ORIGINAL ARTICLE OPEN ACCESS

Spatiotemporal Dynamics of Non-Ecological Speciation in Rubyspot Damselflies (*Hetaerina* spp.)

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Received: 3 January 2025 | Revised: 25 April 2025 | Accepted: 30 April 2025

Handling Editor: Angus Davison

Funding: This work was supported by a NERC Environmental Omics Facility Grant (NEOF1274) to J.D., funding from Durham University (including a Durham Doctoral Studentship to C.P.) and NSF DEB-NERC-2040883 to J.D. and G.F.G. Specimen collection was conducted under permit SGPA/ DGVS/04421/21 issued by the Mexican Secretaría de Medio Ambiente y Recursos Naturales (SEMARNAT) to L.M.C.; permit 08112112 issued by the Florida Department of Environmental Protection to J.D.; licence FD/WL/7/21(04) issued by the Forest Department in Belize to J.D. & G.F.G.; permit No. R-SINAC-SE-DT-PI-003-2021 issued by the Costa Rican Ministry of Environment (Minae) and genetic access permission No. 377 issued by the Comisión Institucional de Biodiversidad, Universidad de Costa Rica. C.P. was provided training and support for laboratory work by Gavin Horsburgh at the Natural Environment Research Council Omics Facility (NEOF) at the University of Sheffield. NEOF also provided C.P. with training and support with bioinformatics.

Keywords: ddRAD | Odonata | population genomics | speciation | Zygoptera

ABSTRACT

Non-ecological speciation is a common mode of speciation, which occurs when allopatric lineages diverge in the absence of pronounced ecological differences. Yet, relative to other speciation mechanisms, non-ecological speciation remains understudied. Numerous damselfly clades are characterised as non-adaptive radiations (the result of several rounds of non-ecological speciation without subsequent divergence), but there are few damselfly lineages for which we have a detailed understanding of the spatiotemporal dynamics of divergence. Recent phylogeographic analyses demonstrate that American rubyspot damsel-flies (*Hetaerina americana* sensu lato) actually comprise at least two cryptic lineages that coexist sympatrically across most of Mexico. To broaden our understanding of the dynamics of diversification to other rubyspot lineages, we investigated the phylogeographic history of smoky rubyspot damselflies (*Hetaerina titia*) using genomic data collected across Central and North America. Unexpectedly, we found evidence of reproductive isolation between the highly genetically differentiated Pacific and Atlantic lineages of *H. titia* in a narrow secondary contact zone on the Isthmus of Tehuantepec, Mexico. We then fit models of historical demography to both *H. americana* sensu lato and *H. titia* to place these comparisons in a temporal context. Our findings indicate that Pacific and Atlantic lineages of *H. titia* split more recently than the broadly sympatric lineages within *H. americana* sensu lato, supporting key assumptions of the non-ecological speciation model and demonstrating that these two pairs of sister lineages are at different stages of the speciation cycle.

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1 | Introduction

Speciation-the process by which a split in one lineage leads to two or more reproductively isolated lineages-is a key process contributing to the accumulation of biodiversity on Earth. Yet many aspects of the process by which one population transitions to two allopatric populations then, upon geographical reunification (secondary contact), becomes two sympatric species remain poorly understood (Tobias et al. 2020). There are multiple outcomes to the speciation cycle, from the admixture and collapse of the two diverging, ephemeral species back into one (Cutter and Gray 2016; van der Valk et al. 2021; Zou et al. 2022), to parapatric species with hybrid zones (Barton and Hewitt 1989; DeRaad et al. 2022; Irwin and Schluter 2022), to sympatric and reproductively isolated species. The outcome of secondary contact is primarily predicted by divergence time, but we lack a comprehensive understanding of how quickly intrinsic reproductive isolation can arise, generating nonephemeral species (Anderson et al. 2023; Matute and Cooper 2021).

Much research into the speciation process examines the role of natural selection in driving divergence between lineages via ecological speciation, where species divergence and extrinsic reproductive isolation are underpinned by adaptation to different ecological niches (Anderson et al. 2023; Nosil 2012; Rundle and Nosil 2005). When sustained over several bursts of speciation, ecological speciation leads to adaptive radiations such as the iconic Galápagos finches, Lake Victoria cichlids or Greater Antillean anoles, and has been the central focus of evolutionary biologists interested in explaining the origin and accumulation of biodiversity (Schluter 2000; Simpson 1944).

However, many speciation events lead to species without discernible ecological differentiation between daughter lineages. An alternative model of speciation is non-ecological speciation (Czekanski-Moir and Rundell 2019; Gittenberger 1991), where divergence between species is not primarily driven by natural selection but rather by the accumulation of reproductive isolation over time. Such isolation can result from intrinsic genomic incompatibilities that arise over time from genetic drift (Dion-Côté and Barbash 2017; Ravinet et al. 2017; Westram et al. 2022) or through divergence in reproductive traits (Arnegard et al. 2010; McEachin et al. 2022; Mendelson et al. 2014; Mendelson and Safran 2021; Okamoto and Grether 2013).

Despite the intense research focus on adaptive radiations, most clades have not diversified via adaptive radiation (Czekanski-Moir and Rundell 2019; Rundell and Price 2009). When sustained through several bouts of speciation without subsequent divergence (e.g., via character displacement), nonecological speciation can lead to a radiation characterised by minimal ecological differentiation between clade members, referred to as a non-adaptive radiation (Czekanski-Moir and Rundell 2019; Gittenberger 1991; Rundell and Price 2009). A recent analysis of insular radiations of birds (including several textbook examples) demonstrates that the majority of such radiations are non-adaptive (Illera et al. 2024). Indeed, examples of non-adaptive radiations are abundant (Czekanski-Moir and Rundell 2019) and likely to increase in frequency as genomics leads to the discovery of new cryptic species (Eme et al. 2018; Struck et al. 2018).

In addition to being common in nature, non-adaptive radiations offer compellingly simplified models for studying the diversification process. For biodiversity to accumulate in a given region, species must be able to both co-occur (e.g., via dispersal into a common area) and coexist (i.e., experience population growth) in one another's presence (Weir and Price 2011; Tobias et al. 2020). Non-adaptive radiations provide useful case studies for characterising the circumstances under which sister lineages attain range overlap in the absence of ecological differentiation.

Damselflies (Odonata, suborder Zygoptera) provide several iconic examples of non-adaptive radiations (Wellenreuther and Sánchez-Guillén 2016). According to the widely accepted conceptual model for diversification in damselflies, diversity accumulates via non-ecological speciation as species come into secondary sympatry after sufficient time has passed in allopatry for divergent lineages to become reproductively isolated via the evolution of species-specific genital morphology (e.g., male claspers and the [pro]thoracic plates of females which come into physical contact with male claspers during mating) (Paulson 1974; Wellenreuther and Sánchez-Guillén 2016). Consistent with this model, sympatric assemblages of congeners often exhibit little ecological differentiation (e.g., Calopteryx spp. [Svensson et al. 2018]; Ischnura spp. [Sánchez-Guillén, Córdoba-Aguilar, et al. 2014; Sánchez-Guillén et al. 2005]; Enallagma spp. [McPeek and Brown 2000]). Species do, however, possess reproductive characters that are highly divergent from those of other congeners (e.g., Calopteryx spp. [Svensson et al. 2010, 2014]; Ischnura spp. [Sánchez-Guillén, Córdoba-Aguilar, et al. 2014; Sánchez-Guillén et al. 2005]; Enallagma spp. [McPeek et al. 2009, 2011]). Yet, while these observations support the hypothesis that these damselfly genera are non-adaptive radiations, no study to date has reconstructed the temporal dynamics of reproductive isolation and secondary contact in damselflies.

Here, we investigate whether divergence time predicts the outcome of secondary contact (two allopatric lineages becoming geographically reunited) within a subset of damselfly species within the genus Hetaerina. Hetaerina damselflies have a crown age estimate of 36.2 million years ago (mya) (Standring et al. 2022) with most species living in sympatry with one or more congeners. There are currently 39 recognised Hetaerina species (Garrison 1990; Standring et al. 2022), but the recent discovery that Hetaerina americana sensu lato consists of at least two highly diverged and sympatric cryptic species (now named H. americana and Hetaerina calverti; Vega-Sánchez et al. 2020, 2024) suggests the number may be higher. The morphology of male claspers is the only way to identify some adult Hetaerina species in the field (Vega-Sánchez et al. 2020, 2024). All Hetaerina species are lotic habitat (stream, river) specialists and closely resemble one another in morphology, diet and reproductive behaviour, despite the wide diversity of forms and behaviours present in Odonata (Corbet 1999). Although Hetaerina spp. show moderate levels of climatic niche and microhabitat differentiation (Grether et al. 2024; McEachin et al. 2022), the ecological and phenotypic similarities between species are more remarkable than the differences considering their ancient divergence. Consequently, Hetaerina damselflies likely represent another example of a non-adaptively radiating damselfly clade.

We investigate two geographically widespread lineages of *Hetaerina* from across North and Central America: *H. americana* sensu lato (i.e., the *H. americana* and *H. calverti* species complex) and *H. titia*. *H. calverti* is found in sympatry with both the Northern and Southern lineages of *H. americana* (Vega-Sánchez et al. 2024). *H. titia* exhibits the largest latitudinal range of any *Hetaerina* species, extending from Canada in the north to Panama in the south (Grether et al. 2024; Paulson 2020). Phylogenies of *H. titia* constructed using mitochondrial and nuclear genes suggest divergence between populations that reside in Pacific and Atlantic drainages (Drury, Anderson, et al. 2019; Drury and Grether 2014). Together, these taxa offer a window into the process of non-ecological speciation.

Here, we use genome-wide markers from specimens across Central and North America to reconstruct the population-level relationships between distinct lineages within the species currently recognised as *H. americana*, *H. calverti and H. titia*. We then estimate the divergence times between these lineages to characterise the timescale of isolation and secondary sympatry in a non-adaptive radiation.

2 | Methods

2.1 | Sampling and Sequencing

Whole organism samples of smoky rubyspot (Hetaerina titia) and American rubyspot (H. americana sensu lato) damselflies were collected between 2006 and 2021 from across Central and North America, submerged in \geq 95% ethanol or RNALater (Invitrogen), and stored at $\leq -20^{\circ}$ C. For DNA extraction, approximately 2 mm³ of wing muscle tissue was removed from the thorax and processed using DNeasy Blood and Tissue Kits (Qiagen) following standard manufacturer protocols. To generate genome-wide sequence data, we followed double digest restriction enzyme associated DNA (ddRAD) protocols (DaCosta and Sorenson 2014; Franchini et al. 2017; Peterson et al. 2012). We used the restriction enzymes PstI and EcoR and generated multiplexed libraries by ligating adapters containing a region of four random nucleotides for PCR clone removal. After pairedend 150 bp sequencing on a NovaSeq 6000 (Illumina), we demultiplexed and filtered clones (see Data S1 for further details on library prep and the full bioinformatics pipelines is outlined in Figure S1 and Table S1). In total, we obtained sequence data for 205 individuals of H. titia and 58 individuals of H. americana sensu lato from across Central and North America.

