.7	-12.0	-15.2	-9.8	-16.4	
SI-T3	mean	-8.7	-0.2	-5.2	-1.2	-12.1	-11.6	-8.7	-14.9	-11.1	-17.5	-11.8	-15.4	-9.6	-16.2	-9.3

	SD	0.2	0.2	0.0	0.5	0.7	0.2	0.0	0.3	0.4	0.2	0.2	0.2	0.3	0.3	
SI-T3	5	-10.8	-3.9	-9.3	-2.8	-13.0	-12.7	-9.7	-15.8	-11.6	-18.0	-12.1	-14.7	-10.3	-16.7	
SI-T3	5	-11.1	-3.8	-8.9	-3.2	-13.7	-12.7	-9.9	-15.7	-12.2	-18.4	-12.7	-15.3	-10.8	-17.3	
SI-T3	mean	-11.0	-3.8	-9.1	-3.0	-13.3	-12.7	-9.8	-15.7	-11.9	-18.2	-12.4	-15.0	-10.6	-17.0	-11.0
	SD	0.2	0.0	0.2	0.3	0.5	0.0	0.1	0.0	0.4	0.2	0.4	0.4	0.3	0.4	
SI-T3	6	-11.7	-2.2	-9.0	-2.6	-13.5	-13.9	-13.0	-16.3	-11.8	-18.3	-13.1	-15.1	-10.8	-16.7	
SI-T3	6	-12.1	-2.9	-8.7	-2.6	-12.5	-13.9	-13.1	-16.4	-11.6	-18.7	-12.5	-15.1	-11.4	-17.0	
SI-T3	mean	-11.9	-2.5	-8.8	-2.6	-13.0	-13.9	-13.0	-16.3	-11.7	-18.5	-12.8	-15.1	-11.1	-16.8	-11.4
	SD	0.3	0.4	0.3	0.0	0.7	0.0	0.1	0.1	0.1	0.3	0.4	0.0	0.4	0.3	
SI-T3	7	-10.8	-2.1	-7.7	-2.4	-11.3	-11.7	-11.7	-15.4	-11.4	-17.6	-12.7	-15.3	-11.3	-16.3	
SI-T3	7	-10.8	-2.1	-8.1	-2.1	-11.0	-12.3	-12.3	-15.4	-12.0	-18.2	-13.1	-15.4	-11.4	-16.7	
SI-T3	mean	-10.8	-2.1	-7.9	-2.2	-11.2	-12.0	-12.0	-15.4	-11.7	-17.9	-12.9	-15.3	-11.3	-16.5	-10.7
	SD	0.0	0.0	0.3	0.2	0.2	0.5	0.4	0.0	0.4	0.4	0.3	0.1	0.1	0.3	
SI-T3	8	-9.4	-1.1	-7.2	-1.0	-11.3	-11.6	-11.7	-15.0	-11.7	-17.7	-11.8	-14.2	-10.3	-16.6	
SI-T3	8	-9.5	-0.7	-7.6	-1.7	-10.4	-12.0	-12.0	-15.0	-11.6	-18.0	-11.3	-14.6	-9.7	-16.5	
SI-T3	mean	-9.5	-0.9	-7.4	-1.3	-10.8	-11.8	-11.8	-15.0	-11.6	-17.9	-11.6	-14.4	-10.0	-16.6	-10.1
	SD	0.1	0.3	0.3	0.5	0.6	0.3	0.2	0.0	0.1	0.2	0.3	0.3	0.4	0.1	
SI-T3	9	-9.2	-2.4	-6.9	-1.1	-11.5	-12.5	-9.5	-14.7	-10.0	-17.3	-11.5	-15.2	-9.4	-15.9	
SI-T3	9	-9.7	-2.1	-6.5	-1.3	-12.6	-12.9	-9.9	-14.5	-10.2	-17.6	-12.1	-15.2	-10.0	-16.3	
SI-T3	mean	-9.4	-2.3	-6.7	-1.2	-12.1	-12.7	-9.7	-14.6	-10.1	-17.5	-11.8	-15.2	-9.7	-16.1	-9.8
	SD	0.4	0.2	0.3	0.1	0.7	0.3	0.3	0.2	0.2	0.2	0.4	0.0	0.4	0.2	
SI-T3	10	-8.6	-2.2	-5.9	-1.3	-11.9	-10.5	-8.5	-15.1	-11.1	-17.5	-12.5	-15.0	-9.7	-16.2	
SI-T3	10	-8.4	-1.5	-6.1	-1.2	-11.1	-10.9	-8.7	-15.2	-11.2	-17.8	-12.4	-15.3	-10.0	-16.6	
SI-T3	mean	-8.5	-1.9	-6.0	-1.2	-11.5	-10.7	-8.6	-15.2	-11.1	-17.6	-12.4	-15.1	-9.8	-16.4	-9.6
	SD	0.1	0.5	0.1	0.1	0.6	0.3	0.1	0.1	0.1	0.2	0.1	0.2	0.2	0.4	

Table C.2. Summary of mean within-individual amino acid δ^{13} C (‰) values for hair keratin.

