



SOFTWARE TOOL ARTICLE

REVISED Facilitating accessible, rapid, and appropriate**processing of ancient metagenomic data with AMDiR**

[version 2; peer review: 1 approved, 3 approved with reservations]

Maxime Borry ^{1,2}, Adrian Forsythe³, Aida Andrades Valtueña²,
Alexander Hübner^{1,2,4}, Anan Ibrahim ⁵, Andrea Quagliariello⁶,
Anna E. White ^{7,8}, Arthur Kocher^{2,9}, Åshild J. Vågene¹⁰,
Bjørn Peare Bartholdy ¹¹, Diāna Spurīte^{2,12}, Gabriel Yaxal Ponce-Soto¹³,
Gunnar Neumann^{2,14}, I-Ting Huang¹⁵, Ian Light¹⁶, Irina M. Velsko²,
Iseult Jackson ^{17,18}, Jasmin Frangenberg ⁵, Javier G. Serrano¹⁹,
Julien Fumey ^{13,20}, Kadir T. Özdoğan^{21,22}, Kelly E. Blevins^{23,24}, Kevin G. Daly¹⁸,
Maria Lopopolo ¹³, Markella Moraitou²⁵, Megan Michel^{2,14,26},
Meriam van Os ²⁷, Miriam J. Bravo-Lopez^{28,29}, Mohamed S. Sarhan ^{30,31},
Nihan D. Dagtas³², Nikolay Oskolkov ^{33,34}, Olivia S. Smith³⁵,
Ophélie Lebrasseur^{36,37}, Piotr Rozwalak³⁸, Raphael Eisenhofer³⁹, Sally Wasef ⁴⁰,
Shreya L. Ramachandran ⁴¹, Valentina Vanghi³², Christina Warinner ^{2,4,42,43},
James A. Fellows Yates ^{2,4,5}

¹Cluster of Excellence "Balance of the Microverse", Leibniz Institute for Natural Product Research and Infection Biology Hans Knöll Institute, Adolf-Reichwein-Straße 23, Jena, Thuringia, 07745, Germany

²Department of Archaeogenetics, Max Planck Institute for Evolutionary Anthropology, Deutscher Pl. 6, Leipzig, Saxony, 04103, Germany

³Department of Animal Zoology, Uppsala Universitet, Norbyvägen 18D, Uppsala, 752 36, Sweden

⁴Associated Research Group of Archaeogenetics, Leibniz Institute for Natural Product Research and Infection Biology Hans Knöll Institute, Adolf-Reichwein-Straße 23, Jena, Thuringia, 07745, Germany

⁵Department of Paleobiotechnology, Leibniz Institute for Natural Product Research and Infection Biology Hans Knöll Institute, Adolf-Reichwein-Straße 23, Jena, Thuringia, 07745, Germany

⁶Department of Comparative Biomedicine and Food Science, Università degli Studi di Padova, Viale dell'Università 16, Legnaro, Padova, 350250, Italy

⁷Section for Molecular Ecology and Evolution, Globe Institute, Faculty of Health and Medical Sciences, Københavns Universitet, Øster Farimagsgade 5, Copenhagen K, 1353, Denmark

⁸BioArCh, Department of Archaeology, University of York, York, England, YO10 5DD, UK

⁹Transmission, Infection, Diversification and Evolution Group, Max Planck Institute for Geoanthropology, Kahlaische Str. 10, Jena, Thuringia, 07745, Germany

¹⁰Section for Hologenomics, Globe Institute, Faculty of Health and Medical Sciences, Københavns Universitet, Oester Voldgade 44747, Copenhagen K, 1350, Denmark

¹¹Department of Archaeological Sciences, Universiteit Leiden, Einsteinweg 2, Leiden, 2333 CC, The Netherlands

¹²Institute of Ecology and Evolution, Friedrich-Schiller-Universität Jena, Jena, Thuringia, 07743, Germany

¹³Microbial Paleogenomics Unit, Institut Pasteur, Université Paris Cité, CNRS UMR 2000, Rue du Docteur Roux 25-28, Paris, Île-de-France, F-75015, France

¹⁴Max Planck-Harvard Research Center for the Archaeoscience of the Ancient Mediterranean (MHAAM), Max Planck Institute for Evolutionary Anthropology, Deutscher Pl. 6, Leipzig, Saxony, 04103, Germany

¹⁵Department of Organismic and Evolutionary Biology, Harvard University, 26 Oxford St., Cambridge, Massachusetts, 02138, USA

¹⁶Max Planck Institute for Infection Biology, Virchowweg 12, Berlin, Berlin, 10117, Germany

¹⁷SFI Centre for Research Training in Genomics Data Science, University of Galway, Galway, H91 TK33, Ireland

¹⁸Smurfit Institute of Genetics, The University of Dublin Trinity College, Dublin, Leinster, D02 VF25, Ireland

¹⁹Department of Biochemistry, Microbiology, Cell Biology and Genetics, Universidad de La Laguna, San Cristóbal de La Laguna, Santa Cruz de Tenerife, 38200, Spain

²⁰Bioinformatics and Biostatistics Hub, Institut Pasteur, Université Paris Cité, Rue du Docteur Roux 25-28, Paris, Île-de-France, F-75015, France

²¹Animal Ecology, Wageningen Environmental Research, P.O box 47, Wageningen, Gelderland, 6700 AA, The Netherlands

²²Department of History and Art History, Universiteit Utrecht, Drift 6, Utrecht, Utrecht, 3512 BS, The Netherlands

²³Center for Bioarchaeological Research, Arizona State University, Candy Mall, Tempe, Arizona, 85281, USA

²⁴Department of Archaeology, Durham University, South Road, Durham, County Durham, England, DH1 3LE, UK

²⁵Institute of Ecology and Evolution, School of Biological Sciences, The University of Edinburgh, Charlotte Auerbach Road, Edinburgh, Scotland, EH9 3FL, UK

²⁶Department of Human Evolutionary Biology, Harvard University, Divinity Avenue 11, Cambridge, Massachusetts, 02138, USA

²⁷Department of Anatomy, University of Otago, 270 Great King St, Dunedin, Otago, 9016, New Zealand

²⁸International Laboratory for Human Genome Research (LIIGH), Universidad Nacional Autonoma de Mexico, La Mesa 3001, Juriquilla, Queretaro, 76230, Mexico

²⁹Center for Genomic Sciences (CCG), Universidad Nacional Autonoma de Mexico, Cuernavaca, Morelos, 62210, Mexico

³⁰Institute for Mummy Studies, Eurac Research, Drususallee 1, Bolzano/Bozen, Autonome Provinz Bozen, 39100, Italy

³¹Centre for Integrative Biology (CIBIO), Università degli Studi di Trento, Via Sommarive 9, Povo, Trentino, 38123, Italy

³²Department of Anatomy and Anthropology and Department of Human Molecular Genetics and Biochemistry, Faculty of Medicine, Tel Aviv University, Ramat Aviv, Tel Aviv-Yafo, 69978, Israel

³³National Bioinformatics Infrastructure Sweden, Science for Life Laboratory, Tomtebodavägen 23, Stockholm, 17165, Sweden

³⁴Department of Biology, Lunds Universitet, Sölvegatan 35, Lund, 223 62, Sweden

³⁵Department of Integrative Biology, The University of Texas at Austin, Speedway 2415, Austin, Texas, 78712, USA

³⁶Instituto Nacional de Antropología y Pensamiento Latinoamericano, 3 de Febrero 1370 (1426), Ciudad Autónoma de Buenos Aires, C1426BJN CABA, Argentina

³⁷Center for Anthropobiology and Genomics of Toulouse, CNRS/Université Toulouse III Paul Sabatier, Allées Jules Guesde 37, Toulouse, Occitanie, 31000, France

³⁸Department of Computational Biology, Adam Mickiewicz University, Poznań, Uniwersytetu Poznańskiego 6, Poznań, Wielkopolska, 61-614, Poland

³⁹Center for Evolutionary Hologenomics, Globe Institute, Københavns Universitet, ester Voldgade 44747, Copenhagen, Copenhagen K, 1350, Denmark

⁴⁰Defence Genomics, Centre for Genomics and Personalised Health, Queensland University of Technology, Musk Ave 60, Kelvin Grove, Queensland, 4059, Australia

⁴¹Department of Human Genetics, The University of Chicago, E. 58th St. 920, Chicago, Illinois, 60637, USA

⁴²Department of Anthropology, Harvard University, Divinity Avenue 11, Cambridge, Massachusetts, 02138, USA

⁴³Faculty of Biological Sciences, Institute of Microbiology, Friedrich-Schiller-Universität Jena, Neugasse 25, Jena, Thuringia, 07743, Germany

v2 First published: 02 Aug 2023, 12:926
<https://doi.org/10.12688/f1000research.134798.1>

Latest published: 28 May 2024, 12:926
<https://doi.org/10.12688/f1000research.134798.2>





Abstract

Background

Access to sample-level metadata is important when selecting public metagenomic sequencing datasets for reuse in new biological analyses. The Standards, Precautions, and Advances in Ancient Metagenomics community (SPAAM, <https://spaam-community.org>)

Open Peer Review

Approval Status 

	1	2	3	4
version 2				
(revision)				
28 May 2024			view	view
version 1				
02 Aug 2023	view	view		

has previously published AncientMetagenomeDir, a collection of curated and standardised sample metadata tables for metagenomic and microbial genome datasets generated from ancient samples. However, while sample-level information is useful for identifying relevant samples for inclusion in new projects, Next Generation Sequencing (NGS) library construction and sequencing metadata are also essential for appropriately reprocessing ancient metagenomic data. Currently, recovering information for downloading and preparing such data is difficult when laboratory and bioinformatic metadata is heterogeneously recorded in prose-based publications.

Methods

Through a series of community-based hackathon events, AncientMetagenomeDir was updated to provide standardised library-level metadata of existing and new ancient metagenomic samples. In tandem, the companion tool 'AMDirT' was developed to facilitate rapid data filtering and downloading of ancient metagenomic data, as well as improving automated metadata curation and validation for AncientMetagenomeDir.

Results

AncientMetagenomeDir was extended to include standardised metadata of over 6000 ancient metagenomic libraries. The companion tool 'AMDirT' provides both graphical- and command-line interface based access to such metadata for users from a wide range of computational backgrounds. We also report on errors with metadata reporting that appear to commonly occur during data upload and provide suggestions on how to improve the quality of data sharing by the community.

Conclusions

Together, both standardised metadata reporting and tooling will help towards easier incorporation and reuse of public ancient metagenomic datasets into future analyses.

Keywords


metagenomics, environmental, palaeogenomics, aDNA, microbiome, metadata, microbial, FAIR data



This article is included in the [Bioinformatics gateway](#).

1. **Timothy Read**, Emory University, Atlanta, USA

Robert Petit III, Wyoming Public Health Laboratory, Cheyenne, USA

2. **Jonas Coelho Kasmanas** , University of São Paulo, São Paulo, Brazil

3. **Intikhab Alam**, King Abdullah University of Science and Technology, Thuwal, Saudi Arabia

4. **Emma Griffiths**, Simon Fraser University, Burnaby, USA

Any reports and responses or comments on the article can be found at the end of the article.



This article is included in the **Max Planck Society** collection.



This article is included in the **Evolutionary Genomics** collection.