2.2 | SNP Calling

Individual sequences were mapped to a *H. americana* reference genome (Grether et al. 2023) and to a *H. titia* reference genome (Patterson et al. 2024) using the Burrow-Wheeler aligner (bwa) *mem* alignment algorithm (Li and Durbin 2009). Genotype calling was done using *bcftools v1.13* (Danecek et al. 2021; Li 2011) using the *mpileup* and *call* commands.

The probability of any two samples having the same restriction site at a particular locus decreases with phylogenetic distance.

As such, multiple SNP libraries were constructed including varying combinations of species for use in different analyses (Table 1, Table S2). Firstly, two SNP libraries were produced that contained all Hetaerina samples (H. americana sensu lato and *H. titia*) and were mapped to either the draft genome of H. americana (Grether et al. 2023) or H. titia (Patterson et al. 2024). Additionally, four different SNP libraries were constructed, again using both draft genomes, for all H. americana sensu lato samples and, separately, all H. titia samples. Finally, we also conducted a de novo (reference-free) SNP assembly to determine if there was any ascertainment bias using draft genomes that were more closely related to either species or population within our sample set. To reduce computation time, we limited the de novo SNP library to 3 of the highest coverage samples from each identified lineage (18 samples in total) from the reference-mapped libraries. The de novo pipeline was constructed using ipyrad (Eaton and Overcast 2020) as outlined in the Supporting Information Methods. As recent introgression (<2 generations) violates assumptions of the phylogenetic and demographic analysis, we created additional sets of SNP libraries excluding the samples from a drainage where preliminary results suggested recent introgression between Atlantic and Pacific population clusters of *H. titia*. The full bioinformatics pipeline is outlined in the Supporting Information Methods, and all scripts are available on GitHub (https://github.com/ ChristophePatterson/Phylogeography-Hetaerina).

The resulting vcf files were imported into R using the package vcfR (Knaus and Grünwald 2017). Further SNP and sample filtering (Supporting Information Methods) and conversion of vcf into compatible formats for each analysis software were done using the R packages ape (Paradis and Schliep 2019), adegenet (Jombart 2008) and poppr (Kamvar et al. 2014). The total number of samples, species included, number of SNPs/loci and read alignment methodology used in each analysis are presented in Table S2.

2.3 | Species Delimitation and Population Structure

To characterise the population structure of *H. americana* sensu lato and *H. titia*, we used the R package LEA (Frichot and François 2015) to conduct principal components analysis (PCA) and non-negative matrix factorisation algorithms (sNMF) for least-squares estimates of ancestry proportions for each sample (Frichot et al. 2014). We restricted the SNPs to those that were biallelic and removed samples that had more than 20% missing data. To maintain equal levels of ploidy we removed SNPs mapped to the X chromosome, as *Hetaerina* has an XX/XO sex determination system (Patterson et al. 2024). In sNMF, we tested for a range of ancestral populations (K = 1 to 10) and plotted the mean cross-entropy values for 100 repetitions. We used hierfstat (Goudet 2005) to calculate F_{st} between each identified cluster.

2.4 | Phylogenetic Inference

We reconstructed phylogenetic trees for each reference mapped SNP library (Table 1) using RAxML/8.2.12 (Stamatakis 2014).

TABLE 1 Overview of all analyses and SNP/loci libraries used for each. Each library consists of different combinations of samples from different
species and reads were aligned to the draft genome of Hetaerina americana (HetAmer1.0, Grether et al. 2023), H. titia (HetTit1.0, Patterson et al. 2024)
or mapped <i>de novo</i> .

		Population structure	Phylogenies		Demographic/ divergence times			Introgression
Species included	Read alignment	sNMF & PCA	RAxML	SVD quartets	DelimitR	SNAPP	G- Phocs	Introgress
H. americana sensu lato & H. titia	HetAmer1.0	_	S	S	_	М	_	_
H. americana sensu lato & H. titia	HetTit1.0	—	S	S	_	М	—	—
<i>H. americana</i> sensu lato & <i>H. titia</i>	de novo	—	_	_	_	М	—	—
<i>H. americana</i> sensu lato	HetAmer1.0	М	М	S	S	—	S	_
<i>H. americana</i> sensu lato	HetTit1.0	S	S	S	S		S	_
<i>H. americana</i> sensu lato	de novo	_	—	—	—	—	М	_
H. titia	HetAmer1.0	S	S	S	S	_	S	_
H. titia	HetTit1.0	М	М	S	S	_	S	М
H. titia	de novo	_	—	_	—	—	М	

Note: Each analysis and library are marked as to whether the results are presented in the main text (M) or in the Supporting Information (S). *H. americana* sensu lato consists of three distinct lineages, including the recently described *H. calverti* (Vega-Sánchez et al. 2024). A breakdown of the number of samples and the SNP/loci number for each analysis is presented in Table S2.

As RAxML requires homozygous SNPs, we filtered the vcfs to include only homozygous-called sites, then excluded sites that were invariant across individuals after removing samples with < 20% missing data. Phylogenies were reconstructed under a general time reversible model (GTR), a gamma distribution of rate heterogeneity and a Lewis ascertainment correction due to the exclusion of invariant sites (-m=ASC_GTRGAMMA) (Devitt et al. 2019; Lozier et al. 2016).

We also reconstructed phylogenies in SVDquartets (Chifman and Kubatko 2014) in PAUP* (Wilgenbusch and Swofford 2003). Heterozygous sites, which are compatible with SVDquartet analysis, were retained (Table S2). We calculated the SVD score of 100,000 unrooted 4-'taxa' trees (quartets) and to infer the optimal phylogenetic relationship between the samples for each quartet, we used the Quartet FM method (Reaz et al. 2014). We then constructed a consensus tree by repeating the process 100 times to produce bootstrap support values for each tree node determined by the percentage of times the node was part of the consensus topology of the tree.

2.5 | Testing for Migration Between Lineages

To test the assumption of the non-ecological speciation hypothesis that little to no migration occurs between diverged lineages, we used the R package delimitR (Smith and Carstens 2020). delimitR uses site frequency spectrums (SFS) built from a SNP data set to predict the most likely demographic history for several potential populations or species. It then uses fastsimcoal2 (v2.6) (Excoffier et al. 2013, 2021) to simulate SFS for each specified demographic scenario under a range of priors and builds a random forest classifier to estimate the most likely demographic scenario for the observed data. For population clusters of *H. titia* and for *H. americana* sensu lato, we simulated each valid combination of several demographic scenarios with and without migration between populations (Figure S2). We simulated each scenario using broad, uniform priors (Supporting Information Methods).

Empirical SFS were calculated using the package easySFS (https://github.com/isaacovercast/easySFS) which builds off the dadi.Spectrum class from the software $\partial a \partial i$ (Gutenkunst et al. 2009). To take into account missing SNPs, which are inherent to ddRAD data, we projected down the SFS to maximise the number of segregating sites following Gutenkunst et al. (2009).

2.6 | Divergence Time Estimation

To place divergence among *Hetaerina* lineages within the broader context of the speciation cycle, we estimated the divergence times

of population clusters using two approaches. Firstly, we ran the Bayesian coalescent analysis SNAPP implemented within the programme Beast v2.7.5 (Bouckaert et al. 2019). Due to computational constraints, we restricted the analysis to four individuals per cluster identified by sNMF (24 individuals in total) with the highest SNP coverage from each distinct ancestral clustering identified by sNMF. We then removed SNPs that were either no longer polymorphic between the selected samples, genotyped in less than one individual from each population or mapped to the X chromosome. We used previously estimated divergence times from Standring et al. (2022) as priors by secondary calibration for divergence time between H. titia and H. americana sensu lato (mean = 33.08 million years ago (mya), standard deviation = 5.53 mya) and for the divergence of *H. americana* and *H. calverti* (mean = 3.76 mya, standard deviation = 1.87 mya). We used a starting tree that had the same relationships identified in RAxML and SVDquartets for each of the clusters and ran MCMC for 1,000,000 generations, sampling every 500 iterations. A SNAPP configuration file was created using a custom R script and the ruby script from https:// github.com/mmatschiner/snapp_prep. We assessed the convergence using tracer and calculated the maximum clade credibility tree, with a 10% burn-in removal, using TreeAnnotator v2.7.5 (Bouckaert et al. 2019).

For an alternative estimate of divergence times not based on a secondary calibration, we fit models of historical demography using G-PhoCS (Gronau et al. 2011) which uses a Bayesian coalescent approach. We present parameter estimates for the demographic models that were best supported by delimitR. We ran G-PhoCS using loci mapped using heterospecific draft genome, conspecific draft genome and loci mapped *de novo* for each species. We converted mutation rate-scaled parameter estimates of G-PhoCS into the number of diploid individuals and the number of years using 2.8e-9 mutations per base pair per generation (Keightley et al. 2014). We converted generations to years using an estimated generation time of 1 year. We present results from G-PhoCS using the loci mapped de novo in the main text as these libraries minimise ascertainment bias (see Supporting Information Methods for further detail).

2.7 | Investigating a Potential Secondary Contact Zone

Preliminary analysis identified an individual of H. titia with admixed ancestry from a site on the Isthmus of Tehuantepec, in Mexico. To determine the number of generations since the putative hybridisation event and see if any other individuals had admixed ancestry, we ran a hybridisation analysis using the R package introgress (Gompert and Buerkle 2010). We subset our data to samples from sites in and around the Isthmus of Tehuantepec. We calculated the allele frequency for each SNP for both Pacific and Atlantic populations, excluding samples from the drainage where the putative hybrid was identified. We then subset our dataset to 914 autosomal SNPs and 19 sex-linked SNPs that had an allele frequency difference < 0.8 between the Pacific and Atlantic, in line with DeRaad et al. (2022). We then assigned each allele to a 'parental' Pacific or Atlantic genotype and calculated both the percentage of Pacific and Atlantic alleles carried by each sample (the hybridisation index) and the average autosomal heterozygosity

across all highly divergent SNPs (the multi-allele heterozygosity) for each sample.

3 | Results

Nearly all analyses, using all different combinations of libraries, produced comparable results. For brevity, we summarise the results in the main text and present the result for each individual library in the Data S1. For an overview of which SNP/loci libraries were used in each analysis, see Table 1.

We retained sequence data for 259–263 samples of *H. americana* sensu lato and *H. titia* which had between 519 and 609 SNPs with adequate genotyping across all samples (Tables S2 and S3). For SNP libraries that only included *H. americana* sensu lato, we retained sequencing for 58 samples with 1816 to 5259 SNPs. For SNP libraries which only included *H. titia*, we retained sequencing for between 205 and 207 samples with 1122 to 3819 SNPs depending on read alignment methodology. Across all SNP libraries, we obtained an average coverage of between 50 and 53× and a median missing genotype rate of around 1.6% to 1.9%.