	Essent	ial Amin	o Acids				Non-Essential Amino Acids								
Individual	Phe	Val	Leu	Ile	Lys	Thr	Ala	Ser	Gly	Asx	Glx	Pro	Arg	Tyr	MB*
SI-T74															
Mean	-19.3	-16.8	-19.1	-12.7	-13.6	-3.6	-11.8	-2.6	-10.4	-9.2	-7.2	-12.1	-12.8	-18.1	-11.4
SD	1.5	0.8	1.0	1.3	1.4	0.9	1.6	1.7	1.5	1.1	1.8	2.2	1.3	1.0	1.0

Range	4.3	2.8	3.2	4.2	4.4	3.2	6.1	4.3	4.3	3.7	6.3	6.8	4.2	3.2	3.3
Min	-21.3	-18.5	-20.4	-14.5	-16.0	-5.5	-14.1	-5.0	-12.9	-10.4	-10.1	-15.2	-14.7	-19.9	-12.5
Max	-17.0	-15.7	-17.2	-10.3	-11.6	-2.3	-8.0	-0.7	-8.6	-6.7	-3.8	-8.4	-10.5	-16.7	-9.2
SD-T24															
Mean	-16.6	-15.8	-17.9	-11.8	-11.6	-3.2	-11.2	-3.5	-11.4	-8.9	-7.1	-10.5	-10.8	-15.2	-10.5
SD	1.6	1.2	1.4	1.6	1.4	1.4	3.4	2.1	1.6	2.4	2.8	3.3	2.5	1.4	1.9
Range	5.3	3.9	4.4	5.2	4.4	4.0	9.1	7.5	5.3	8.4	8.9	9.4	8.3	4.2	5.9
Min	-19.0	-17.6	-19.8	-14.1	-13.2	-4.9	-16.0	-8.7	-14.2	-13.5	-11.6	-15.9	-15.6	-17.5	-14.0
Max	-13.7	-13.7	-15.4	-8.9	-8.8	-0.9	-6.9	-1.2	-8.9	-5.1	-2.7	-6.5	-7.3	-13.3	-8.1
SI-T32															
Mean	-15.3	-14.8	-16.9	-10.6	-10.8	-3.2	-11.3	-1.7	-13.1	-7.8	-5.4	-8.3	-8.8	-14.5	-9.3
SD	0.7	0.7	0.6	0.8	0.8	0.8	1.3	0.8	1.3	1.5	0.8	0.6	1.0	0.8	0.5
Range	2.0	2.1	1.7	3.0	2.3	2.5	3.5	2.2	4.2	4.8	2.4	1.6	3.4	2.3	1.5
Min	-16.6	-15.9	-17.7	-12.4	-11.9	-4.2	-12.9	-2.7	-14.5	-10.2	-6.6	-9.0	-10.9	-15.7	-10.1
Max	-14.6	-13.8	-16.0	-9.4	-9.6	-1.7	-9.4	-0.5	-10.3	-5.4	-4.2	-7.4	-7.5	-13.4	-8.6
SE-T3															
Mean	-18.1	-18.2	-19.6	-13.2	-14.0	-4.5	-12.2	-2.8	-15.2	-10.9	-7.9	-10.2	-11.3	-16.2	-11.6
SD	2.1	2.5	2.3	2.3	2.0	2.6	2.3	2.7	2.2	2.5	2.1	2.3	2.8	2.6	2.3
Range	5.8	6.5	5.8	5.5	5.1	8.1	6.9	8.0	6.7	6.8	5.4	7.2	7.0	6.0	6.1
Min	-21.2	-22.2	-23.1	-16.3	-16.5	-8.8	-14.5	-7.4	-16.7	-14.8	-10.7	-14.3	-15.7	-19.7	-15.0
Max	-15.4	-15.7	-17.3	-10.8	-11.4	-0.7	-7.6	0.6	-10.0	-8.0	-5.3	-7.1	-8.7	-13.7	-8.9
SF-T4															
Mean	-15.9	-15.1	-17.5	-11.0	-11.2	-2.0	-11.0	-1.5	-12.7	-8.9	-6.6	-9.0	-9.7	-14.3	-9.7
SD	1.5	1.3	1.2	1.1	1.1	1.3	2.3	1.6	2.4	1.8	2.0	1.8	1.6	1.7	1.4
Range	4.9	4.0	4.2	3.0	3.7	4.0	7.7	4.2	6.6	5.0	5.7	5.6	4.2	5.9	3.8
Min	-18.1	-17.3	-19.6	-12.3	-12.8	-4.4	-14.7	-3.3	-16.1	-11.8	-9.0	-12.0	-11.7	-17.9	-11.8
Max	-13.2	-13.3	-15.4	-9.3	-9.1	-0.4	-7.0	0.9	-9.5	-6.8	-3.3	-6.4	-7.5	-12.0	-8.0
SI-T3															
Mean	-16.4	-15.0	-17.6	-11.0	-12.1	-1.6	-11.7	-1.6	-11.9	-9.7	-6.8	-10.0	-10.1	-14.9	-9.9
SD	0.5	0.7	0.6	0.9	0.6	0.7	1.2	1.1	0.8	1.1	1.5	1.7	0.7	0.4	0.9
Range	1.9	2.7	2.2	2.8	1.8	2.2	4.0	3.6	2.5	3.4	4.6	4.9	2.4	1.1	2.8
Min	-17.0	-16.3	-18.5	-11.9	-12.9	-3.0	-13.9	-3.8	-13.3	-11.9	-9.1	-13.0	-11.3	-15.4	-11.4
Max	-15.1	-13.6	-16.3	-9.1	-11.1	-0.8	-9.9	-0.2	-10.8	-8.5	-4.5	-8.1	-8.9	-14.3	-8.6

*MB indicates calculated δ^{13} C mass balance values.