Corresponding authors: Maxime Borry (maxime_borry@eva.mpg.de), Christina Warinner (christina_warinner@eva.mpg.de), James A. Fellows Yates (james_fellows_yates@eva.mpg.de)

Author roles: **Borry M:** Conceptualization, Methodology, Software, Supervision, Validation, Writing – Original Draft Preparation, Writing – Review & Editing; **Forsythe A:** Data Curation, Writing – Review & Editing; **Andrades Valtueña A:** Data Curation, Writing – Review & Editing; **Hübner A:** Data Curation, Software, Writing – Review & Editing; **Ibrahim A:** Data Curation, Writing – Review & Editing; **Quagliariello A:** Data Curation, Writing – Review & Editing; **White AE:** Data Curation, Writing – Review & Editing; **Kocher A:** Data Curation, Writing – Review & Editing; **Vågene AJ:** Data Curation, Writing – Review & Editing; **Bartholdy BP:** Data Curation, Writing – Review & Editing; **Spurite D:** Data Curation, Writing – Review & Editing; **Ponce-Soto GY:** Data Curation, Writing – Review & Editing; **Neumann G:** Data Curation, Writing – Review & Editing; **Huang IT:** Data Curation, Writing – Review & Editing; **Light I:** Data Curation, Software, Writing – Review & Editing; **Velsko IM:** Data Curation, Writing – Review & Editing; **Jackson I:** Data Curation, Writing – Review & Editing; **Frangenberg J:** Data Curation, Software, Writing – Review & Editing; **Serrano JG:** Data Curation, Writing – Review & Editing; **Fumey J:** Software, Writing – Review & Editing; **Özdoğan KT:** Data Curation, Writing – Review & Editing; **Blevins KE:** Data Curation, Writing – Review & Editing; **Daly KG:** Data Curation, Writing – Review & Editing; **Lopopolo M:** Data Curation, Writing – Review & Editing; **Moraitou M:** Data Curation, Writing – Review & Editing; **Michel M:** Data Curation, Writing – Review & Editing; **van Os M:** Data Curation, Writing – Review & Editing; **Bravo-Lopez MJ:** Data Curation, Writing – Review & Editing; **Sarhan MS:** Data Curation, Writing – Review & Editing; **Dagtas ND:** Data Curation, Writing – Review & Editing; **Oskolkov N:** Software, Writing – Review & Editing; **Smith OS:** Data Curation, Writing – Review & Editing; **Lebrasseur O:** Data Curation, Writing – Review & Editing; **Rozwalak P:** Software, Writing – Review & Editing; **Eisenhofer R:** Data Curation, Writing – Review & Editing; **Wasef S:** Data Curation, Writing – Review & Editing; **Ramachandran SL:** Data Curation, Writing – Review & Editing; **Vanghi V:** Data Curation, Writing – Review & Editing; **Warinner C:** Conceptualization, Funding Acquisition, Supervision, Writing – Original Draft Preparation, Writing – Review & Editing; **Fellows Yates JA:** Conceptualization, Data Curation, Investigation, Methodology, Project Administration, Software, Supervision, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: M.B., A.A.V., A.H., A.K., D.S., G.N., I.L., I.M.V., M.Mi., C.W., and J.A.F.Y. were supported by Max Planck Society. A.F. was supported by Swedish Research Council (Formas), Science for Life Laboratory National Sequencing Projects (SciLife) and the Carl Tryggers Stiftelse. A.H. was supported by the European Research Council under the European Union's Horizon 2020 research and innovation programme (grant agreement number 804884-DAIRYCULTURES awarded to C.W.). M.B. and A.H. were supported under Germany's Excellence Strategy EXC 2051 (Project-ID 390713860, "Balance of the Microverse"). A.I., J.F., M.B., I.M.V., C.W., and J.A.F.Y. were supported by Werner Siemens Foundation grant 'Palaeobiotechnology' (awarded to Prof. Pierre Stallforth and C.W.). A.Q. was supported by the S.T.A.R.S 2019 program from the University of Padua. A.E.W. was supported by the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No. 956351. Å.J.V. was supported by Carlsbergfondet Semper Ardens grant CF18-1109 (to Prof. M. Thomas P. Gilbert). D.S. was supported by Honours-Programm für forschungsorientierte Studierende (Friedrich-Schiller-Universität Jena). G.N. was supported by the European Research Council under the European Union's Horizon 2020 research and innovation program (grant agreement number 771234-PALEORIDER). G.N., M.M., and C.W. were supported by the Max Planck-Harvard Research Center for the Archaeoscience of the Ancient Mediterranean (MHAAM). G.Y.P.S. and M.L. was supported by the European Research Council under the European Union's Horizon 2020 research and innovation program Starting Grant agreement number 948800 PaleoMetAmerica (to Dr. Nicolás Rascovan). I.J. was supported by the Science Foundation Ireland Centre for Research Training in Genomics Data Science (Grant 18/CRT/6214). J.G.S. was supported by European Research Council under the European Union's Horizon 2020 research and innovation program Starting Grant agreement number 851733 IsoCAN (to Dr. Rosa Irene Fregel Lorenzo). K.T.Ö. was supported by 'Constructing the Limes: Employing citizen science to understand borders and border systems from the Roman period until today' (C-Limes), funded by the Dutch Research Council (NWO) as part of the Dutch Research Agenda (NWA, 2021-2026, project number: NWA.1292.19.364). K.E.B. was supported by Leverhulme Trust Research Grant 'What's in a house? Exploring the kinship structure of the world's first houses' (Project Ref. 84009). K.G.D. was supported by a Science Foundation Ireland – Irish Research Council (SFI-IRC) Pathways grant (Grant 21/PATH-S/9515). M.L. was supported the Institut Pasteur's INCEPTION-program (Investissement d'Avenir grant ANR-16-CONV-0005). N.O. was supported by the Knut and Alice Wallenberg Foundation. O.S.S. was supported by the NSF Graduate Research Fellowship Program (Grant no. DGE 2137420). O.L. was supported by the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement no. 895107. R.E. was supported by the Carlsberg Fellowship for Associate Professors CF20-0460 (awarded to Assoc. Prof. Antton Alberdi). S.L.R. was supported by the NIH Genetic Mechanisms of Evolution Training Grant (Grant No. T32 GM139782).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2024 Borry M *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Borry M, Forsythe A, Andrades Valtueña A *et al.* **Facilitating accessible, rapid, and appropriate processing of ancient metagenomic data with AMDiR [version 2; peer review: 1 approved, 3 approved with reservations]** F1000Research 2024, 12:926 <https://doi.org/10.12688/f1000research.134798.2>

First published: 02 Aug 2023, 12:926 <https://doi.org/10.12688/f1000research.134798.1>

REVISED Amendments from Version 1

In the revised version of this manuscript we describe a new command line subcommand 'download', as well as an updated viewer GUI and convert CLI that allows for library metadata filtering. At the request of a reviewer, we've added support for SRAToolkit as additional sequencing data download option. We have also improved installation and hardware requirement documentation, and fixed a few bugs also identified by reviewers.

Any further responses from the reviewers can be found at the end of the article

Introduction

The field of palaeogenomics has been praised as a role model for scientific data reporting and data availability.¹ When compared against FAIR principles (Findability, Accessibility, Interoperability, and Reusability),² ancient DNA (aDNA) sequencing data have been consistently made available in standard data formats on public data repositories, satisfying the principles of *accessibility*, *interoperability* and, to a certain extent, *reusability*. However, the *findability* of the uploaded data still poses challenges, often due to the lack of inclusion of key metadata specific for aDNA in the standardised sample metadata fields used by public sequencing repositories such as the European Bioinformatics Institute's European Nucleotide Archive (EBI ENA), the US National Center for Biotechnology Information's Sequence Read Archive (NCBI SRA), and the Japanese National Institute of Genetics' DNA Data Bank of Japan (NIG DDBJ). To improve findability of ancient metagenomic samples in public data repositories, the SPAAM community (<https://spaam-community.org>) previously developed the AncientMetagenomeDir project, a set of curated standard sample metadata for ancient host-associated shotgun-sequenced metagenomes, ancient environmental metagenomes, and/or host-associated microbial genomes.³ However, while sample-level metadata already help with the discovery of suitable comparative data, library-level metadata are also needed to further facilitate data reuse in dedicated aDNA analysis pipelines such as PALEOMIX,⁴ nf-core/eager,⁵ aMeta,⁶ and nf-core/mag.⁷ aDNA researchers often build many different types of NGS libraries⁸ and may generate (meta)genomic data using multiple different sequencing platforms that require different bioinformatic pre-processing workflows. Furthermore, the library-level metadata currently available in public repositories often lack key information about aDNA library treatments and other laboratory information needed to reproducibly reanalyse palaeogenomic datasets obtained from different studies.

An ancient metagenome can be generally described as the entire genetic content of a sample, within which at least a portion of the DNA has degraded over time.³ As the number of ancient metagenomics samples and shotgun sequenced library files steadily increases (currently >2500 host-associated metagenome, >3000 single-genome, and >700 environmental metagenome sequencing run accessions as of April 2024; **Figure 1**), the need to efficiently identify, curate, and

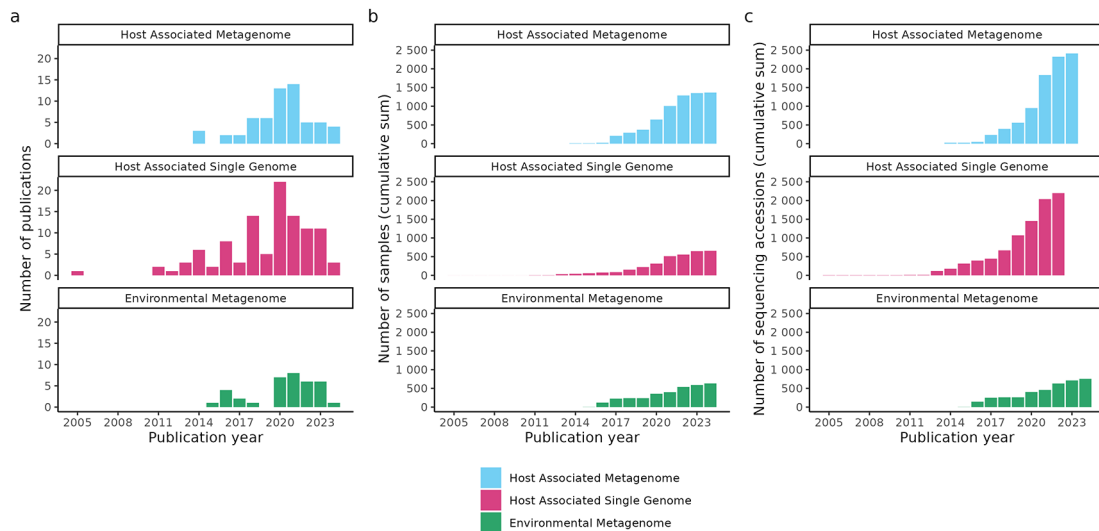


Figure 1. Growth of studies curated in the AncientMetagenomeDir as of v24.03. (a) Number of ancient metagenomic publications published per year with open sequencing data and included in AncientMetagenomeDir. The original AncientMetagenomeDir publication was in 2020. (b) Cumulative sum of the number of published samples with publicly accessible sequencing data. (c) Cumulative sum of the number of ancient metagenomic sequencing data accessions of the samples in panel b. Data from Fellows Yates et al.¹²

download such data is becoming more pressing. Although the original AncientMetagenomeDir releases provided project- or sample-level accession numbers that point to data primarily hosted by the ENA, SRA, and DDBJ, the metadata tables did not provide direct links to the data themselves. This meant that researchers still needed to manually search for each project or sample accession number in public data repositories and then manually identify and download the relevant associated files. Researchers were then required to parse and evaluate each sequencing file for inclusion in their study by consulting the original scientific publications for laboratory, library, and sequencing metadata. As with sample metadata, the reporting of this information within publications can be heterogeneous, may appear in the main text or supplement, and may take the form of prose text, tables, supplementary spreadsheets, or citations to other publications or protocols. While other tools for exploring public data repositories exist, such as NCBImeta,⁹ SRA-Explorer,¹⁰ and ffq,¹¹ they are generally limited to a restricted set of metadata available for inspection or require the use of command-line filtering tools, an interface not always accessible to all palaeogenomics researchers, who often have varying levels of computational experience.