3.1 | Hetaerina titia Population Structure

sNMF admixture analyses and principal component analyses both identified three distinct clusters in *H. titia* (Figures 1 and 2, Figure S3): (1) a Caribbean and Southern Gulf of Mexico cluster, (2) a Northern Gulf of Mexico and Atlantic cluster and (3) a Pacific Coast cluster (Figures 1 and 2, Figures S3–S7). Hereafter, we refer to these three clusters as the Southern Atlantic H. titia cluster, the Northern Atlantic H. titia cluster and the Pacific H. titia cluster, respectively. The pairwise $F_{\rm st}$ values between the three groups indicate high levels of differentiation. Using the 3819 SNPs mapped to the *H. titia* draft genome, the $F_{\rm st}$ was 0.818 between the Pacific and Northern Atlantic, 0.730 between the Pacific and Southern Atlantic, and 0.521 between the Northern and Southern Atlantic. Further population genetic summary statistics are presented in Table S5. We identified one sample with extensive admixture between the Pacific and Southern Atlantic clusters from site CUAJ01 in Cuajinicuil, Oaxaca (16°47'24.00" N, 95°0'36.00" W) on the Gulf slope of the Isthmus of Tehuantepec (Mexico).

3.2 | *Hetaerina americana* Sensu Lato Population Structure

Consistent with previous work conducted on a different set of specimens with different restriction enzymes (Vega-Sánchez et al. 2020, 2024), analyses of *H. americana* sensu lato also grouped samples into three distinct clusters (Figures S8–S13). *Hetaerina calverti* forms one cluster, and *H. americana* is split into two distinct clusters—a Northern population in the continental United States and a Southern population found on both the Gulf and Pacific slopes of Mexico. We refer to these lineages as Northern *H. americana* and Southern *H. americana* going forward. Using SNPs mapped to the draft genome of *H. americana*, pairwise F_{st} values between the identified groups were 0.833 (*H. calverti* vs. Northern *H. americana*), 0.791 (*H. calverti* vs. Southern *H. americana*).

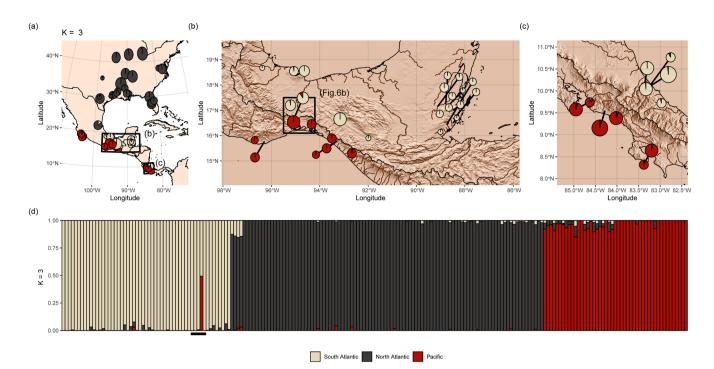


FIGURE 1 | Ancestry estimates for 205 *Hetaerina titia* with a dataset of 3819 unlinked biallelic autosomal SNPs. SNPs were generated by mapping ddRAD reads to the draft genome of *H. titia*. LEA was run for 20 repetitions and an alpha value of 100. (a) The mean estimate of ancestry proportion for all samples within each sample site of *H. titia* across Central and North America, (b) Isthmus of Tehuantepec and Belize and (c) Costa Rica. Within panels a, b and c, the area of each pie chart is proportional to the number of samples from each site and then coloured by the mean proportion of estimated ancestry (either South Atlantic, North Atlantic or Pacific) across all samples from each site. (d) Estimate of ancestry for each individual. Samples are ordered by drainage, then country and then latitude. Rivers and drainage basins from Hydrosheds. Topography data from the R package elevatr. The black boxes shown in panel (a) are the bounding areas for panels (b) and (c). The black box in panel (b) is the bounding box for Figure 6b and the five samples from the site with an identified hybrid individual are underlined in panel (d).

3.3 | Phylogenetic Inference of Hetaerina

In agreement with population structure analyses, populations of *H. titia*, which reside in drainages that flow into the Atlantic, including the Gulf of Mexico and the Caribbean, are more closely related to each other than populations that reside in drainages that flow into the Pacific. The Atlantic lineage is split into two groups: (1) samples that originated from the continental United States and the most Northern sample site in Mexico, and (2) the remaining samples from Mexico, Belize and Costa Rica (Figure 3). Within the Pacific *H. titia* lineage, there are three distinct groups, one group from Costa Rica and two separate Central and Southern lineages in Mexico (Figure 3, Figure S14).

Within the *H. americana* sensu lato lineage there is a distinct split between populations in the continental United States and populations in Mexico. Unlike *H. titia* lineages, neither *H. americana* sensu lato lineage is restricted to either Pacific or Atlantic drainages—the *H. americana* south and *H. calverti* lineages ranges broadly overlap and are commonly found coexisting sympatrically (Figures S10, S11 and S15).

Key inferences from SVDquartet analyses were qualitatively similar to those derived from RAxML (Figures S16 and S17).

3.4 | Tests for Migration Between Lineages

The best-supported demographic scenarios for *H. titia* suggest that Pacific and Atlantic lineages are completely isolated, with no evidence of ancient or contemporary migration between them. The best-fit demographic scenarios did contain ancestral—but not contemporary—gene flow between the Northern and Southern Atlantic lineages (Model 13 in Figure S2, receiving 85.3% of support). There was some support for the demographic scenarios with no migration, neither contemporary nor ancestral, between any lineage (Model 5 in Figure S2, receiving 14.3% of support). The outof-the-bag error rate varied among *H. titia* demographic scenarios but was low for models 5 and 13 (20% and 10%). Furthermore, incorrect classifications of models 5 and 13 were limited to the alternative of these two scenarios. No other demographic scenarios for *H. titia* received more than 2% support (Table S4).

Similarly, for *H. americana* and *H. calverti*, models suggest no ancient or contemporary migration between *H. americana* and *H. calverti*. The most favoured model had three separate lineages with no migration (Model 5 in Figure S3, receiving 72.6% of support), followed by a model with three lineages with isolation with ancient migration between the Northern and Southern lineages of *H. americana* (Model 13 in Figure S3, receiving 13.3% of support). The out-of-the-bag error rate for

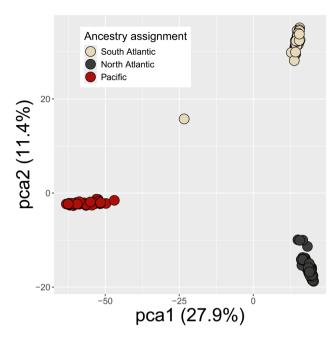


FIGURE 2 | Principal component analysis of 205 *Hetaerina titia* with a dataset of 3819 unlinked biallelic autosomal SNPs that were generated by mapping ddRAD reads to the draft genome of *H. titia*. Percentages indicate how much variation is explained by each component and colour indicates the highest assigned ancestry population from sNMF for each individual. The single point directly between the main Pacific and Atlantic cluster is the putative F_1 hybrid. A PCA plot for *H. americana* sensu lato showing broadly the same level of differentiation between samples, without any individuals showing introgression, is included in Figure S10.

H. americana and *H. calverti* demographic scenarios varied but was again low for models 5 and 13 (26% and 11%, respectively) and incorrect classifications were limited to the alternative of these two scenarios. No other demographic scenarios received any support.

3.5 | Divergence Times

SNAPP analysis using the SNPs mapped to the H. americana genome, the *H. titia* genome and mapped de novo, converged on the same tree and estimates of divergence times between each species and sub-population overlapped (Figure 4, Figures S18-S20). Based on the de novo SNP data, the divergence time between H. titia and H. americana sensu lato was estimated to be 24.5 mya (95% highest posterior density [HPD] 15.67–33.59 mya). The divergence between H. calverti and H. americana was estimated to be 6.83 mya (HPD 4.23-9.17 mya). SNAPP analysis also identified relatively distant dates for the divergence between the sub-populations within H. titia and H. americana. Populations of H. titia that reside in Atlantic drainages were estimated to have diverged from populations in the Pacific 3.74 mya (HPD 2.18-5.59 mya). The two lineages of H. titia that reside within Atlantic drainages separated at an estimated 1.11 mya (HPD 0.52-1.75 mya). The two identified lineages of H. americana diverged 3.25 mya (HPD 1.79-4.90 mya). Across the posterior distribution of trees of the de novo SNAPP run, the split between Pacific and Atlantic H. titia was younger than the split between

H.americana and *H. calverti* (mean 3.09 million years, HPD +1.48 to +4.86 million years). In the SNAPP analysis using the SNPs mapped to the draft genome of *H. americana*, in 99.8% of posterior distribution trees, the split between Pacific and Atlantic *H. titia* was younger than the split between *H. americana* and *H. calverti* (mean + 2.38 million years HPD +0.90 to +3.95 million years). Using the SNPs mapped to the draft genome of *H. titia*, in 75.3% of posterior distribution trees, the split between Pacific and Atlantic *H. titia* was younger than the split between Pacific and Atlantic *H. titia* was younger than the split between Pacific and Atlantic *H. titia* was younger than the split between *H. americana* and *H. calverti* (mean + 0.5 million HPD -1.17 to +1.92 million years).

Divergence times estimated by G-PhoCS were generally more recent than those estimated by SNAPP (Figure 5, Figures S21 and S22), but in all cases the divergence between *H. americana* and *H. calverti* was estimated as approximately twice as old as the split between Pacific and Atlantic clusters of *H. titia*.

The estimated effective population sizes for each lineage of *H. titia* were consistent between models and runs, with the exception being larger effective population sizes in runs that used loci mapped de novo (Figure S23). For *H. titia*, the Southern Atlantic *H. titia* lineage had the largest effective population size, around 1.0 million individuals (0.96–1.11 HPD), and the Northern Atlantic lineage had the smallest, around 0.28 million individuals (0.26–0.310 HPD). The Pacific lineage's effective population size was estimated to be around 0.39 million individuals (0.37–0.42 HPD). *Hetaerina calverti* was estimated to have a much greater effective population size than either lineage of *H. americana*: 1.1 million individuals (1.08–1.16 HPD) compared to 0.67 million (0.65–0.70 HPD) for the Southern *H. americana* lineage and 0.41 million (0.39–0.43 HPD) for the Northern *H. americana* lineage.

Where migration was included in the demographic models, the estimated migration rate between populations was low and consistent across all runs (Figure S23b). For all migration bands, the percentage of individuals within each population per generation that were estimated to have originated by migration was between 0.01 and 0.08 individuals per generation. For both *H. titia* and *H. americana*, migration from southern populations to northern populations was estimated to occur more often than migration from northern to southern populations (Figure 5). Posterior distributions of effective population size and migration rates across all runs are presented in Figure S23.

3.6 | An F₁ Hybrid at a Zone of Secondary Contact

Calculations of hybrid index and heterozygosity indicated that sample CUAJa02 from site CUAJ01 (an Atlantic drainage near the continental divide) is an F_1 hybrid between Pacific and Atlantic lineages (Figure 6). Sample CUAJa02 had an autosome heterozygosity of 93.3% and a hybrid index of 0.50, close to the theoretical level of an F_1 hybrid (100% and 0.5%, respectively) and markedly above the heterozygosity of an F_2 hybrid (50%). A second-generation backcross would produce a hybrid index of 0.25 or 0.75, depending on the proportion of Pacific versus Atlantic parentage. The X chromosome of sample CUAJa02 was nearly entirely homozygous for Pacific alleles. As *Hetaerina* exhibit an XO sex determination system, its parents were likely a

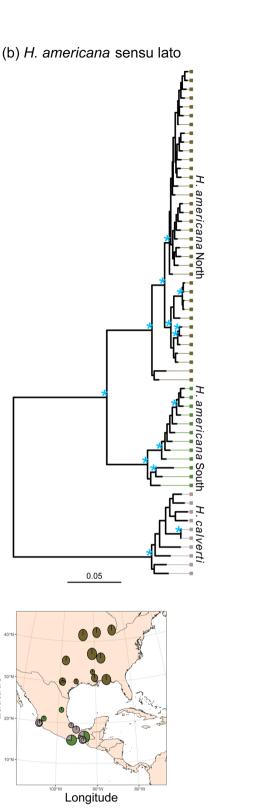


FIGURE 3 | The maximum likelihood tree for (a) *Hetaerina titia* and (b) *H. americana* sensu lato calculated using RAxML with 3020 SNPs for *H. titia* and 3949 SNPs for *H. americana* and *H. calverti* and mapped onto the genome of *H. titia*. Scale bar indicates the mean number of substitutions per SNP site. Due to exclusion of invariant sites and differences in the total number of SNPs used in each analysis, scale bars should not be used to compare phylogenetic distances within *H. titia* to distance within *H. americana* sensu lato. The nodes marked with a blue star '*' indicate a bootstrap support value (out of 100) of <95%. The tree tips are coloured according to the species and the max sNMF ancestry assignment (K=3). The geographical location of each sample is shown in the bottom two maps. Each pie chart shows the number of samples assigned to each ancestry cluster from each sample site, split between *H. titia* and *H. americana* sensu lato (*H. americana/calverti*).

-atitude

North Atlantic

South Atlantic

Pacific

0.05

Longitude

-atitude

(a) H. titia

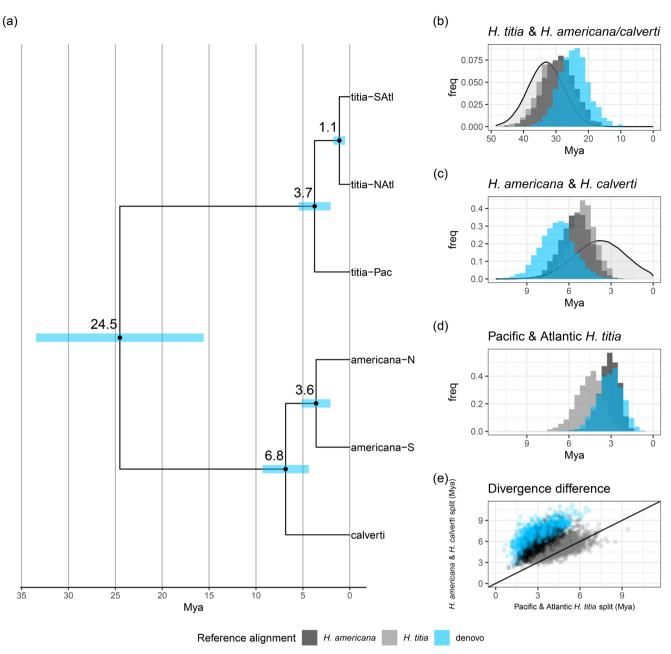


FIGURE 4 | (a) Estimates of divergence dates (million years ago—mya) between populations of Pacific *Hetaerina titia* (titia-Pac), Atlantic *H. titia* (titia-NAtl and titia-SAtl), *H. americana* (americana-N and americana-S) and *H. calverti* (calverti) calculated using SNAPP analysis in Beast. Node labels indicate the mean estimated divergence date with 95% highest posterior density in blue. All branches had a posterior distribution of 1. Tree plotted in R using the packages *treeio* and *ggtree*. Input data was 552 autosomal SNPs called using a de novo method of SNP calling (ipyrad). (b, c, d) The prior and posterior distribution (where applicable) of divergence times between the major lineages. The three different histograms denote the posterior distribution of the divergence times using three different SNP datasets, those mapped the draft genome of *H. americana* (dark grey), mapped the draft genome of *H. titia* (grey) and *de novo* SNP calling. The prior, where applicable, is denoted by the grey density distribution. (e) Comparison between the divergence times of *H. americana* and *H. calverti* and Atlantic and Pacific *H. titia*. Each point is the divergence times from a tree in the posterior distribution, the black line indicates values where the divergence times between the lineages are equal.

female from the Pacific lineage and a male from the Atlantic lineage. The single sex-linked heterozygous site in the hybrid individual had markedly higher read depth than the other SNPs on the X chromosome, suggesting the SNP was autosomal and incorrectly mapped to the X chromosome (Figure S24). The rate of heterozygosity across the highly divergent sites was close to zero for all other samples. All other samples either had nearly entirely Pacific or Atlantic genotypes.

4 | Discussion

We reconstructed the spatiotemporal dynamics of divergence in multiple lineages of rubyspot damselflies. As predicted by the non-adaptive radiation model commonly invoked for damselflies (Wellenreuther and Sánchez-Guillén 2016), we found evidence consistent with divergence times between lineages being positively related to levels of reproductive isolation and spatial

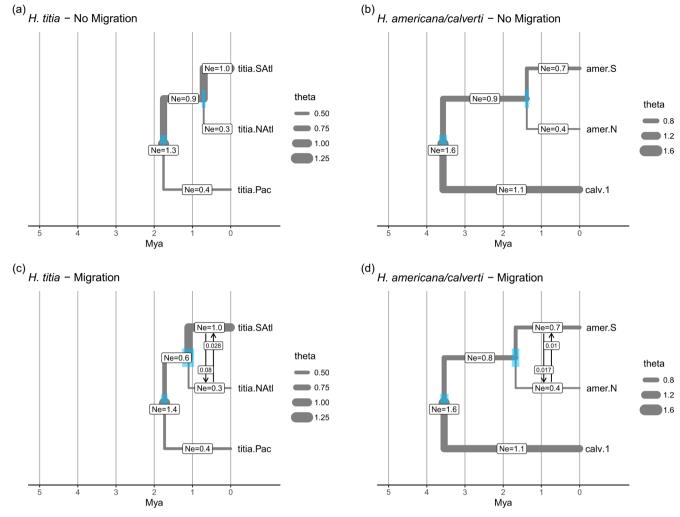


FIGURE 5 | The estimated divergence times (Mya = million years ago) and effective population size (theta—Ne in millions of individuals) from G-PhoCS analysis of *Hetaerina titia* and *H. americana*. Migration rate is the number of individuals per generation with vertical arrows indicating direction of migration (from and to). All models ran for 1,000,000 iterations with 10% burn in. Blue bars show 95% highest posterior density for each divergence date (a) Model estimates for *H. titia* with no migration bands. The estimated divergence time for Atlantic and Pacific *H. titia* was 2.72 mya (2.65–2.80 mya HPD) and divergence time between Northern and Southern Atlantic clusters was estimated as 0.59 mya (0.55–0.62 mya HPD). (b) Model estimates for *H. americana* and *H. calverti* with no migration bands. The divergence time for *H. calverti* and *H. americana* was estimated to be 4.7 mya (3.47–4.68 mya HPD). The Northern and Southern *H. americana* clusters divergence time between Northern and Southern Atlantic *H. titia* was 2.71 mya (2.64–2.79 HPD). (d) Model estimates for *H. titia* was 0.87 mya (0.76–1.01 mya HPD) and the split between North and Southern *H. americana*. The divergence time for *H. calverti* and *H. americana* was estimated for *H. americana* and *H. calverti* with migration bands between Northern and Southern *H. titia*. The divergence time between Northern and Southern Atlantic *H. titia* was 2.71 mya (2.64–2.79 HPD). (d) Model estimates for *H. americana* and *H. calverti* with migration bands between Northern and Southern *H. americana*. The divergence time for *H. calverti* and *H. americana* was estimated to be 4.68 mya (4.57–4.79 mya HPD). The Northern and Southern *H. americana* clusters diverged 1.75 mya (1.68–1.83 mya). G-PhoCS runs presented here are conducted on the RAD loci mapped de novo using ipyrad.

overlap. Specifically, the older species pair (*H. americana* and *H. calverti*, estimated to have diverged 6.8 mya in our SNAPP analysis) are those whose ranges broadly overlap and exhibit no evidence of introgression; the younger lineages (Pacific *H. titia* and Atlantic *H. titia*, estimated to have diverged 3.7 mya) are found largely in allopatry, with evidence of limited hybridisation suggesting strong post-zygotic isolation at a narrow point of secondary contact (See Figure 3 for the spatial distribution and geographical overlap of all *H. titia* and *H. americana* sensu lato lineages). In addition, our divergence time estimates reveal deep splits between sister lineages, in agreement with theory (Anderson et al. 2023; Czekanski-Moir and Rundell 2019; Rundell and Price 2009) and other studies that have estimated slow diversification rates in non-adaptive radiations—salamanders

(Kozak et al. 2005), killifish (Lambert et al. 2019), blindsnakes (Tiatragul et al. 2023, 2024) and snails (Fehér et al. 2013; Koch et al. 2020). We note, however, that non-adaptive radiations can also occur quickly when reproductive isolation is driven by rapid sexual selection coupled with strong geographic isolation (Blankers and Shaw 2024). Similarly, models of historical demography estimate extremely little to no migration between rubyspot lineages, consistent with theoretical requirements for non-ecological speciation (Nosil and Flaxman 2011). The deep divergence times between our identified lineages are further indicated by high $F_{\rm st}$ values, ranging from 0.521 to 0.833. Overall, these findings demonstrate that these rubyspot damselfly lineages are at different stages of the non-ecological speciation cycle. These observations also reinforce the converse notion