Here we present AMDiT (AncientMetagenomeDir Toolkit), a tool designed to assist researchers in using a new extension of AncientMetagenomeDir that now includes aDNA library- and sequencing-level metadata. AMDiT is designed to provide a solution to four different challenges, thanks to new command line interfaces (CLI), a new graphical user interface (GUI), and a hosted web version (<https://spaam-community.org/AMDirT>). First, new CLI tooling helps contributors to AncientMetagenomeDir to curate newly published aDNA sequencing data in AncientMetagenomeDir by automatically retrieving relevant library-level metadata available from sequencing archives (`autofill` CLI command) and preparing semi-filled data entry tables for submission. Second, AMDiT also helps project reviewers automate a variety of data validation tasks on completed entry tables to ensure consistency (`validate` CLI command, an improved version of the AncientMetagenomeDirCheck tool from Ref. 3). Third, AMDiT now provides users with a web browser-based GUI that allows researchers to explore relevant ancient metagenomics-related sequencing datasets in AncientMetagenomeDir tables (`viewer` command) and CLI interfaces (`download` and `convert` CLI commands) to download metadata and export data download scripts from International Nucleotide Sequence Database Collaboration (INSDC) repositories. Finally, as an additional functionality, both AMDiT viewer GUI and CLI interfaces can generate template input configuration files for a suite of standard aDNA metagenomics-related pipelines in order to further automate and accelerate the processing of such aDNA data. AMDiT is available for installation via PyPI (<https://pypi.org/>) or Bioconda,¹³ with source code on GitHub under <https://github.com/SPAAM-community/AMDirT>.

Methods

Implementation

AMDirT tool implementation. Members of the SPAAM community an international and open community of nearly 500 researchers work on ancient metagenomics (<https://spaam-community.org>), developed AMDiT through a series of code sprints and hackathons, using Python (v3.9, <https://www.python.org/>; RRID:SCR_008394). It is accessible via a command-line-interface written using Click or via a python API (<https://click.palletsprojects.com/>). The `autofill` command uses the ENA portal API (<https://www.ebi.ac.uk/ena/portal/api/>)¹⁴ to automatically query and return metadata associated with the sequencing library level, such as all project, samples and library accessions, location and size of FASTQ files, sequencing instrument model, library strategy and layout, as well as read count. Data validation in the `validate` subcommand is performed using the jsonschema python library (<https://python-jsonschema.readthedocs.io/>) by validating the dataframes containing the sample level and library level metadata against their respective JSON schema, and using a variety of checks written using Pandas¹⁵ to avoid data duplication and ensure consistency of new entries. Additionally, `validate` will also check that each publication has its own valid DOI, that each sequence archive accession is valid, unique, and associated with the correct project accession. Any errors will be reported in table format, indicating the type of error, the line and column location of the error, and a short explanation of the error, and how to fix it. Both tools are primarily used within automated GitHub actions processes on the AncientMetagenomeDir GitHub repository, however are also usable by submitters and curators running on their own machines.

The GUI data exploration interface of the `viewer` command was developed using Streamlit (<https://streamlit.io/>), and the streamlit-aggrid library¹⁶ is used to allow the end-user to interactively filter and prepare configuration files to process ancient (meta)genomic data in bioinformatics pipelines. AMDiT is packaged thanks to setuptools,¹⁷ and is distributed on PyPi and Bioconda.¹³ The source code is available on GitHub (github.com/SPAAM-community/AMDirT), and associated documentation is provided online (amdirt.readthedocs.io). Furthermore, an online serverless version of the AMDiT viewer tool is available at <https://spaam-community.org/AMDirT> thanks to the stlite library (<https://github.com/whitphx/stlite>), a port of streamlit to WebAssembly that is supported (at the time of writing) by most Chrome-based browsers.

The CLI based `convert` command reuses the backend of the `viewer` command to provide a terminal based filtering functionality for more advanced users. Finally, the `download` command provides a CLI interface to download the

different AncientMetagenomeDir tables using the standard python library, with the possibility of specifying the release and the table type.

AncientMetagenomeDir library metadata aggregation. To extend the original AncientMetagenomeDir³ repository to include library metadata, we created new tab-separated value (TSV) tables and their associated validation checks in the form of JSON schema files, following the original AncientMetagenomeDir structure.

We retained the TSV format for maximum software compatibility, as originally described in Ref. 3. Fields included in the new library-level schema were selected after consultation with ancient metagenomics researchers of the SPAAM community, and, where relevant and possible, by mirroring existing metadata fields and controlled vocabulary from the ENA repository. Newly added library information columns include the library name (how data are typically reported in original publications), the aDNA library generation method (e.g., double-stranded or single-stranded libraries), the library indexing polymerase (e.g., proof-reading or non-proofreading), and the library pretreatment method (e.g., non-Uracil-DNA Glycosylase (UDG), full-UDG, or half-UDG treatments). The latter three fields represent information about the sequencing library construction that influences the presence of aDNA damage, a factor that is critical for the processing of aDNA NGS data.^{8,18} Sequencing metadata columns include instrument model, library layout (single- or paired-end), library strategy (whole genome sequencing, targeted capture, etc.), and read count. These metadata are also critical for correct processing of aDNA data. For example, whether an instrument uses 2- or 4-colour sequencing chemistry determines if poly-G tail trimming is required to remove sequence artefacts that arise in aDNA reads that are normally shorter than the number of sequencing cycles. Library layout is also necessary to indicate whether read-pair merging needs to be applied prior to mapping, or whether unmerged read pairs are available for *de novo* assembly. The remaining columns provide information about storage and file retrieval of sequencing data: direct URLs to FASTQ files, 'md5 checksum' strings (for post-download integrity verification), and download sizes (for storage space usage estimation). Tables may also be extended to contain field-specific metadata useful for data processing under specific conditions. For new non-ENA/SRA supported fields, such as library polymerase or library treatment, we defined fixed lists via new JSON-based 'enum' files stored in the AncientMetagenomeDir repository, as with the sample-level metadata.

Via a series of community events, we then manually carried out data entry and curation for the new columns of metadata by comparing the ENA stored metadata with the methods descriptions in original publications. This procedure identified multiple instances of inconsistencies between the two sources, as well as incorrectly uploaded data and metadata in previously published articles. We describe some of the common issues we encountered in the Discussion section below. In cases of conflict between the publication and the ENA metadata, we attempted to contact the original authors of the publication for confirmation. When this was not possible, we used 'unknown' or another missing-data value to indicate uncertainty. Each library-level metadata addition underwent automated validation and peer-review following the same procedure described in Ref. 3. Since the community events, the AMDirT `autofill` command has been developed to improve the library metadata submission experience by community members contributing new metadata (Figure 3). The `autofill` sub-command automates the pulling of ENA metadata into a 'draft' library-table format during the continuous integration tests (CI) of a GitHub pull request of sample-level metadata, replacing and improving upon the manually executed R scripts used in the initial pull-down for the community events. Submitters to AncientMetagenomeDir can then copy over much of the metadata and fill in the remaining missing metadata not covered by the existing ENA metadata fields.

Operation

AMDirT requires a UNIX-based terminal (e.g., Linux, OSX, Windows Subshell for Linux) for both installation and initial usage; however, the toolkit is written in Python and can therefore be used on a wide range of platforms and operating systems.

To install, users are recommended to use the pip or conda package managers. Users who wish to use the GUI based table viewer and downloader will also require any modern web browser supported by Streamlit (<https://streamlit.io/>).

For example, to install and load the help message:

```
$ pip install amdirt
```

or via conda in a dedicated environment

```
$ conda create -n amdirt -c bioconda amdirt
```

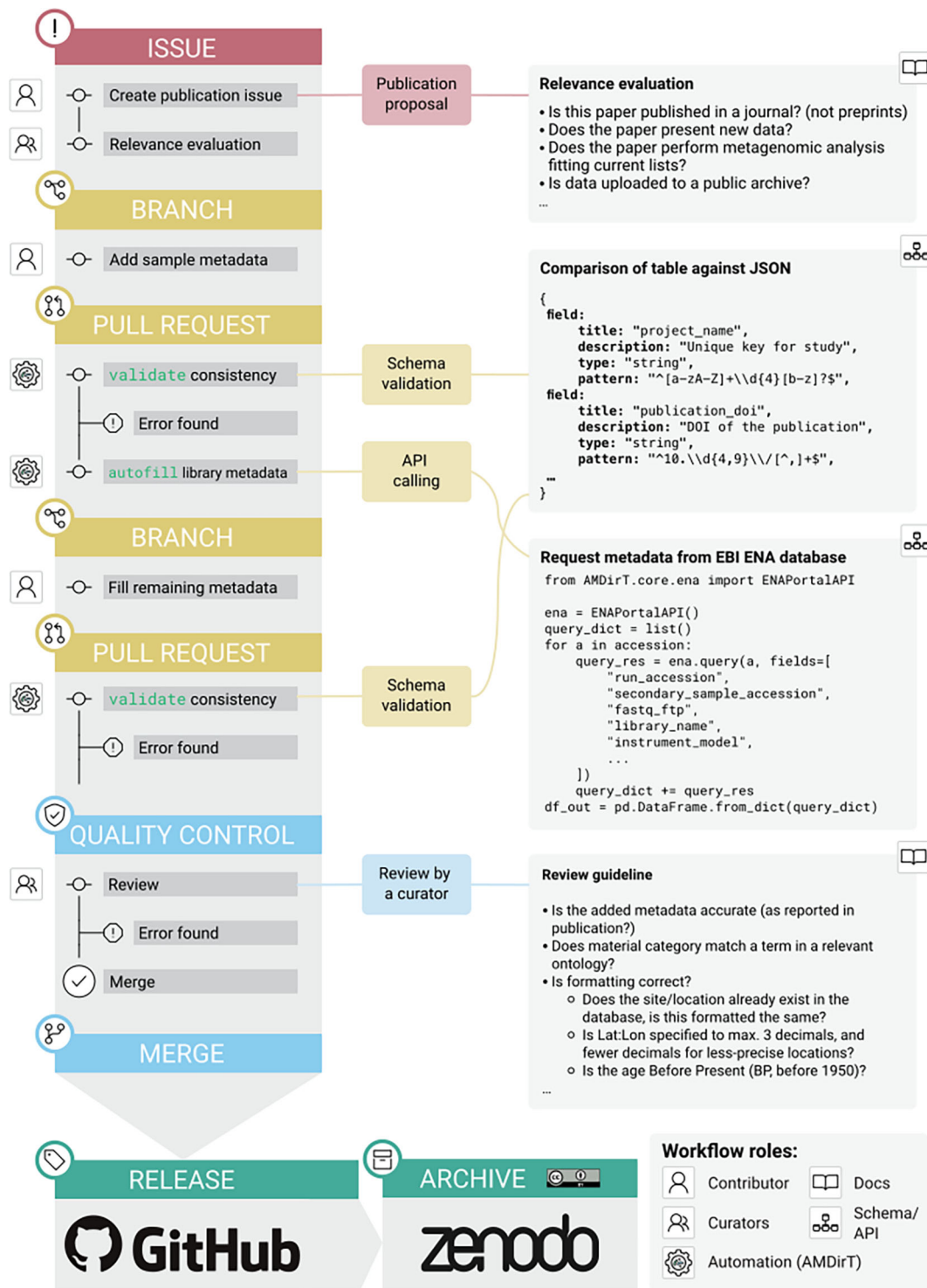



Figure 2. Updated workflow for submission to AncientMetagenomeDir using the AMDirT autofill functionality. The AncientMetagenomeDir submission workflow, as updated since.³ The general workflow remains the same, with issue creation for publication proposals, metadata submission by contributors via a branch and pull request, something that undergoes automated validation (with AMDirT validate), and later peer-review by AncientMetagenomeDir curators. The new addition is the use of autofill that is called via a GitHub Actions 'bot'. This generates and uploads to the pull request in a comment a partially completed library metadata table that can be filled in, reviewed for accuracy and appended to the corresponding AncientMetagenomeDir library table as a part of the original sample pull request.