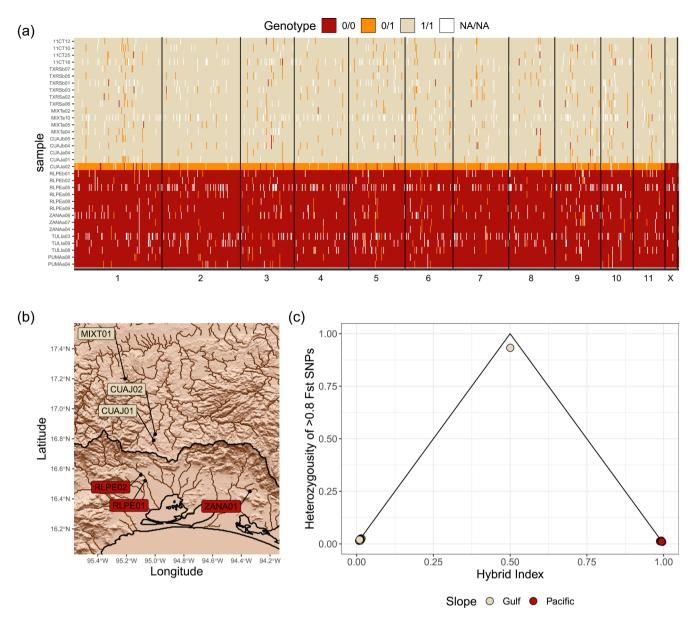


FIGURE 6 | Hybrid zone between Pacific and Southern Atlantic *Hetaerina titia* in the Isthmus of Tehuantepec. (a) Genotypes for 914 autosomal SNPs and 19 sex-linked SNPs that had a <0.8 allele frequency difference between Pacific and Atlantic individuals (calculations excluded samples from CUAJ01/02). Each sample is positioned along the *y*-axis with each SNP ordered by the position along each chromosome along the *x*-axis. The F_1 hybrid is sample CUAJa02. Each SNP is coloured by whether they were homozygous for the Pacific allele (0/0—red), homozygous for the Atlantic allele (1/1—beige) or heterozygous (0/1—orange). (b) Sample locations around the Isthmus of Tehuantepec. The Atlantic and Pacific watershed boundary is shown in black (see Figure 1b for a map of wider region and Figure S23 for a map of terrain height rather than a shaded relief). (c) A triangle plot showing the hybrid index, measuring the percentage of 'parental' genotype and the heterozygosity of each sample. A theoretical F_1 hybrid would be placed at the top corner of the triangle. SNPs on the X chromosome were excluded when calculating the hybrid index and heterozygosity.

that niche divergence accelerates speciation and highlight the usefulness of non-ecologically speciating taxa—with their lack of niche divergence and, in some cases, pre-zygotic isolation as simplified models for studying the accumulation of intrinsic reproductive isolation during the speciation process (Anderson et al. 2023). Further analyses in additional *Hetaerina* lineages would help to disentangle the mechanisms leading to reproductive isolation, such as mutation-order processes (Mendelson et al. 2014) and/or reproductive character displacement acting on traits that mediate isolation (e.g., genital morphology and mate recognition; Pfennig and Pfennig 2012). Moreover, geographical isolating barriers are not uniform in space and time, which is likely to influence the time it takes for daughter lineages to attain secondary contact and overlap in sympatry as well as the probability that speciation is nonephemeral.

Although our results support the non-ecological speciation model for damselfly diversification, a persistent challenge for determining whether speciation has occurred non-ecologically is the possibility that niche divergence has occurred along unmeasured axes, and therefore, that ecological speciation cannot be ruled out (Anderson and Weir 2022). Yet, this dichotomous framework is unrealistic, as speciation events are unlikely to proceed without any niche evolution, even if such divergence is minor and unlikely to have driven extensive reproductive isolation (Sobel et al. 2010; Anderson and Weir 2022). Indeed, in rubyspot damselflies, there is evidence for climate niche and microhabitat divergence between species (Grether et al. 2024; McEachin et al. 2022). Nevertheless, given the deep divergence time of *Hetaerina* (36 mya, Standring et al. 2022), we argue that such relatively minor divergence is unlikely to have driven pronounced reproductive isolation during speciation.

Our analyses shed light on a number of biogeographic factors that have influenced dynamics in Hetaerina. For instance, the three population clusters in H. titia identified in our analyses are separated by pronounced barriers to dispersal—the Continental Divide (separating the Pacific and Atlantic clusters) and the Trans-Mexican Volcanic Belt, separating the Northern and Southern Atlantic clusters. These have emerged as important phylogeographic barriers in other studies (Edwards et al. 2022; Mastretta-Yanes et al. 2015). Samples from San Luis Potosí, just north of the Trans-Mexican Volcanic Belt, had a majority Northern H. titia ancestry but with a potential small proportion of ancestry from Southern H. titia. Further sampling is required in the zone between Northern and Southern Atlantic lineages of *H. titia*, which occurs near a similar divide between Northern and Southern lineages of H. americana (Vega-Sánchez et al. 2024). The timing of the split between Pacific and Atlantic lineages of H. titia overlaps with the timing of the formation of the Isthmus of Panama; given its phylogenetic affinity with species found in South America (Standring et al. 2022), therefore, one hypothesis is that this split arose from northward dispersal. In other words, the last common ancestor of Pacific and Atlantic lineages of *H. titia* could have occurred in southern Central or northern South America before going locally extinct in that region. Our discovery of an F1 hybrid between Pacific and Southern Atlantic H. titia on the Isthmus of Tehuantepec demonstrates that the Pacific and Atlantic clusters have come into secondary contact in this region. The site with a hybrid individual is only ~27km from the nearest Pacific site where we have found *H. titia*. Here, the barrier to dispersal across the Continental Divide is reduced: the elevation of the Isthmus of Tehuantepec is around 200 m (having dropped from a higher elevation during the Late Miocene and Early Pliocene [Barrier et al. 1998]). In comparison, the mountains east and west of the region extend to over 2000 m in elevation with limited suitable riparian habitat. Finally, our demographic models estimated the lowest effective population sizes in the population clusters furthest north-a result consistent with demographic declines as a result of glaciation (Hewitt 2000).

Despite the presence of the hybrid individual, we did not detect any further admixture within these lineages that would suggest a history of introgression, suggesting that post-zygotic isolation may be complete, even if pre-zygotic isolation is not. In combination with the deep divergence time estimated for these lineages, it is likely that *H. titia* sensu lato represents a species complex containing multiple cryptic, reproductively isolated lineages. Further study is required to test whether there is divergence in mating preferences or reproductive traits (e.g., wing colour, genitalia morphology) between the two lineages and/ or low hybrid fitness related to reproductive traits. Male mate recognition in rubyspot damselflies is based largely on female wing colour (Drury, Anderson, et al. 2015; Drury, Okamoto, et al. 2015; Drury, Anderson, et al. 2019). Pacific and Atlantic H. titia exhibit marked differences in seasonal melanisation, which could allow discrimination between Pacific and Atlantic *H. titia*, but only during the peak-breeding season when newly emerged Atlantic H. titia exhibit high levels of wing melanisation (Drury, Anderson, et al. 2015; Drury, Barnes, et al. 2019). The F₁ individual male had wing pigmentation intermediate between Pacific and Atlantic lineages (with more red pigment than typical Atlantic individuals and more dark pigment than typical Pacific individuals, Figure S26). We have also obtained preliminary whole genome resequencing for an individual with a fully Pacific genotype, which was collected from the same Atlantic site and on the same date as the F₁ hybrid. Characterising mate recognition in Pacific and Atlantic H. titia within the site of secondary contact could further our understanding of the evolution of pre-zygotic mating barriers.

We find it unlikely that our sampling coincided with the first contact between Pacific and Atlantic H. titia in ~3.7 million years. What, therefore, has prevented Pacific H. titia from becoming more widely sympatric with Atlantic H. titia? Sympatry can be prevented by the production of low-fitness hybrids, which can cause population decline and local extinction (i.e., sexual exclusion [Irwin and Schluter 2022; Kuno 1992; Mikkelsen and Irwin 2021]). For instance, an increase in hybrid zones, due to climate-driven range shifts, has been identified as a conservation concern for an endangered species of damselfly (Sánchez-Guillén, Muñoz, et al. 2014). Within H. titia, there also may be differences in fitness between Pacific and Atlantic lineages. The high level of melanisation seen in the Atlantic lineages of H. titia is beneficial in reducing interspecific behavioural interference (Anderson and Grether 2011; Drury, Anderson, et al. 2015). Therefore, Atlantic H. titia may have an advantage over Pacific H. titia within river drainages that contain other species of Hetaerina, such as H. occisa and H. americana, which are found within the river drainage of the hybrid site (personal observation Patterson CW & Drury JP). Interspecific behavioural interference can itself influence the range dynamics of populations (Patterson and Drury 2023). Consequently, mating and territorial interactions between Pacific and Atlantic H. titia, as well as behavioural interference between Pacific H. titia and other Hetaerina spp. in Atlantic drainages, may be restricting the dispersal of the Pacific H. titia.

Our analyses and those of Vega-Sánchez et al. (2024) have uncovered unexpectedly deep splits between lineages of rubyspot damselflies in North and Central America. Such cryptic diversity appears to be common in damselflies (e.g., Polythore procera (Sánchez-Herrera and Realpe 2010), Megaloprepus caerulatus (Feindt et al. 2014), Matrona basilaris (Xue et al. 2019) Euphaea yayeyamana (Kanke et al. 2021), Ischnura senegalensis (Jiang et al. 2023), Rhinocypha fenestrella (Noorhidayah et al. 2024)), likely owing to recurring non-ecological speciation resulting from low dispersal and the presence of biogeographic barriers. Several studies of species distributed at higher latitudes, however, report relatively low levels of genetic differentiation between populations of Coenagrion spp. (Johansson et al. 2013), Ischnura elegans (Wellenreuther et al. 2011) and Calopteryx spp. (Kahilainen et al. 2014). This may be due to a relative lack of pronounced biogeographical barriers in northern Europe and/

or the effects of postglacial or contemporary northward range expansions (Dudaniec et al. 2018; Swaegers et al. 2013; Watts et al. 2010), which could lead to more ephemeral speciation in northern latitudes (Cutter and Gray 2016; Willink et al. 2024). Further genomic studies of additional large-range damselfly species will help to clarify the circumstances promoting repeated ecological speciation.

5 | Conclusion

We estimated divergence times for multiple lineages in a nonadaptive radiation. Divergence times correlate well with the stage of the non-ecological speciation cycle of each lineage pair, with the most distantly related lineages found in sympatry and the most closely related being in allopatry. We identified a site where there is contemporary but limited hybridisation between two highly differentiated lineages of the same (currently recognised) species. Collectively, this research provides insight into multiple stages of the non-ecological speciation cycle and paves the way for future work on diversification dynamics in nonadaptive radiations.

Author Contributions

C.P.: conceptualization, data curation, formal analysis, investigation, methodology, project administration, software, visualisation, writing – original draft, writing – review and editing. A.B.: methodology, supervision, writing – review and editing. H.C.: investigation, writing – review and editing. A.G.-R.: resources, writing – review and editing. G.F.G.: conceptualization, funding acquisition, resources, writing – review and editing. L.M.C.: resources, writing – review and editing. J.D.: conceptualization, funding acquisition, methodology, resources, supervision, project administration, writing – original draft, writing – review and editing.

Acknowledgements

This work was supported by a NERC Environmental Omics Facility Grant (NEOF1274) to J.D., funding from Durham University (including a Durham Doctoral Studentship to C.P.) and NSF DEB-NERC-2040883 to J.D. and G.F.G. Specimen collection was conducted under permit SGPA/DGVS/04421/21 issued by the Mexican Secretaría de Medio Ambiente y Recursos Naturales (SEMARNAT) to L.M.C.; permit 08112112 issued by the Florida Department of Environmental Protection to J.D.; licence FD/WL/7/21(04) issued by the Forest Department in Belize to J.D. & G.F.G.; permit No. R-SINAC-SE-DT-PI-003-2021 issued by the Costa Rican Ministry of Environment (Minae) and genetic access permission No. 377 issued by the Comisión Institucional de Biodiversidad, Universidad de Costa Rica. C.P. was provided training and support for laboratory work by Gavin Horsburgh at the Natural Environment Research Council Omics Facility (NEOF) at the University of Sheffield. NEOF also provided C.P. with training and support with bioinformatics. We thank Andreanna Welch, Lesley Lancaster, Erandi Bonillas-Monge and Dan Nesbit for helpful feedback on an earlier draft of the manuscript. We thank Erik Svensson, Thomas Blankers, and an anonymous reviewer for their additional helpful comments.

Disclosure

We forged a research collaboration between scientists from the countries where samples were obtained, and all collaborators are included as co-authors. Benefits from this research include the sharing of all sequence data in public databases, filing reports with relevant wildlife permitting agencies, and presentation of results in research seminars in Mexico and Costa Rica. Our groups are committed to international scientific partnerships and providing training opportunities for early career scientists from countries where the damselflies occur.

Conflicts of Interest

The authors declare no conflicts of interest.

Data Availability Statement

All code is available on GitHub: https://github.com/ChristophePatte rson/Phylogeography-Hetaerina and includes SNP libraries in vcf format. Raw demultiplexed sequence reads are available on NCBI under the accession number PRJNA1251623.

References

Anderson, C. N., and G. F. Grether. 2011. "Multiple Routes to Reduced Interspecific Territorial Fighting in *Hetaerina* Damselflies." *Behavioral Ecology* 22, no. 3: 527–534. https://doi.org/10.1093/beheco/arr013.

Anderson, S. A., H. López-Fernández, and J. T. Weir. 2023. "Ecology and the Origin of Nonephemeral Species." *American Naturalist* 201, no. 5: 619–638.

Anderson, S. A. S., and J. T. Weir. 2022. "The Role of Divergent Ecological Adaptation During Allopatric Speciation in Vertebrates." *Science* 378, no. 6625: 1214–1218. https://doi.org/10.1126/science.abo7719.

Arnegard, M. E., P. B. McIntyre, L. J. Harmon, et al. 2010. "Sexual Signal Evolution Outpaces Ecological Divergence During Electric Fish Species Radiation." *American Naturalist* 176, no. 3: 335–356. https://doi.org/10.1086/655221.

Barrier, E., L. Velasquillo, M. Chavez, and R. Gaulon. 1998. "Neotectonic Evolution of the Isthmus of Tehuantepec (Southeastern Mexico)." *Tectonophysics* 287, no. 1: 77–96. https://doi.org/10.1016/S0040-1951(98) 80062-0.

Barton, N. H., and G. M. Hewitt. 1989. "Adaptation, Speciation and Hybrid Zones." *Nature* 341, no. 6242: 497–503.

Blankers, T., and K. L. Shaw. 2024. "The Biogeographic and Evolutionary Processes Shaping Population Divergence in Laupala." *Molecular Ecology* 33, no. 15: e17444.

Bouckaert, R., T. G. Vaughan, J. Barido-Sottani, et al. 2019. "BEAST 2.5: An Advanced Software Platform for Bayesian Evolutionary Analysis." *PLoS Computational Biology* 15, no. 4: e1006650. https://doi.org/10. 1371/journal.pcbi.1006650.

Chifman, J., and L. Kubatko. 2014. "Quartet Inference From SNP Data Under the Coalescent Model." *Bioinformatics* 30, no. 23: 3317–3324. https://doi.org/10.1093/bioinformatics/btu530.

Corbet, P. S. 1999. "Dragonflies: Behavior and Ecology of Odonata." In *Dragonflies: Behavior and Ecology of Odonata*. Cornell Univ. Press. https://doi.org/10.1111/j.1365-2427.2001.00664.x.

Cutter, A. D., and J. C. Gray. 2016. "Ephemeral Ecological Speciation and the Latitudinal Biodiversity Gradient." *Evolution* 70, no. 10: 2171–2185.

Czekanski-Moir, J. E., and R. J. Rundell. 2019. "The Ecology of Nonecological Speciation and Nonadaptive Radiations." *Trends in Ecology & Evolution* 34, no. 5: 400–415. https://doi.org/10.1016/j.tree. 2019.01.012.

DaCosta, J. M., and M. D. Sorenson. 2014. "Amplification Biases and Consistent Recovery of Loci in a Double-Digest RAD-Seq Protocol." *PLoS One* 9, no. 9: e106713. https://doi.org/10.1371/journal.pone.0106713.

Danecek, P., J. K. Bonfield, J. Liddle, et al. 2021. "Twelve Years of SAMtools and BCFtools." *GigaScience* 10, no. 2: giab008. https://doi.org/10.1093/gigascience/giab008.

DeRaad, D. A., J. E. McCormack, N. Chen, A. T. Peterson, and R. G. Moyle. 2022. "Combining Species Delimitation, Species Trees, and Tests for Gene Flow Clarifies Complex Speciation in Scrub-Jays." *Systematic Biology* 71, no. 6: 1453–1470. https://doi.org/10.1093/sysbio/syac034.

Devitt, T. J., A. M. Wright, D. C. Cannatella, and D. M. Hillis. 2019. "Species Delimitation in Endangered Groundwater Salamanders: Implications for Aquifer Management and Biodiversity Conservation." *Proceedings of the National Academy of Sciences of the United States of America* 116: 2624–2633. https://doi.org/10.1073/pnas.1815014116.

Dion-Côté, A.-M., and D. A. Barbash. 2017. "Beyond Speciation Genes: An Overview of Genome Stability in Evolution and Speciation." *Current Opinion in Genetics & Development* 47: 17–23. https://doi.org/10.1016/j. gde.2017.07.014.

Drury, J. P., C. N. Anderson, M. B. C. Castillo, J. Fisher, S. McEachin, and G. F. Grether. 2019. "A General Explanation for the Persistence of Reproductive Interference." *American Naturalist* 194, no. 2: 268–275. https://doi.org/10.1086/704102.

Drury, J. P., C. N. Anderson, and G. F. Grether. 2015. "Seasonal Polyphenism in Wing Coloration Affects Species Recognition in Rubyspot Damselflies (*Hetaerina* spp.)." *Journal of Evolutionary Biology* 28, no. 8: 1439–1452. https://doi.org/10.1111/jeb.12665.

Drury, J. P., M. Barnes, A. E. Finneran, M. Harris, and G. F. Grether. 2019. "Continent-Scale Phenotype Mapping Using Citizen Scientists' Photographs." *Ecography* 42, no. 8: 1436–1445. https://doi.org/10.1111/ecog.04469.

Drury, J. P., and G. F. Grether. 2014. "Interspecific Aggression, Not Interspecific Mating, Drives Character Displacement in the Wing Coloration of Male Rubyspot Damselflies (*Hetaerina*)." *Proceedings of the Royal Society B: Biological Sciences* 281, no. 1796: 20141737. https://doi.org/10.1098/rspb.2014.1737.

Drury, J. P., K. W. Okamoto, C. N. Anderson, and G. F. Grether. 2015. "Reproductive Interference Explains Persistence of Aggression Between Species." *Proceedings of the Royal Society B: Biological Sciences* 282, no. 1804: 20142256. https://doi.org/10.1098/rspb.2014.2256.

Dudaniec, R. Y., C. J. Yong, L. T. Lancaster, E. I. Svensson, and B. Hansson. 2018. "Signatures of Local Adaptation Along Environmental Gradients in a Range-Expanding Damselfly (*Ischnura elegans*)." *Molecular Ecology* 27, no. 11: 2576–2593.

Eaton, D. A. R., and I. Overcast. 2020. "Ipyrad: Interactive Assembly and Analysis of RADseq Datasets." *Bioinformatics* 36, no. 8: 2592–2594. https://doi.org/10.1093/bioinformatics/btz966.

Edwards, S. V., V. V. Robin, N. Ferrand, and C. Moritz. 2022. "The Evolution of Comparative Phylogeography: Putting the Geography (And More) Into Comparative Population Genomics." *Genome Biology and Evolution* 14, no. 1: evab176. https://doi.org/10.1093/gbe/evab176.

Eme, D., M. Zagmajster, T. Delić, et al. 2018. "Do Cryptic Species Matter in Macroecology? Sequencing European Groundwater Crustaceans Yields Smaller Ranges but Does Not Challenge Biodiversity Determinants." *Ecography* 41, no. 2: 424–436. https://doi.org/10.1111/ ecog.02683.

Excoffier, L., I. Dupanloup, E. Huerta-Sánchez, V. C. Sousa, and M. Foll. 2013. "Robust Demographic Inference From Genomic and SNP Data." *PLoS Genetics* 9, no. 10: pgen.1003905. https://doi.org/10.1371/journal. pgen.1003905.

Excoffier, L., N. Marchi, D. A. Marques, R. Matthey-Doret, A. Gouy, and V. C. Sousa. 2021. "Fastsimcoal2: Demographic Inference Under Complex Evolutionary Scenarios." *Bioinformatics* 37, no. 24: 4882–4885. https://doi.org/10.1093/bioinformatics/btab468.

Fehér, Z., L. Németh, A. Nicoară, and M. Szekeres. 2013. "Molecular Phylogeny of the Land Snail Genus *Alopia* (Gastropoda: Clausiliidae) Reveals Multiple Inversions of Chirality." *Zoological Journal of the Linnean Society* 167, no. 2: 259–272. https://doi.org/10.1111/zoj.12002.

Feindt, W., O. Fincke, and H. Hadrys. 2014. "Still a One Species Genus? Strong Genetic Diversification in the World's Largest Living Odonate, the Neotropical Damselfly *Megaloprepus caerulatus.*" *Conservation Genetics* 15: 469–481.

Franchini, P., D. Monné Parera, A. F. Kautt, and A. Meyer. 2017. "quaddRAD: A New High-Multiplexing and PCR Duplicate Removal ddRAD Protocol Produces Novel Evolutionary Insights in a Nonradiating Cichlid Lineage." *Molecular Ecology* 26, no. 10: 2783–2795. https://doi. org/10.1111/mec.14077.

Frichot, E., and O. François. 2015. "LEA: An R Package for Landscape and Ecological Association Studies." *Methods in Ecology and Evolution* 6, no. 8: 925–929. https://doi.org/10.1111/2041-210X.12382.

Frichot, E., F. Mathieu, T. Trouillon, G. Bouchard, and O. François. 2014. "Fast and Efficient Estimation of Individual Ancestry Coefficients." *Genetics* 196, no. 4: 973–983. https://doi.org/10.1534/GENETICS.113. 160572.