The general help of AMDirT (v1.6) is available from the CLI:

```
$ AMDirT --help
Usage: AMDirT [OPTIONS] COMMAND [ARGS] ...

AMDirT: Performs validity check of AncientMetagenomeDir datasets
Authors: AMDirT development team and the SPAAM community
Homepage & Documentation: https://github.com/SPAAM-community/AMDirT

Options:
  --version Show the version and exit.
  --verbose Verbose mode
  --help Show this message and exit.

Commands:
  autofill Autofills library and/or sample table(s) using ENA API and..
  convert  Converts filtered samples and libraries tables to eager,..
  download Download a table from the AMDirT repository
  merge    Merges new dataset with existing table
  validate Run validity check of AncientMetagenomeDir datasets...
  viewer   Launch interactive filtering tool
```

Most tools follow a standard CLI based interface. For example, converting a user-filtered ancient metagenome host-associated AncientMetagenomeDir table (e.g. in R) to a curl download script can be performed as follows:

```
$ AMDirT convert --curl <filtered_table>.tsv ancientmetagenome-hostassociated -o ./
```

In the command above, options, input files and output files are defined with standard command line flags and positional arguments.

The resulting file AncientMetagenomeDir_curl_download_script.sh file from the command above will be present in the directory specified in the command. The user can then simply run the bash script to download all libraries of the samples present in the input table.

```
$ bash AncientMetagenomeDir_curl_download_script.sh
```

For the template pipeline input sheets, these can be supplied to the pipelines themselves, after checking for accuracy.

The other AMDirT tools follow a similar scheme, with help messages and documentation on the AMDirT website providing more how-to information (<https://amdir.readthedocs.io/>).

For the GUI-based viewer tool, a user simply enters the following command in their terminal, after which their web browser will automatically load. Alternatively, the reported local or network address can be manually entered into the user's web browser. In comparison to the convert subcommand, the input tables are automatically pulled from the AncientMetagenomeDir for the user, without requiring any manual input.

```
$ AMDirT viewer
AMDirT [INFO] :
[AMDirT] To close app, press on your keyboard: ctrl+c

You can now view your Streamlit app in your browser.

Local URL: http://localhost:8501
Network URL: http://172.16.9.75:8501
```

Once completed, the user can close the tab and cancel the command in their terminal (e.g., with ctrl + c).

Alternatively, for individuals who wish to use the viewer but do not wish to deal with software installation and/or are not comfortable with command line interfaces, a hosted online version of the AMDiRT viewer is available at <https://spaam-community.org/AMDiRT> accessible with a web browser.

Use cases

Here we will describe a common use case for when users may wish to use the AMDiRT package, namely filtering for a particular subset of metagenomic aDNA data, downloading the resulting data, generating a corresponding semi-prepared input sheet for nf-core/eager,⁵ and creating a citations file. Full tutorials in text and video format for the following and other AMDiRT commands can be found on the AMDiRT website (<https://amdir.readthedocs.io/>).

This example scenario demonstrates how a user can download all publicly available ancient host-associated metagenomes published since 2020 from samples originating from Spain using the AMDiRT GUI interface. In this hypothetical example, a user may wish to compare the microbial taxonomic profiles of archaeological dental calculus and other skeletal elements in Spain at different time points. In order to distinguish modern and aDNA, the user will likely want to examine the DNA for evidence of chemical degradation, which can be used to authenticate aDNA. To do this, the user will already have selected their preferred dedicated aDNA analysis workflow, such as nf-core/eager, that integrates DNA damage analysis into its pipeline. nf-core/eager requires an input 'sample sheet' that describes whether a particular sample has been sequenced over multiple lanes or libraries, and whether aDNA damage has been already removed during laboratory processing. We will show how AMDiRT can assist with the creation of this sample sheet with the desired dataset.

This example assumes that the user has already installed AMDiRT and nf-core/eager, and has downloaded a *Homo sapiens* reference genome for host DNA removal.

To load the GUI based viewer and downloading tool, a user enters the following command into their terminal:

```
$ AMDiRT viewer
```

A) Select Table

AMDiRT viewer tool
Version: 1.6.0
Select an AncientMetagenomeDir release: v23.12.0
Select a table: ancientmetagenome-hostassociated

Displayed table: ancientmetagenome-hostassociated

Project	Publication	Publication DOI	Site	Latitude	Longitude	anc. DNA source
Warren2014	2014	10.1038/ng.2006	Dachau	51.505	8.84	Germany
Warren2017	2017	10.1038/nature21118	Grotto Fc...	7.637	-85.862	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Stutgar...	48.859	9.227	Germany

B) Filter and select samples

AMDiRT viewer tool
Version: 1.6.0
Select an AncientMetagenomeDir release: v23.12.0
Select a table: ancientmetagenome-hostassociated

Displayed table: ancientmetagenome-hostassociated

Project	Publication	Publication DOI	Site	Latitude	Longitude	anc. DNA source
Warren2014	2014	10.1038/ng.2006	Dachau	51.505	8.84	Germany
Warren2017	2017	10.1038/nature21118	Grotto Fc...	7.637	-85.862	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Stutgar...	48.859	9.227	Germany

C) Filter and select libraries

AMDiRT viewer tool
Version: 1.6.0
Select an AncientMetagenomeDir release: v23.12.0
Select a table: ancientmetagenome-hostassociated

Displayed table: ancientmetagenome-hostassociated

Project	Publication	Publication DOI	Site	Latitude	Longitude	anc. DNA source
Warren2014	2014	10.1038/ng.2006	Dachau	51.505	8.84	Germany
Warren2017	2017	10.1038/nature21118	Grotto Fc...	7.637	-85.862	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Stutgar...	48.859	9.227	Germany

D) Download converted metadata

AMDiRT viewer tool
Version: 1.6.0
Select an AncientMetagenomeDir release: v23.12.0
Select a table: ancientmetagenome-hostassociated

Displayed table: ancientmetagenome-hostassociated

Project	Publication	Publication DOI	Site	Latitude	Longitude	anc. DNA source
Warren2014	2014	10.1038/ng.2006	Dachau	51.505	8.84	Germany
Warren2017	2017	10.1038/nature21118	Grotto Fc...	7.637	-85.862	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	El Sidr...	42.386	-5.028	Spain
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Soy Cave	50.68	4.074	Belgium
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Durka	51.96	21.86	Poland
Warren2017	2017	10.1038/nature21118	Stutgar...	48.859	9.227	Germany

Figure 3. Example workflow of using AMDiRT viewer. (a) The viewer opens in a user's web browser, where the desired AncientMetagenomeDir version and table is selected. (b) Interaction with columns follows standard operations common to most spreadsheet software. Samples for download are selected using checkboxes. (c) The same interface can be used for the subsequent library metadata filter table. (d) After pressing 'Validate library selection', buttons appear for downloading various download scripts, reference, and pipeline input sheets.

As shown in [Figure 3a](#), the viewer is loaded into the user's web browser. Using the left sidebar, the user can navigate drop-down menus to select the release of AncientMetagenomeDir to use (for reproducibility purposes), and the desired AncientMetagenomeDir table to explore (i.e., ancient environmental metagenomes, ancient host-associated metagenomes, or ancient microbial single-genomes). The user can also specify the number of rows for the table to display and which tool to use for the download scripts generated later in the example.

Once the AncientMetagenomeDir table is selected, the main window will load the corresponding table. The user can manipulate the columns as customary in most spreadsheet software, such as resizing the columns by dragging the bars between each column, dragging column names to reorder them, etc. Users can use keyboard arrows or scroll bars to navigate further along the columns. To filter the columns, the user can press the 'hamburger' menu of each column, which reveals a range of column operation options. In the example in [Figure 3b](#), the 'geo_loc_name' column has already been filtered to display only samples with the value 'Spain', and by pressing the funnel on an integer column, such as 'publication_year', the user can specify to only display rows 'greater than or equals' to 2020. Additional filter specifications can be added using the AND/OR operators in each filter menu. The user can then select which samples to be exported from AMDiT. They can either use the 'Select All' checkbox in the top left of the table, or select each sample using the checkboxes at the beginning of each row on a sample-by-sample basis.

Once the user is satisfied with their selection, they can press 'Validate sample selection' ([Figure 3c](#)). Once pressed, a new table will appear below the Sample metadata table. This new table contains all library-level metadata of the selected samples from the previous step. Users can then filter library-level metadata with the same interface and in the same way as the sample-level metadata. Once selected, and the 'Validate library selection' button has been pressed, a range of buttons representing different downloading options will appear.

Here, we recommend that users always download the corresponding AncientMetagenomeDir libraries table, as well as the BibTeX citation file. In this example, the user would also download the 'curl download script' and the 'nf-core/eager' input TSV using the corresponding buttons. Hovering over the download script button also provides an estimate for the user of how much hard-drive space the download of all selected data will use.

To close the GUI viewer, the user can close the tab in their browser, and then in their terminal press `ctrl + c` on their keyboard to stop the Streamlit server.

Alternatively, a user could also follow the same process but via the command line interface. The user first can run the new download (download) command to retrieve a particular samples or libraries table.

```
$ AMDirT download --table ancientmetagenome-hostassociated --table_type samples
--release v24.03.0
```

The user could then use typical shell commands, or load the table into languages such as R or Python and Pandas for filtering the table to the data they would be interested in. To replicate the same conditions as in [Figure 3b](#) above, a user could run the following command with the common CLI tool `awk`

```
$ awk -F "\t" 'NR==1 || $2 >= 2020 && $7 == "Spain"' ancientmetagenome-
hostassociated_samples_v24.03.0.tsv > ancientmetagenome-hostassociated_samples_
gt2020_spain.tsv
```

Finally, the user can pass the resulting filtered table to the AMDirT `convert` command, to perform the same download script generation, pipeline-input file conversion, and citation file download as with the GUI interface

```
$ AMDirT convert --librarymetadata --curl --eager --bibliography ancientmetagenome-
hostassociated_samples_gt2020_spain.tsv ancientmetagenome-hostassociated
```

The `convert` command will then generate the same files downloaded by the GUI (example above).