Garrison, R. W. 1990. "A Synopsis of the Genus *Hetaerina* With Descriptions of Four New Species." *Transactions of the American Entomological Society* 116, no. 1: 175–259.

Gittenberger, E. 1991. "What About Non-Adaptive Radiation?" *Biological Journal of the Linnean Society* 43, no. 4: 263–272. https://doi.org/10.1111/j.1095-8312.1991.tb00598.x.

Gompert, Z., and A. C. Buerkle. 2010. "Introgress: A Software Package for Mapping Components of Isolation in Hybrids." *Molecular Ecology Resources* 10, no. 2: 378–384. https://doi.org/10.1111/j.1755-0998.2009. 02733.x.

Goudet, J. 2005. "Hierfstat, a Package for r to Compute and Test Hierarchical F-Statistics." *Molecular Ecology Notes* 5, no. 1: 184–186. https://doi.org/10.1111/j.1471-8286.2004.00828.x.

Grether, G. F., J. Beninde, E. Beraut, et al. 2023. "Reference Genome for the American Rubyspot Damselfly, *Hetaerina americana*." *Journal of Heredity* 114, no. 4: 385–394. https://doi.org/10.1093/jhered/esad031.

Grether, G. F., A. E. Finneran, and J. P. Drury. 2024. "Niche Differentiation, Reproductive Interference, and Range Expansion." *Ecology Letters* 27, no. 1: e14350. https://doi.org/10.1111/ele.14350.

Gronau, I., M. J. Hubisz, B. Gulko, C. G. Danko, and A. Siepel. 2011. "Bayesian Inference of Ancient Human Demography From Individual Genome Sequences." *Nature Genetics* 43, no. 10: 1031–1035. https://doi. org/10.1038/ng.937.

Gutenkunst, R. N., R. D. Hernandez, S. H. Williamson, and C. D. Bustamante. 2009. "Inferring the Joint Demographic History of Multiple Populations From Multidimensional SNP Frequency Data." *PLoS Genetics* 5, no. 10: e1000695. https://doi.org/10.1371/journal.pgen. 1000695.

Hewitt, G. 2000. "The Genetic Legacy of the Quaternary Ice Ages." *Nature* 405, no. 6789: 907–913. https://doi.org/10.1038/35016000.

Illera, J. C., J. C. Rando, M. Melo, L. Valente, and M. Stervander. 2024. "Avian Island Radiations Shed Light on the Dynamics of Adaptive and Nonadaptive Radiation." *Cold Spring Harbor Perspectives in Biology* 16, no. 12: a041451. https://doi.org/10.1101/cshperspect.a041451.

Irwin, D., and D. Schluter. 2022. "Hybridization and the Coexistence of Species." *American Naturalist* 200: E93–E109. https://doi.org/10.1086/720365.

Jiang, B., J. Zhang, X. Bai, et al. 2023. "Genetic Variation and Population Structure of a Widely Distributed Damselfly (*Ischnura senegalensis*)." *Archives of Insect Biochemistry and Physiology* 114, no. 2: 1–14.

Johansson, H., R. Stoks, V. Nilsson-Örtman, P. K. Ingvarsson, and F. Johansson. 2013. "Large-Scale Patterns in Genetic Variation, Gene Flow and Differentiation in Five Species of European Coenagrionid Damselfly Provide Mixed Support for the Central-Marginal Hypothesis." *Ecography* 36, no. 6: 744–755.

Jombart, T. 2008. "Adegenet: A R Package for the Multivariate Analysis of Genetic Markers." *Bioinformatics* 24, no. 11: 1403–1405. https://doi. org/10.1093/bioinformatics/btn129.

Kahilainen, A., I. Keränen, K. Kuitunen, J. S. Kotiaho, and K. E. Knott. 2014. "Interspecific Interactions Influence Contrasting Spatial Genetic Structures in Two Closely Related Damselfly Species." *Molecular Ecology* 23, no. 20: 4976–4988.

Kamvar, Z. N., J. F. Tabima, and N. J. Grünwald. 2014. "Poppr: An R Package for Genetic Analysis of Populations With Clonal, Partially Clonal, and/or Sexual Reproduction." *PeerJ* 2: e281. https://doi.org/10. 7717/peerj.281.

Kanke, E., K. Suzuki, K. Sekiné, et al. 2021. "Unexpected Population Genetic Structure in Two Closely Related Euphaeid Damselflies From the Yaeyama and Taiwan Islands (Odonata: Euphaeidae)." *Biological Journal of the Linnean Society* 134, no. 1: 214–228.

Keightley, P. D., R. W. Ness, D. L. Halligan, and P. R. Haddrill. 2014. "Estimation of the Spontaneous Mutation Rate per Nucleotide Site in a *Drosophila melanogaster* Full-Sib Family." *Genetics* 196, no. 1: 313–320. https://doi.org/10.1534/genetics.113.158758.

Knaus, B. J., and N. J. Grünwald. 2017. "Vcfr: A Package to Manipulate and Visualize Variant Call Format Data in R." *Molecular Ecology Resources* 17, no. 1: 44–53. https://doi.org/10.1111/1755-0998.12549.

Koch, E. L., M. T. Neiber, F. Walther, and B. Hausdorf. 2020. "Patterns and Processes in a Non-Adaptive Radiation: Alopia (Gastropoda, Clausiliidae) in the Bucegi Mountains." *Zoologica Scripta* 49, no. 3: 280–294. https://doi.org/10.1111/zsc.12406.

Kozak, K. H., D. W. Weisrock, and A. Larson. 2005. "Rapid Lineage Accumulation in a Non-Adaptive Radiation: Phylogenetic Analysis of Diversification Rates in Eastern North American Woodland Salamanders (Plethodontidae: Plethodon)." *Proceedings of the Royal Society B: Biological Sciences* 273, no. 1586: 539–546. https://doi.org/10. 1098/rspb.2005.3326.

Kuno, E. 1992. "Competitive Exclusion Through Reproductive Interference." *Researches on Population Ecology* 34, no. 2: 275–284. https://doi.org/10.1007/BF02514797.

Lambert, J. W., M. Reichard, and D. Pincheira-Donoso. 2019. "Live Fast, Diversify Non-Adaptively: Evolutionary Diversification of Exceptionally Short-Lived Annual Killifishes." *BMC Evolutionary Biology* 19, no. 1: 10. https://doi.org/10.1186/s12862-019-1344-0.

Li, H. 2011. "A Statistical Framework for SNP Calling, Mutation Discovery, Association Mapping and Population Genetical Parameter Estimation From Sequencing Data." *Bioinformatics* 27, no. 21: 2987–2993. https://doi.org/10.1093/bioinformatics/btr509.

Li, H., and R. Durbin. 2009. "Fast and Accurate Short Read Alignment With Burrows-Wheeler Transform." *Bioinformatics* 25, no. 14: 1754–1760. https://doi.org/10.1093/bioinformatics/btp324.

Lozier, J. D., J. M. Jackson, M. E. Dillon, and J. P. Strange. 2016. "Population Genomics of Divergence Among Extreme and Intermediate Color Forms in a Polymorphic Insect." *Ecology and Evolution* 6, no. 4: 1075–1091. https://doi.org/10.1002/ece3.1928.

Mastretta-Yanes, A., A. Moreno-Letelier, D. Piñero, T. H. Jorgensen, and B. C. Emerson. 2015. "Biodiversity in the Mexican Highlands and the Interaction of Geology, Geography and Climate Within the Trans-Mexican Volcanic Belt." *Journal of Biogeography* 42, no. 9: 1586–1600. https://doi.org/10.1111/jbi.12546.

Matute, D. R., and B. S. Cooper. 2021. "Comparative Studies on Speciation: 30 Years Since Coyne and Orr." *Evolution* 75, no. 4: 764–778. https://doi.org/10.1111/evo.14181.

McEachin, S., J. P. Drury, C. N. Anderson, and G. F. Grether. 2022. "Mechanisms of Reduced Interspecific Interference Between Territorial Species." *Behavioral Ecology* 33, no. 1: 126–136. https://doi.org/10.1093/ beheco/arab115. McPeek, M. A., and J. M. Brown. 2000. "Building a Regional Species Pool: Diversification of the Enallagma Damselflies in Eastern North America." *Ecology* 81, no. 4: 904–920. https://doi.org/10.1890/0012-9658(2000)081[0904:BARSPD]2.0.CO;2.

McPeek, M. A., L. Shen, and H. Farid. 2009. "The Correlated Evolution of Three-Dimensional Reproductive Structures Between Male and Female Damselflies." *Evolution* 63, no. 1: 73–83.

McPeek, M. A., L. B. Symes, D. M. Zong, and C. L. McPeek. 2011. "Species Recognition and Patterns of Population Variation in the Reproductive Structures of a Damselfly Genus." *Evolution* 65, no. 2: 419–428.

Mendelson, T. C., M. D. Martin, and S. M. Flaxman. 2014. "Mutation-Order Divergence by Sexual Selection: Diversification of Sexual Signals in Similar Environments as a First Step in Speciation." *Ecology Letters* 17, no. 9: 1053–1066. https://doi.org/10.1111/ele.12313.

Mendelson, T. C., and R. J. Safran. 2021. "Speciation by Sexual Selection: 20 Years of Progress." *Trends in Ecology & Evolution* 36, no. 12: 1153–1163. https://doi.org/10.1016/j.tree.2021.09.004.

Mikkelsen, E. K., and D. Irwin. 2021. "Ongoing Production of Low-Fitness Hybrids Limits Range Overlap Between Divergent Cryptic Species." *Molecular Ecology* 30, no. 16: 4090–4102. https://doi.org/10. 1111/mec.16015.

Noorhidayah, M., N. Azrizal-Wahid, V. L. Low, and N.-R. Yusoff. 2024. "Genetic Diversity and Phylogeographic Patterns of the Peacock Jewel-Damselfly, *Rhinocypha fenestrella* (Rambur, 1842)." *PLoS One* 19, no. 4: e0301392.

Nosil, P. 2012. Ecological Speciation. Oxford University Press.

Nosil, P., and S. M. Flaxman. 2011. "Conditions for Mutation-Order Speciation." *Proceedings of the Royal Society B: Biological Sciences* 278: 399–407. https://doi.org/10.1098/rspb.2010.1215.

Okamoto, K. W., and G. F. Grether. 2013. "The Evolution of Species Recognition in Competitive and Mating Contexts: The Relative Efficacy of Alternative Mechanisms of Character Displacement." *Ecology Letters* 16, no. 5: 670–678. https://doi.org/10.1111/ele.12100.

Paradis, E., and K. Schliep. 2019. "Ape 5.0: An Environment for Modern Phylogenetics and Evolutionary Analyses in R." *Bioinformatics* 35, no. 3: 526–528. https://doi.org/10.1093/bioinformatics/bty633.

Patterson, C. W., E. Bonillas-Monge, A. Brennan, et al. 2024. "A Chromosome-Level Genome Assembly for the Smoky Rubyspot Damselfly (*Hetaerina titia*)." *Journal of Heredity* 115, no. 1: esad070. https://doi.org/10.1093/jhered/esad070.

Patterson, C. W., and J. P. Drury. 2023. "Interspecific Behavioural Interference and Range Dynamics: Current Insights and Future Directions." *Biological Reviews* 98, no. 6: 2012–2027. https://doi.org/10. 1111/brv.12993.

Paulson, D. R. 1974. "Reproductive Isolation in Damselflies." *Systematic Biology* 23, no. 1: 40–49. https://doi.org/10.1093/sysbio/23.1.40.

Paulson, D. 2020. "Hetaerina titia in Panamá." Argia 32, no. 2: 850.

Peterson, B. K., J. N. Weber, E. H. Kay, H. S. Fisher, and H. E. Hoekstra. 2012. "Double Digest RADseq: An Inexpensive Method for De Novo SNP Discovery and Genotyping in Model and Non-Model Species." *PLoS One* 7, no. 5: 1–11. https://doi.org/10.1371/journal.pone.0037135.

Pfennig, D. W., and K. S. Pfennig. 2012. *Evolution's Wedge: Competition and the Origins of Diversity*. University of California Press. https://doi.org/10.1525/california/9780520274181.001.0001.

Ravinet, M., R. Faria, R. K. Butlin, et al. 2017. "Interpreting the Genomic Landscape of Speciation: A Road Map for Finding Barriers to Gene Flow." *Journal of Evolutionary Biology* 30, no. 8: 1450–1477. https://doi.org/10.1111/jeb.13047.

Reaz, R., M. S. Bayzid, and M. S. Rahman. 2014. "Accurate Phylogenetic Tree Reconstruction From Quartets: A Heuristic Approach." *PLoS One* 9, no. 8: e104008. https://doi.org/10.1371/journal.pone.0104008.

Rundell, R. J., and T. D. Price. 2009. "Adaptive Radiation, Nonadaptive Radiation, Ecological Speciation and Nonecological Speciation." *Trends in Ecology & Evolution* 24, no. 7: 394–399. https://doi.org/10.1016/j.tree. 2009.02.007.

Rundle, H. D., and P. Nosil. 2005. "Ecological Speciation." *Ecology Letters* 8, no. 3: 336–352. https://doi.org/10.1111/j.1461-0248.2004.00715.x.

Sánchez-Guillén, R. A., A. Córdoba-Aguilar, A. Cordero-Rivera, and M. Wellenreuther. 2014. "Genetic Divergence Predicts Reproductive Isolation in Damselflies." *Journal of Evolutionary Biology* 27, no. 1: 76–87. https://doi.org/10.1111/jeb.12274.

Sánchez-Guillén, R. A., J. Muñoz, J. Hafernik, M. Tierney, G. Rodriguez-Tapia, and A. Córdoba-Aguilar. 2014. "Hybridization Rate and Climate Change: Are Endangered Species at Risk?" *Journal of Insect Conservation* 18, no. 3: 295–305. https://doi.org/10.1007/s1084 1-014-9637-5.

Sánchez-Guillén, R. A., H. Van Gossum, and A. Cordero Rivera. 2005. "Hybridization and the Inheritance of Female Colour Polymorphism in Two Ischnurid Damselflies (Odonata: Coenagrionidae)." *Biological Journal of the Linnean Society* 85, no. 4: 471–481. https://doi.org/10. 1111/j.1095-8312.2005.00506.x.

Sánchez-Herrera, M., and E. Realpe. 2010. "Population Structure of *Polythore procera* at a Colombian Stream (Odonata: Polythoridae)." *International Journal of Odonatology* 13, no. 1: 27–37.

Schluter, D. 2000. The Ecology of Adaptive Radiation. OUP Oxford.

Simpson, G. G. 1944. *Tempo and Mode in Evolution*. Columbia University Press.

Smith, M. L., and B. C. Carstens. 2020. "Process-Based Species Delimitation Leads to Identification of More Biologically Relevant Species." *Evolution* 74, no. 2: 216–229. https://doi.org/10.1111/evo.13878.

Sobel, J. M., G. F. Chen, L. R. Watt, and D. W. Schemske. 2010. "The Biology of Speciation." *Evolution* 64, no. 2: 295–315. https://doi.org/10. 1111/j.1558-5646.2009.00877.x.

Stamatakis, A. 2014. "RAxML Version 8: A Tool for Phylogenetic Analysis and Post-Analysis of Large Phylogenies." *Bioinformatics* 30, no. 9: 1312–1313. https://doi.org/10.1093/bioinformatics/btu033.

Standring, S., M. Sánchez-Herrera, R. Guillermo-Ferreira, et al. 2022. "Evolution and Biogeographic History of Rubyspot Damselflies (Hetaerininae: Calopterygidae: Odonata)." *Diversity* 14, no. 9: 757. https://doi.org/10.3390/d14090757.

Struck, T. H., J. L. Feder, M. Bendiksby, et al. 2018. "Finding Evolutionary Processes Hidden in Cryptic Species." *Trends in Ecology & Evolution* 33, no. 3: 153–163. https://doi.org/10.1016/j.tree.2017.11.007.

Svensson, E. I., F. Eroukhmanoff, K. Karlsson, A. Runemark, and A. Brodin. 2010. "A Role for Learning in Population Divergence of Mate Preferences." *Evolution* 64, no. 11: 3101–3113. https://doi.org/10.1111/j. 1558-5646.2010.01085.x.

Svensson, E. I., M. A. Gómez-Llano, A. R. Torres, and H. M. Bensch. 2018. "Frequency Dependence and Ecological Drift Shape Coexistence of Species With Similar Niches." *American Naturalist* 191, no. 6: 691–703. https://doi.org/10.1086/697201.

Svensson, E. I., A. Runemark, M. N. Verzijden, and M. Wellenreuther. 2014. "Sex Differences in Developmental Plasticity and Canalization Shape Population Divergence in Mate Preferences." *Proceedings of the Royal Society B: Biological Sciences* 281, no. 1797: 20141636. https://doi.org/10.1098/rspb.2014.1636.

Swaegers, J., J. Mergeay, L. Therry, M. Larmuseau, D. Bonte, and R. Stoks. 2013. "Rapid Range Expansion Increases Genetic Differentiation While Causing Limited Reduction in Genetic Diversity in a Damselfly." *Heredity* 111, no. 5: 422–429.

Tiatragul, S., I. G. Brennan, E. S. Broady, and J. S. Keogh. 2023. "Australia's Hidden Radiation: Phylogenomics Analysis Reveals Rapid Miocene Radiation of Blindsnakes." *Molecular Phylogenetics and Evolution* 185: 107812. https://doi.org/10.1016/j.ympev.2023.107812.

Tiatragul, S., A. Skeels, and J. S. Keogh. 2024. "Morphological Evolution and Niche Conservatism Across a Continental Radiation of Australian Blindsnakes." *Evolution* 78, no. 11: 1854–1868. https://doi.org/10.1093/ evolut/qpae132.

Tobias, J. A., J. Ottenburghs, and A. L. Pigot. 2020. "Avian Diversity: Speciation, Macroevolution, and Ecological Function." *Annual Review of Ecology, Evolution, and Systematics* 51, no. 1: 533–560. https://doi.org/10.1146/annurev-ecolsys-110218-025023.

van der Valk, T., P. Pečnerová, D. Díez-del-Molino, et al. 2021. "Million-Year-Old DNA Sheds Light on the Genomic History of Mammoths." *Nature* 591, no. 7849: 265–269. https://doi.org/10.1038/s41586-021-03224-9.

Vega-Sánchez, Y. M., L. F. Mendoza-Cuenca, and A. González-Rodríguez. 2020. "*Hetaerina calverti* (Odonata: Zygoptera: Calopterygidae) sp. Nov., a New Cryptic Species of the American Rubyspot Complex." *Zootaxa* 4766, no. 3: 485–497. https://doi.org/10.11646/zootaxa.4766.3.7.

Vega-Sánchez, Y. M., K. Oyama, L. F. Mendoza-Cuenca, R. Gaytán-Legaria, and A. González-Rodríguez. 2024. "Genomic Differentiation and Niche Divergence in the *Hetaerina americana* (Odonata) Cryptic Species Complex." *Molecular Ecology* 33, no. 2: e17207. https://doi.org/ 10.1111/mec.17207.

Watts, P. C., S. Keat, and D. J. Thompson. 2010. "Patterns of Spatial Genetic Structure and Diversity at the Onset of a Rapid Range Expansion: Colonisation of the UK by the Small Red-Eyed Damselfly *Erythromma viridulum.*" *Biological Invasions* 12: 3887–3903.

Weir, J. T., and T. D. Price. 2011. "Limits to Speciation Inferred from Times to Secondary Sympatry and Ages of Hybridizing Species along a Latitudinal Gradient." *American Naturalist* 177, no. 4: 462–469. https://doi.org/10.1086/658910.

Wellenreuther, M., and R. A. Sánchez-Guillén. 2016. "Nonadaptive Radiation in Damselflies." *Evolutionary Applications* 9, no. 1: 103–118. https://doi.org/10.1111/eva.12269.

Wellenreuther, M., R. A. Sanchez-Guillen, A. Cordero-Rivera, E. I. Svensson, and B. Hansson. 2011. "Environmental and Climatic Determinants of Molecular Diversity and Genetic Population Structure in a Coenagrionid Damselfly." *PLoS One* 6, no. 6: e20440.

Westram, A. M., S. Stankowski, P. Surendranadh, and N. Barton. 2022. "What Is Reproductive Isolation?" *Journal of Evolutionary Biology* 35, no. 9: 1143–1164. https://doi.org/10.1111/jeb.14005.

Wilgenbusch, J. C., and D. Swofford. 2003. "Inferring Evolutionary Trees With PAUP*." *Current Protocols in Bioinformatics* 00, no. 1: 6.4.1–6.4.28. https://doi.org/10.1002/0471250953.bi0604s00.

Willink, B., J. L. Ware, and E. I. Svensson. 2024. "Tropical Origin, Global Diversification, and Dispersal in the Pond Damselflies (Coenagrionoidea) Revealed by a New Molecular Phylogeny." *Systematic Biology* 73, no. 2: 290–307. https://doi.org/10.1093/sysbio/syae004.

Xue, J., H. Zhang, X. Ning, W. Bu, and X. Yu. 2019. "Evolutionary History of a Beautiful Damselfly, *Matrona basilaris*, Revealed by Phylogeographic Analyses: The First Study of an Odonate Species in Mainland China." *Heredity* 122, no. 5: 570–581.

Zou, T., W. Kuang, T. Yin, et al. 2022. "Uncovering the Enigmatic Evolution of Bears in Greater Depth: The Hybrid Origin of the Asiatic Black Bear." *Proceedings of the National Academy of Sciences of the United States of America* 119, no. 31: e2120307119. https://doi.org/10. 1073/pnas.2120307119.

Supporting Information

Additional supporting information can be found online in the Supporting Information section.