Once downloaded, the user should check the AncientMetagenomeDir libraries table to ensure that all desired libraries are present. If there are extra libraries with specifications that the user does not want, they should remove those entries from the curl download script and nf-core/eager input TSV files. For the CLI `convert` command, a user can also optionally supply a pre-filtered *libraries* table in addition to the samples table to reduce the need for manual editing of the

downloaded files. The user should also review the generated nf-core/eager pipeline input sheet to check for accuracy in regards to the pipeline's specifications.

After reviewing and filtering the scripts and pipeline input TSV sheets, the user can then use their terminal to navigate to a directory, move the curl script into it and begin the download. Due to the large sizes of sequencing data, in most cases we recommend that a user do this in a 'screen' or 'tmux' session (or similar) to ensure that the downloading can continue in the background:

```
$ bash AncientMetagenomeDir_curl_download_script.sh
```

Once the sequencing data are downloaded, the user can provide their AMDirT generated nf-core/eager input sheet to the following Nextflow¹⁹ command (the nf-core/eager input sheet assumes that the command is being run in the same directory as the downloaded data):

```
$ nextflow run nf-core/eager -r 2.4.6 -profile conda
-input AncientMetagenomeDir_nf_core_eager_input_table.tsv
-fasta hg19.fasta -outdir ./results
```

Discussion

Results of library level metadata aggregation

Since the original publication of AncientMetagenomeDir³ and the release of version v20.09, the SPAAM community has doubled the number of manually curated publications in the AncientMetagenomeDir from 87 to 187 studies as of version v24.03. The number of samples has increased from 443 to 1427 for ancient host-associated metagenome samples, 269 to 667 for ancient microbial genome level sequences, and 312 to 662 for sediment samples (Figure 1).

During the series of 'hackathon' events carried out by the community to scrape library metadata from previous publications and subsequent submissions of new studies, a total of 2557 ancient host-associated metagenome libraries, 3048 ancient microbial genomes libraries, and 754 ancient environmental metagenome libraries have been curated and included.

Common issues in ancient metagenomic library metadata

During the aggregation and clean-up of the library metadata by the SPAAM community, a range of problems were repeatedly encountered across multiple studies that made data entry and the determination of appropriate preprocessing procedures difficult. Here, we describe the most common issues encountered, as well as possible solutions, listed from most to least severe. By highlighting these common mistakes and problems, we hope to help improve (meta)data uploads to sequencing archives, which in turn will both benefit the AncientMetagenomeDir users, but also the field as a whole.

Inconsistent sample and library naming.

Problem: A common problem encountered when cross-referencing ENA or SRA metadata with information provided in original publications was inconsistencies in sample, library, and/or sequencing file names. This often made it difficult for the community member to correctly infer which library was associated with which sample, or even which sequencing file went with which library.

Example: In studies where two sets of libraries were generated (for example, one with UDG-based DNA damage removal and one without), this palaeogenomic-specific information was often not indicated in the library or file names. Given that this information is not supported in the ENA/SRA metadata schema, this is the only location where such aDNA-specific information could be reasonably recorded. In such cases, we found that while library pretreatment procedures were documented in the original publication, the uploaded metadata and sequencing files generally lacked this information and in some instances used internal laboratory IDs instead of the sample or library codes recorded in the publication. Without a key linking the published IDs with the internal laboratory IDs, other researchers cannot know which files to use for their particular analyses or how to process the data appropriately, and this can lead to downstream problems. For example, if a user does not know that a sequencing file was generated from damage-removed libraries, they may inappropriately apply additional *in silico* trimming steps to remove DNA damage, and thus unnecessarily truncate the sequences.

Solutions: We suggest two solutions: first, ancient metagenomic researchers should ensure that library and sample names are descriptive (i.e., in a structured system in which a certain level of information can be inferred just by the name) and that sequencing metadata uploads match those reported in the publication; second, where this is not possible (e.g., if an upload

is carried out by a third party), then researchers should at a minimum include a key in their supplementary files. This could be in the form of a table that includes all ID codes for each sample, library, and sequencing batch, including internal laboratory codes, other-analysis codes, and external sequencing archive accession codes.

Metadata discrepancies about sequencing methods.

Problem: Another relatively common issue was the discrepancies between the metadata reported in the sequence archive and in the original publication. It was generally difficult to resolve such discrepancies without contacting the authors. Discrepancies occurred most frequently in the reporting of the sequencing platform.

Example: In several cases, the particular sequencing platform recorded in the sequence archive metadata, such as ‘Illumina HiSeq 4000’, did not match that reported in the publication, e.g. ‘NextSeq 500’.

Solutions: Researchers should be sure to cross-reference their metadata upload sheets with their manuscripts prior to upload. In cases where Illumina sequencing was carried out externally (and where limited information may have been provided by the sequencing centre), researchers can generally inspect the headers of the FASTQ file to determine which platform was used, as in the example [provided here](#).

Methods description in secondary or tertiary citations.

Problem: For journals with strict word or character count limits, it was qualitatively observed that there was an increased tendency in these publications to rely on secondary or tertiary non-protocol specific citations for describing laboratory methods used for DNA library construction and sequencing. This practice is problematic as secondary or tertiary citations may describe multiple protocols, and it was not always possible to determine which protocol was actually used in the study.

Example: In one case, a publication reporting an ancient microbial genome reconstruction referred to library protocols used in an earlier related publication that described data generation for a human population genetics study. However, upon closer inspection, this cited study itself referred to an even earlier publication that included extensive protocol experimentation and development. Neither the primary nor secondary publication indicated which of the experimental protocols from the original methods study was actually used.

Solutions: Ancient metagenomic researchers should make an effort to more clearly describe their protocols and explicitly indicate which library protocol is linked to each sequencing file. At a minimum, the information provided should include critical metadata for downstream analysis, such as library treatment protocols that affect DNA damage. This can be accomplished by providing expanded, plain-language descriptions of laboratory methods in article supplementary information files (rather than simply citing and re-citing) and providing a supplementary table that lists each library name and their corresponding treatments. For improved compliance with FAIR principles, researchers are encouraged to further provide or cite a protocol written up in a citable protocol format and/or on open platforms. For example, platforms such as [protocols.io](#)²⁰ allow critical protocol information to be communicated consistently and unambiguously, by providing a persistent identifier (DOI) that points to a specific version of a given protocol.

Uploading of mapped BAM files or merged FASTQ files rather than raw metagenomic data.

Problem: Occasionally, we found that in some cases ancient metagenomic researchers uploaded mapped BAM files or merged FASTQ files rather than ‘raw’ FASTQ files (i.e., against ENA/SRA specifications). Both formats present obstacles for downstream analysis. For example, mapped BAM files include only reads mapped to a particular reference genome and thus do not represent a full metagenomic dataset. For BAM files containing reads mapped to the human genome, microbial DNA will be absent, including ancient pathogen DNA that could be highly relevant for an archaeological study. While an ‘unmapped’ BAM ([uBAM](#)) format exists and FASTQ files can be partly reconstructed from such data, raw FASTQ files are the preferred format for data reporting. Unmapped BAM files indicate that a certain level of data preprocessing has already occurred, and such files often combine multiple libraries into a single BAM file in order to achieve sufficient genomic coverage for analysis. If the process of generating the BAM file is not sufficiently described, it can be difficult for other researchers to disentangle the data originating from different libraries or sequencing batches prior to reanalysis. Providing FASTQ files containing merged paired-end reads also limits data reuse. Although read merging is a common first step in some ancient bioinformatics pipelines, it is incompatible with others. Base quality scores are often altered during the read merging process, which can interfere with tools reliant on such scores, and most *de*

novo sequence assemblers either require or perform better on unmerged reads. Furthermore, many metagenomic tools, including taxonomic classifiers, do not accept merged paired-end FASTQ files or BAM files as an input format.

Example: An ancient metagenomic researcher generates both damage and damage-removed libraries, but merges them together in BAM format and uploads to a sequencing archive. However another researcher wishes to analyse only the data deriving from the damage-removed libraries.

Solution: Ideally researchers should upload FASTQ files that match the ‘raw’ output from sequencing, i.e., demultiplexed datasets separated per library, applying only the preprocessing steps recommended by the sequence repository (e.g., for the ENA/SRA, adapter removal but not read merging). If this is not possible, authors should, at a minimum, describe exactly how the merging steps were performed so that other researchers can manually separate merged sequences (e.g., using sequencing read headers) when required for downstream analysis.

Unique sample accessions applied to multiple libraries of the same sample.

Problem: Another common error was found to occur when researchers mistakenly uploaded each library or sequencing dataset with a unique sample accession code. While often not a critical error, because in these cases the correct sample could usually be inferred from the file name, this nevertheless makes automated data processing more difficult and requires manual intervention. To reuse such data, a researcher must manually reassociate the library datasets with the correct sample based on the file names, rather than relying on the sequence archive sample accession ID, as expected by metadata schemas of the ENA/SRA data repositories.

Solution: Researchers should review sequencing archive documentation to ensure they correctly construct upload sheets at both the library and sample levels (e.g. <https://ena-docs.readthedocs.io/en/latest/submit/general-guide/metadata.html>). Furthermore, researchers should ensure that library names have a consistent pattern such that other researchers can unambiguously associate each library with the correct sample.

Note on AMDiT generated pipeline scripts

It is important to note that the aim of the pipeline TSV sheets generated by AMDiT is to provide a *template* for data input to the pipelines. Due to the high heterogeneity in the way that sequence (meta) data are uploaded, some information in AncientMetagenomeDir may be missing or erroneous, despite the best efforts of the SPAAM community experts to standardise the information, resolve ambiguities, and correct errors. However, we hope that by providing this functionality, it reduces the time it takes to create such input sheets from scratch.

Future development

We envision that the future development of the AncientMetagenomeDir project will be to further extend and also standardise the types of metadata currently recorded. For example, when recording the age of samples, AncientMetagenomeDir currently only records a single value of an approximate date. This poses challenges for analyses requiring exact dates and probability intervals such as tip dating for phylogenetic trees and other analyses of evolutionary divergence. At present, however, heterogeneity in the reporting of radiocarbon dates (the most common dating method in archaeology and palaeogenomics) and associated modelling information currently limits our ability to add such dating information to AncientMetagenomeDir and to consistently apply calibration and reservoir effect correction across studies. This is despite the fact that there is already standard reporting guidance.²¹ However, we also call on ancient metagenomics researchers to report both uncalibrated *and* calibrated dates and associated metadata (radiocarbon lab code, calibration curve, software, etc.), and not to rely solely on secondary citations to facilitate adding such data to repositories such as AncientMetagenomeDir, as well as refinement of chronological modelling in the future.

In the same vein, we also aim to synchronise AncientMetagenomeDir with upstream standardised sequencing data metadata schemas and repositories such as MInS checklists²² via another SPAAM-established project, MInAS (<https://mixs-minas.org/>), to further ensure common standards across both modern and ancient sequencing data.

Given that the functionalities of AMDiT provide simple data exploration without requiring advanced computational knowledge, but also offers semi-prepared templates for aDNA and metagenomics, the dataset and tooling are ideal for further generation of community resources. Following other projects that have been developed for modern microbiome data,²³ the ancient metagenomics community could also consider providing standardised and pre-made taxonomic profiles (e.g., for microbiome or environmental samples) or VCF files (for single genomes) that could allow integration into current analysis workflows to assist users in more rapidly integrating public data into their analyses from a single source. This could be particularly useful for screening ancient microbiome samples for preservation (e.g., by comparing a

newly sequenced sample to all previously published ancient metagenomes), in order to assess whether a sample falls within the variation of known well-preserved or environmentally degraded samples.

Conclusions

By extending AncientMetagenomeDir to include library-level metadata, not only do we make ancient metagenomics data more findable, but also we make them more accessible by providing improved transparency of the diverse library and sequencing treatments performed in the field of palaeogenomics. Furthermore, AMDirT has been designed to improve the experience of researchers in the downloading and processing of previously published ancient metagenomics data. By providing both a graphical user interface and a command-line interface to filter and generate relevant download scripts and input sample sheets for aDNA analyses, we provide more flexibility and choice for the wide range of computational backgrounds that ancient metagenomic researchers can have. Finally, we hope that by informing researchers about inconsistencies in past data and metadata uploads and providing templates of standardised metadata for future publications, we will contribute to improving aDNA data reporting and FAIR data sharing.

Data availability

Source data

The source data for the sample-level metadata used by AMDirT is from the AncientMetagenomeDir project originally published in Ref. 3 under a CC-BY 4.0 license.

The existing sample-level and new library-level data is stored on GitHub:

<https://github.com/SPAAM-community/AncientMetagenomeDir>

Each release is archived on Zenodo: <https://doi.org/10.5281/zenodo.3980833>.

Underlying data

New sequencing library-level metadata is also stored in the AncientMetagenomeDir project from version v22.09 (Pyu Ancient Cities) onwards.¹² The version of the dataset used for the demonstration of AMDirT, statistics, and figures in this updated manuscript is v24.03 (Monticello).²⁴

Zenodo: SPAAM-community/AncientMetagenomeDir: Monticello (v24.03). <https://doi.org/10.5281/zenodo.10942606>.

Data are available under the terms of the [Creative Commons Attribution 4.0 International license](#) (CC-BY 4.0).

Software availability

- Software available from:
 - PyPi: <https://pypi.org/project/AMDirT/>
 - Bioconda: <https://bioconda.github.io/recipes/amdirt/README.html>
 - Hosted web version of AMDirT viewer: <https://spaam-community.org/AMDirT>
- Source code available from: <https://github.com/SPAAM-community/AMDirT>
- Archived source code at time of publication revision (AMDirT v1.6): <https://doi.org/10.5281/zenodo.10941007>
- License: GNU General Public License v3.0.

Acknowledgements

The authors thank the SPAAM community for the ongoing maintenance, testing, and general support of both the AMDirT and AncientMetagenomeDir projects. We also thank all the supervisors and managers of all the authors for allowing us to contribute to the AncientMetagenomeDir and AMDirT projects. We are also grateful for all authors of publications who we contacted with queries about their metadata, particularly those who subsequently took the time to update their original sequencing archive uploads to correct mistakes or improve metadata of their given study.

References

1. Anagnostou P, Capocasa M, Milia N, *et al.*: **When data sharing gets close to 100%: what human paleogenetics can teach the open science movement.** *PLoS One*. March 2015; **10**(3): e0121409. 1932-6203.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
2. Wilkinson MD, Dumontier M, Aalbersberg IJJ, *et al.*: **The FAIR guiding principles for scientific data management and stewardship.** *Sci. Data*. March 2016; **3**: 160018. 2052-4463.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
3. Fellows Yates JA, Andrades Valtueña Á, Vågane AJ, *et al.*: **Community-curated and standardized metadata of published ancient metagenomic samples with AncientMetagenomeDir.** *Sci. Data*. January 2021; **8**(1): 31. 2052-4463.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
4. Schubert M, Ermini L, Der Sarkissian C, *et al.*: **Characterization of ancient and modern genomes by SNP detection and phylogenomic and metagenomic analysis using PALEOMIX.** *Nat. Protoc.* May 2014; **9**(5): 1056-1082. 1754-2189, 1750-2799.
[PubMed Abstract](#) | [Publisher Full Text](#)
5. Fellows Yates JA, Lamnidis TC, *et al.*: **Reproducible, portable, and efficient ancient genome reconstruction with nf-core/eager.** *PeerJ*. March 2021; **9**: e10947. 2167-8359.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
6. Pochon Z, Bergfeldt N, Kirdök E, *et al.*: **aMeta: an accurate and memory-efficient ancient Metagenomic profiling workflow.** *bioRxiv*. October 2022; page 2022.
[Publisher Full Text](#)
7. Krakau S, Straub D, Gourel H, *et al.*: **nf-core/mag: a best-practice pipeline for metagenome hybrid assembly and binning.** *NAR Genom. Bioinform.* January 2022; **4**(1).
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [Reference Source](#)
8. Orlando L, Allaby R, Skoglund P, *et al.*: **Ancient DNA analysis.** *Nat. Rev. Methods Primers*. February 2021; **1**(1): 1-26.
[Publisher Full Text](#)
9. Eaton K: **NCBImeta: efficient and comprehensive metadata retrieval from NCBI databases.** *J. Open Source Softw.* February 2020; **5**(46): 1990. 2475-9066.
[Publisher Full Text](#)
10. Ewels P, Duncan A, Fellows Yates JA: **ewels/sra-explorer: Version 1.0.** March 2023.
[Reference Source](#)
11. Gálvez-Merchán Á, Min KHJ, Pachter L, *et al.*: **ffq: A tool to find sequencing data and metadata from public databases.** 2022.
[Reference Source](#)
12. Fellows Yates JA, Andrades Valtueña A, Vågane AJ, *et al.*: **SPAAM-community/AncientMetagenomeDir: v22.09.2.** August 2022.
[Reference Source](#)
13. Grünig B, Dale R, Sjödin A, *et al.*: **Bioconda: sustainable and comprehensive software distribution for the life sciences.** *Nat. Methods*. July 2018; **15**(7): 475-476.
[PubMed Abstract](#) | [Publisher Full Text](#)
14. Harrison PW, Ahamed A, Aslam R, *et al.*: **The european nucleotide archive in 2020.** *Nucleic Acids Res.* January 2021; **49**(D1): D82-D85.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [Reference Source](#)
15. The pandas development team: **pandas-dev/pandas: Pandas.** 2020.
[Publisher Full Text](#)
16. Fonseca P: **streamlit-aggrid: Implementation of Ag-Grid component for streamlit.** 2023.
[Reference Source](#)
17. Python Packaging Authority: **setuptools: Official project repository for the setuptools build system.** 2023.
[Reference Source](#)
18. Dabney J, Meyer M, Pääbo S: **Ancient DNA damage.** *Cold Spring Harb. Perspect. Biol.* July 2013; **5**(7): 1943-0264.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
19. Di Tommaso P, Chatzou M, Floden EW, *et al.*: **Nextflow enables reproducible computational workflows.** *Nat. Biotechnol.* April 2017; **35**(4): 316-319.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. Teytelman L, Stoliartchouk A, Kindler L, *et al.*: **Protocols.io: Virtual communities for protocol development and discussion.** *PLoS Biol.* August 2016; **14**(8): e1002538.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Millard AR: **Conventions for reporting radiocarbon determinations.** *Radiocarbon*. 2014; **56**(2): 555-559.
[Publisher Full Text](#) | [Reference Source](#)
22. Yilmaz P, Kottmann R, Field D, *et al.*: **Minimum information about a marker gene sequence (MIMARKS) and minimum information about any (x) sequence (MIXS) specifications.** *Nat. Biotechnol.* May 2011; **29**(5): 415-420.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
23. Pasolli E, Schiffer L, Manghi P, *et al.*: **Accessible, curated metagenomic data through ExperimentHub.** *Nat. Methods*. October 2017; **14**(11): 1023-1024.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
24. Fellows Yates JA, Andrades Valtueña A, Vågane AJ, *et al.*: **SPAAM-community/AncientMetagenomeDir: v23.03.0: Rocky necropolis of pantolica.** March 2023.
[Reference Source](#)

Open Peer Review

Current Peer Review Status:    

Version 2

Reviewer Report 27 September 2024

<https://doi.org/10.5256/f1000research.164792.r318837>

© 2024 Griffiths E. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Emma Griffiths

Simon Fraser University, Burnaby, USA

The manuscript entitled “Facilitating accessible, rapid, and appropriate processing of ancient metagenomic data with AMDirT” describes a suite of tools enabling automated population and validation of contextual data (metadata) for samples and libraries in the AncientMetagenomeDir collection. AncientMetagenomeDir is a previously published resource containing sets of tables of curated and standardized metadata for metagenomic and microbial genome datasets generated from ancient samples. AMDirT (AncientMetagenomeDir Toolkit), was developed to facilitate/automate data retrieval and curation from INSDC BioProjects, BioSamples, and SRA/GenBank to support AncientMetagenomeDir. The tooling enabled the addition of library metadata to AncientMetagenomeDir, which is required for many downstream analytical pipelines. Improvements included the addition of a CLI, a GUI, and a web version of AncientMetagenomeDir. Both the GUI and CLI methods can generate standardized templates of metadata, and both a wealth of supporting materials and documentation for users are provided. Code for AMDirT is openly provided on GitHub under a CCBY 4.0 license, and is also available via BioConda.

Development of AMDirT was very community-driven, via different hackathons. The developers appear to be highly collaborative (evidenced by the long author and affiliation list acknowledging the efforts of different community members). Improving the quality and findability of metadata and datasets is important, if unfortunately underappreciated, work. AncientMetagenomeDir is a well-curated resource that will likely be very useful to those investigating ancient DNA. Automation of curation and validation is key to sustaining metadata quality and for growing the resource. As such, AMDirT provides important tooling. Furthermore, many use cases and scripts were provided to illustrate how AMDirT can be implemented. The Problem-Solution section in the Discussion was great, and really highlighted issues that aren’t solved by the tooling but could be solved by the adoption of best (or at least better) practices by data providers.

To really understand the importance of AMDirT, I had to backtrack through several citations, and review a lot of the AncientMetagenomeDir documentation (click a lot of links). The authors have done their best to be concise and not be redundant in describing the AncientMetagenomeDir as it is already published, but this is sometimes to the detriment of clarity for the reader of the current manuscript. As such, Figure 2 was critical for making sense of AMDirT. Figure 3 was helpful in illustrating important components of the interface. What would have helped in the reading of the

current manuscript, would be a table of the fields for samples and libraries so the reader could understand the metadata types that AMDirT was retrieving at-a-glance. Furthermore, an expanded discussion of the need/activities to align AncientMetagenomeDir fields and terms with community standards (i.e. MInS) that is currently underway, would further demonstrate the authors' commitment to FAIR data and interoperability. This is important work and should not be glossed over. Further suggestions about the integration of data standards are provided below.

This manuscript has already been reviewed several times, and the authors have worked hard to improve the software and paper. If the authors expand their discussion of their plans for integrating community standards, then I would recommend this manuscript for indexing.

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?

Figure 2 mentions the use of ontologies, but from my investigations, ontologies are not used (there is some controlled vocabulary for the picklist enums for some fields, but these are not ontologies - ontologies use logical relationships to link the controlled vocabulary, and terms are defined and assigned universal identifiers e.g. OBO Foundry ontologies). Are the enums sourced from particular ontologies e.g. ENVO, Uberon? The authors mention that they are participating in another project to implement/further develop a new MInS package, which is wonderful. It would be great if the authors could expand on that effort in the discussion so the reader doesn't have to go to so many external sites to learn more, as this seems to be a crucial effort for building interoperability. In addition to this alignment with community standards, the enum controlled vocabulary would benefit from being aligned with community-based ontologies. Ontology terms can be sourced using the EBI Ontology Look-up Service. Furthermore, there has been work in the ontology community to better structure sample material descriptions. For example, there are some enums that precompose concepts that may make them difficult to compare across resources e.g. "skin from leg" precomposes anatomical material (skin) and anatomical parts (leg), which could also be written as "leg skin" as a synonym in other databases, and the precomposition opens the resource up to the "word bomb" problem i.e. you would need lots of terms to capture skin from other parts of the body (skin from arm, skin from foot, skin from knee). Better to separate "material" into different fields e.g. anatomical material, anatomical part, environmental material, environmental site, collection device, collection method. Some work has been done on this by PHA4GE and a similar effort can be seen in the NCBI One Health Enterics package. Separating environmental fields from anatomical fields has also been helpful, and could improve AncientMetagenomeDir annotations. For example, a data provider could include that a Homo sapiens dental calculus (anatomical material) sample was sourced from a desert (environmental site) vs a tomb, which could provide more context for the data.

Since all of the data is derived from published work, have the authors considered including additional fields for sample processing (e.g. pooling, subsampling), quality control, bioinformatic processing (e.g. dehosting)? There are ontology-based standardized fields available for these methods that may also be informative for users.

References

1. Borry M, Forsythe A, Andrades Valtueña A, Hübner A, et al.: Facilitating accessible, rapid, and appropriate processing of ancient metagenomic data with AMDirT. *F1000Res.* 2023; **12**: 926
[PubMed Abstract](#) | [Publisher Full Text](#)

Is the rationale for developing the new software tool clearly explained?

Yes

Is the description of the software tool technically sound?

Yes

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?

Partly

Is sufficient information provided to allow interpretation of the expected output datasets and any results generated using the tool?

Partly

Are the conclusions about the tool and its performance adequately supported by the findings presented in the article?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: data curation, data standards, ontologies, pathogen genomics, data sharing, data interoperability

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Reviewer Report 10 September 2024

<https://doi.org/10.5256/f1000research.164792.r318840>

© 2024 Alam I. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Intikhab Alam

King Abdullah University of Science and Technology, Thuwal, Makkah Province, Saudi Arabia

Borri et al. work entitled "" covers an important topic on metadata of samples. Historically, this is a developing field where initially such data sets were available with minimal metadata. As the field progressed, importance of comprehensive metadata was acknowledged and standards were proposed. However integrating older and new samples with different level of depth in metadata curation is a challenge. This work tries to improve upon such data curation and developed a tool AMDirT with GUI and commandline interface for users to explore metadata.

Looking at the GUI interface, it appears to have issues when using Firefox browser, the page does not load. Looking at developer tools there are some issues with the javascripts (e.g. stitle.js) saying

"not same-origin") and the following details:

Security Error: Content at <https://www.spaam-community.org/AMDirT/> may not load data from <https://cdn.jsdelivr.net/npm/@stlite/mountable@0.52.1/build/static/js/8779.995e8fcf.chunk.js>.

However, it appears to work when using google chrome browser, allowing to explore the data and ability to download relevant tables.

The command line version is difficult to install due to upgrades and other dependencies. It would be great if a version of AMDirT is available for singularity etc.

Overall, it looks like a very good tool and useful for the community and it is recommended to expand this tool to other types of metagenomes.

Is the rationale for developing the new software tool clearly explained?

Yes

Is the description of the software tool technically sound?

Yes

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?

Partly

Is sufficient information provided to allow interpretation of the expected output datasets and any results generated using the tool?

Partly

Are the conclusions about the tool and its performance adequately supported by the findings presented in the article?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Metagenomics of the biosphere

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 11 Sep 2024

Maxime Borry

Thank you for your positive comments on your manuscript. In regards to the two reservations (using the serverless version in Firefox, and installation)

1. We already explicitly state in the text (Methods > Implementation > Second

paragraph), that the streamlit to WebAssembly functionality that is only supported on Chromium-based browsers. This is a known limitation of the Firefox security policies regarding WebAssembly, and unfortunately not something we have influence on for now. We have added a permanent issue in the AMDirT Github repository (github.com/SPAAM-community/AMDirT/issues/158) to inform users who might be facing this issue.

2. Installation is already possible via pip and conda (both of which handle dependencies). As stated in the README, and the manuscript, we recommend installation in a dedicated environment when using conda. Furthermore, one of the benefit of having our software on bioconda is the complementary biocontainer project that automatically generates docker and singularity containers for the tool. You can get a docker version of our tool from [quay.io](https://quay.io/repository/biocontainers/amdirt?tab=tags&tag=latest) (<https://quay.io/repository/biocontainers/amdirt?tab=tags&tag=latest>), and singularity from the galaxy project (<https://depot.galaxyproject.org/singularity/>)

We hope these two points address your comments!

Maxime Borry & James Fellows Yates

Competing Interests: No competing interests were disclosed.

Version 1

Reviewer Report 18 October 2023

<https://doi.org/10.5256/f1000research.147881.r210768>

© 2023 Kasmanas J. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Jonas Coelho Kasmanas

¹ University of São Paulo, São Paulo, Brazil

² University of São Paulo, São Paulo, Brazil

The submitted manuscript sheds light on an imperative yet often overlooked aspect of current research, specifically metagenomic research: the accessibility and standardization of sample-level metadata, particularly concerning ancient samples. The topic is relevant to the current scientific landscape, especially as the volume of metagenomic sequencing datasets continues to burgeon. With the surge in data, the need for standardized, accessible, and reusable metadata becomes paramount. This manuscript aptly addresses this gap, making it an essential read for researchers in metagenomics, palaeogenomics, and related fields.

The authors have channeled the collective effort of the research community through hackathon events to update and enhance the 'AncientMetagenomeDir.' Even contacting researchers and

actively encouraging them to correct their metadata annotation. Therefore, I understand that this manuscript evolves the AncientMetagenomeDir and provides an additional resource to further increase its accessibility and usability: the AMDiT.

The introduction of 'AMDirT' as the companion tool to 'AncientMetagenomeDir' is a significant stride forward. By offering functionalities ranging from metadata template generation, and guidance to data validation, the tool exhibits promise in addressing the prevalent challenges in metadata management. Additionally, it helps researchers with less computational background to download samples systematically and gives them access to a GUI. My review goes toward the message delivery, especially during the initial sections of the manuscript.

During the introduction section, I missed a clear definition of ancient metagenome. This should include a description of what requires the ancient samples to receive special treatment regarding metadata annotation. Naturally, this leads to comparing your effort to past works like the GOLD (<https://doi.org/10.1093/nar/gky977>), the MetaSRA (<https://doi.org/10.1093/bioinformatics/btx334>), the gcMeta (<https://doi.org/10.1093/nar/gky1008>), or the HumanMetagenomeDB (<https://doi.org/10.1093/nar/gkaa1031>).

I appreciate the extensive AMDiT documentation, including tutorials and videos. However, the manuscript lacks in the description of the tool's workflow and usability. The "Use cases" section of the manuscript makes the tool much clearer. At the same time, it gives the impression that AMDiT is limited to the GUI exploration and template generation guidance. Specifically, in my opinion, the manuscript poorly describes and explores the "Validate" and "Autofill" commands.

For example, the Validate command does "a variety of checks." It would help if you made it more transparent. Do you have a document detailing the checks and standards? If so, it should be placed here; if not, I would like to see it in the manual. For the Autofill command, I missed a more detailed description of the "improvements" done using R scripts after pulling the metadata from ENA. Specifically, since those commands could be used outside the realm of ancient metagenomic samples. I believe a figure containing the complete schematic workflow from AMDiT would help.

Regarding the software operation, I missed a storage requirement specification. The paper mentions that the installation requires Python 3.9, but nothing is specified in the GitHub. The installation via pip and conda should include guidance on accessing those tools and creating a specific environment for the AMDiT. Finally, as a minor opinion, have you considered using specialized sample accessing software? For instance, the SRAtoolkit can download samples from the SRA more efficiently than curl.

If possible, I would like to see a slight reformat in the discussion "Problem/Solutions" section to make it more visible. A simple extra paragraph or bullet points that clearly separate problems and solutions should solve the problem.

Running and installing the pipeline.

I had a relatively smooth experience. However, I kept receiving the following warning upon any usage:

```
"WARNING streamlit.runtime.caching.cache_data_api: No runtime found, using  
MemoryCacheStorageManager"
```

Additionally, the convert command did not work. After successfully installing the tool and

downloading the AncientMetagenomeDir. I received the following:

```
$ AMDirT convert --curl SPAAM-community-AncientMetagenomeDir-60f8a00/ancientmetagenome-hostassociated/libraries/ancientmetagenome-hostassociated_libraries.tsv ancientmetagenome-hostassociated -o ./
```

2023-10-17 19:50:32.289 No runtime found, using MemoryCacheStorageManager

Traceback (most recent call last):

```
File "/mnt/tools/miniconda3/envs/amdirt/lib/python3.9/site-packages/streamlit/runtime/caching/storage/in_memory_cache_storage_wrapper.py", line 87, in get
```

```
    entry_bytes = self._read_from_mem_cache(key)
```

```
File "/mnt/tools/miniconda3/envs/amdirt/lib/python3.9/site-packages/streamlit/runtime/caching/storage/in_memory_cache_storage_wrapper.py", line 137, in _read_from_mem_cache
```

```
    raise CacheStorageKeyNotFoundError("Key not found in mem cache")
```

```
streamlit.runtime.caching.storage.cache_storage_protocol.CacheStorageKeyNotFoundError: Key not found in mem cache
```

Traceback (most recent call last):

```
File "/mnt/tools/miniconda3/envs/amdirt/lib/python3.9/site-packages/pandas/core/indexes/base.py", line 3790, in get_loc
```

```
    return self._engine.get_loc(casted_key)
```

```
File "index.pyx", line 152, in pandas._libs.index.IndexEngine.get_loc
```

```
File "index.pyx", line 181, in pandas._libs.index.IndexEngine.get_loc
```

```
File "pandas/_libs/hashtable_class_helper.pxi", line 7080, in pandas._libs.hashtable.PyObjectHashTable.get_item
```

```
File "pandas/_libs/hashtable_class_helper.pxi", line 7088, in pandas._libs.hashtable.PyObjectHashTable.get_item
```

```
KeyError: 'archive_accession'
```

```
File "/mnt/tools/miniconda3/envs/amdirt/lib/python3.9/site-packages/streamlit/runtime/caching/storage/in_memory_cache_storage_wrapper.py", line 137, in _read_from_mem_cache
```

```
    raise CacheStorageKeyNotFoundError("Key not found in mem cache")
```

```
streamlit.runtime.caching.storage.cache_storage_protocol.CacheStorageKeyNotFoundError: Key not found in mem cache
```

References

1. Mukherjee S, Stamatis D, Bertsch J, Ovchinnikova G, et al.: Genomes OnLine database (GOLD) v.7: updates and new features. *Nucleic Acids Res.* 2019; **47** (D1): D649-D659 [PubMed Abstract](#) | [Publisher Full Text](#)
2. Bernstein MN, Doan A, Dewey CN: MetaSRA: normalized human sample-specific metadata for the Sequence Read Archive. *Bioinformatics.* 2017; **33** (18): 2914-2923 [PubMed Abstract](#) | [Publisher Full Text](#)
3. Shi W, Qi H, Sun Q, Fan G, et al.: gcMeta: a Global Catalogue of Metagenomics platform to support the archiving, standardization and analysis of microbiome data. *Nucleic Acids Res.* 2019; **47** (D1): D637-D648 [PubMed Abstract](#) | [Publisher Full Text](#)
4. Kasmanas JC, Bartholomäus A, Corrêa FB, Tal T, et al.: HumanMetagenomeDB: a public

repository of curated and standardized metadata for human metagenomes. *Nucleic Acids Res.* 2021; **49** (D1): D743-D750 [PubMed Abstract](#) | [Publisher Full Text](#)

Is the rationale for developing the new software tool clearly explained?

Yes

Is the description of the software tool technically sound?

Yes

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?

Yes

Is sufficient information provided to allow interpretation of the expected output datasets and any results generated using the tool?

Yes

Are the conclusions about the tool and its performance adequately supported by the findings presented in the article?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Bioinformatics, metagenome, metadata standardization, machine learning

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 06 May 2024

James Fellows Yates

Thank you for your generally positive review. We apologise for the long time between your review and our revision, this was due to parental leave and extensive travelling of the corresponding authors.

Our responses to your specific comments are below. Line numbers in our revised manuscript should correspond to the submitted revised word document (required by F1000), once track changes are accepted. Please note depending on which software we opened it, the line numbers were not indicating the same content. The line numbers should correspond to what is opened in Microsoft Office for Mac Word v16.78.3.

Manuscript Comments

- *During the introduction section, I missed a clear definition of ancient metagenome. This should include a description of what requires the ancient samples to receive special*

treatment regarding metadata annotation. Naturally, this leads to comparing your effort to past works like the GOLD (<https://doi.org/10.1093/nar/gky977>), the MetaSRA (<https://doi.org/10.1093/bioinformatics/btx334>), the gcMeta (<https://doi.org/10.1093/nar/gky1008>), or the HumanMetagenomeDB (<https://doi.org/10.1093/nar/gkaa1031>). I appreciate the extensive AMDiT documentation, including tutorials and videos. However, the manuscript lacks in the description of the tool's workflow and usability. The "Use cases" section of the manuscript makes the tool much clearer. At the same time, it gives the impression that AMDiT is limited to the GUI exploration and template generation guidance. Specifically, in my opinion, the manuscript poorly describes and explores the "Validate" and "Autofill" commands. For example, the Validate command does "a variety of checks." It would help if you made it more transparent. Do you have a document detailing the checks and standards? If so, it should be placed here; if not, I would like to see it in the manual.

Response: For the ancient metagenome definition: this was originally defined in the AncientMetagenomeDir publication but we've added a short sentence rephrasing and citing that definition on line 36-37.

For validate: we do not go into much detail on this, as it again was originally described in the first AncientMetagenomeDir paper (as cited in the main text) and as such does not necessarily represent 'new' functionality other than a renaming. Furthermore, as it is not a user facing component of the tool kit, we feel it is less relevant for readers (who we presume many more will be potential users rather than developers wishing to adopt a similar scheme). However we have rephrased the text slightly to indicate which person each command is most relevant to (e.g. contributor vs maintainer vs user etc.). The same goes for autofill, in that it is not a user facing tool - as it typically will be just within GitHub actions. We have however updated the manuscript to provide a little more detail on the validate command (line 105-140) and for autofill (which calls the ENA API for information) in lines 100-103. We have otherwise also expanded the use case to also describe in more detail the 'convert' CLI command, which is also a primarily user-facing subcommand.

- *For the Autofill command, I missed a more detailed description of the "improvements" done using R scripts after pulling the metadata from ENA. Specifically, since those commands could be used outside the realm of ancient metagenomic samples. I believe a figure containing the complete schematic workflow from AMDiT would help.*

Response: We have removed references to the R scripts used in the very initial pulling of data in preparation for the first library-level hackathon. On reflection, indeed this could be confusing to the reader, particularly as these are no longer used - and were only used for a one off event. The same concepts have been added to autofill (which came later). In autofill, the ENA portal API is directly queried using Python. We have added more details on the retrieved metadata fields (line 100-103).

We have further now reduced the necessity of a user to run this command at all (see above about not being a user-facing tool), instead having a GitHub actions bot to run this command for a user on the AncientMetagenomeDir repository, during a pull request. We have included a new figure (Figure 2) that shows the updated AncientMetagenomeDir workflow modified after the original AncientMetagenomeDir publication workflow diagram.

- *Regarding the software operation, I missed a storage requirement specification. The paper*

mentions that the installation requires Python 3.9, but nothing is specified in the GitHub. The installation via pip and conda should include guidance on accessing those tools and creating a specific environment for the AMDirT.

Response: We unfortunately don't fully understand what the reviewer means exactly by 'storage requirement specification' here. However, based on our interpretation: all downloadable AMDirT files are small bash scripts, TSV, or bibtext files, all of which take up (bioinformatically) negligible amounts of space - single digit megabytes or less. Thus we do not feel it is necessary to mention this. However if the storage requirements refers to FASTQ files etc, as a convenience, AMDirT offers an estimate of the potential total size of the files if you were to run the download script. This is displayed by hovering over the download script download button - however AMDirT does not download FASTQ files itself. This functionality was already described on lines 452.

The AMDirT package zip archive itself is also less than a megabyte so we feel is not relevant here as it will not make an impact on storage requirements in the vast majority of cases. For installation requirements: we are not entirely sure what the reviewer means by 'accessing these tools' - whether this is the downstream dependencies, or the package managers pip/conda themselves. If the former, the version of dependencies management is automatically handled by pip/conda, and is completely transparent for the user as this is reported by each package manager when installing AMDirT. For advanced bioinformaticians, they are likely familiar with python packages setup.py and conda environment files, so should be able to find these themselves (as an everyday user does not need this). If the reviewer is referring to installation instructions of pip and conda, this is a fair point and we have added in the installation instructions URLs to the installation documentation pages of the respective package managers, and we have updated the README to provide updated guidelines to install AMDirT in a dedicated conda environment.

- *Finally, as a minor opinion, have you considered using specialized sample accessing software? For instance, the SRAToolkit can download samples from the SRA more efficiently than curl.*

Response: As an alternative to curl, AMDirT already provides download scripts using other tools such as aspera and [nf-core/fetchngs](#), which itself, already, provides the option to download with SRA-tools. We picked these tools based on the interest of AncientMetagenomeDir contributors - of which SRAToolkit was not one at the time, however we have added this option in AMDirT v1.6 at the reviewers request.

- *If possible, I would like to see a slight reformat in the discussion "Problem/Solutions" section to make it more visible. A simple extra paragraph or bullet points that clearly separate problems and solutions should solve the problem.*

Response: We are unfortunately somewhat beholden to the formatting of the publisher during typesetting, but we have attempted to reformat the paragraph according to the reviewer's wish with paragraphs (as each text is too long for a bullet point)

Running and installing the pipeline.

- *I had a relatively smooth experience. However, I kept receiving the following warning upon*

any usage:

"WARNING streamlit.runtime.caching.cache_data_api: No runtime found, using MemoryCacheStorageManager"

This is unfortunately a [known issue](#) resulting from a design decision of one of AMDirT dependencies: the streamlit library and its caching system. As a stop gap solution, we now have 'monkeypatched' the streamlit function triggering this warning to silence it (<https://github.com/SPAAM-community/AMDirT/pull/132>).

- Additionally, the convert command did not work. After successfully installing the tool and downloading the AncientMetagenomeDir. I received the following:

```
$ AMDirT convert --curl SPAAM-community-AncientMetagenomeDir-60f8a00/ancientmetagenome-hostassociated/libraries/ancientmetagenome-hostassociated_libraries.tsv ancientmetagenome-hostassociated -o ./2023-10-17 19:50:32.289 No runtime found, using MemoryCacheStorageManager
```

Response: The convert command by default takes as input a *sample* table, not a library table, which is already reflected in the documentation of the convert command (amdirt.readthedocs.io/en/latest/how_to/convert) and the convert command CLI help.

...

```
$ AMDirT convert --help
```

```
Usage: AMDirT convert [OPTIONS] SAMPLES TABLE_NAME
```

```
Converts filtered samples and libraries tables to eager, ameta, taxprofiler, and fetchNGS input tables
```

```
Note: When supplying a pre-filtered libraries table with `--libraries`, the corresponding sample table is still required!
```

```
SAMPLES: path to filtered AncientMetagenomeDir samples tsv file
```

```
TABLE_NAME: name of table to convert
```

```
Options:
```

```
-t, --tables PATH (Optional) JSON file listing AncientMetagenomeDir tables
```

```
--libraries FILE (Optional) Path to a pre-filtered libraries table
```

```
NOTE: This argument is mutually exclusive with arguments: [librarymetadata].
```

```
--librarymetadata Generate AncientMetagenomeDir libraries table of all samples in input table NOTE: This argument is mutually exclusive with arguments: [libraries].
```

```
-o, --output DIRECTORY conversion output directory [default: .]
```

```
--bibliography Generate BibTeX file of all publications in input table
```

```
--curl Generate bash script with curl-based download commands for all libraries of samples in input table
```

```
--aspera Generate bash script with Aspera-based download commands for all libraries of samples in input table
```

```
--fetchngs Convert filtered samples and libraries tables to nf-core/fetchngs input tables
```

```
--sratoolkit    Generate bash script with SRA Toolkit fasterq-dump
                based download commands for all libraries of samples
                in input table
--eager        Convert filtered samples and libraries tables to
                eager input tables
--ameta        Convert filtered samples and libraries tables to
                aMeta input tables
--mag         Convert filtered samples and libraries tables to nf-
                core/mag input tables
--taxprofiler  Convert filtered samples and libraries tables to nf-
                core/taxprofiler input tables
--help        Show this message and exit.
...
```

Thus, for example, running the convert command with a sample table will generate this output:

```
...
$ AMDirT download -t ancientmetagenome-hostassociated -y samples -r v23.12.0
$ head -n 10 ancientmetagenome-hostassociated_samples_v23.12.0.tsv > test.tsv
$ AMDirT convert --curl test.tsv ancientmetagenome-hostassociated
AMDirT [WARNING]: We provide no warranty to the accuracy of the generated input sheets.
AMDirT [INFO]: Writing curl download script
$ head AncientMetagenomeDir_curl_download_script.sh
#!/usr/bin/env bash
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR132/022/SRR13263122/SRR13263122_2.fastq.gz -
o SRR13263122_2.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR132/024/SRR13263124/SRR13263124_2.fastq.gz -
o SRR13263124_2.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR132/031/SRR13263131/SRR13263131_2.fastq.gz -
o SRR13263131_2.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR132/030/SRR13263130/SRR13263130_2.fastq.gz -
o SRR13263130_2.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR957/SRR957740/SRR957740.fastq.gz -o
SRR957740.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR957/SRR957744/SRR957744.fastq.gz -o
SRR957744.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR957/SRR957741/SRR957741.fastq.gz -o
SRR957741.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR957/SRR957738/SRR957738.fastq.gz -o
SRR957738.fastq.gz
curl -L ftp://ftp.sra.ebi.ac.uk/vol1/fastq/SRR132/028/SRR13263128/SRR13263128_2.fastq.gz -
o SRR13263128_2.fastq.gz
...
```

However we've added an additional validation check to ensure that the input SAMPLE object is in the valid format expected by the tool, and if not asks the user to check if it is a sample

table (<https://github.com/SPAAM-community/AMDirT/pull/140>, commit 7bb6b55)
We have also added support for supplying *library* level metadata tables (supplied alongside a corresponding samples table) to the convert command with the --libraries parameter. This, alongside the new command download (added since the submission of the original manuscript), allows for an almost end to end command line based interaction with AMDirT: download → (filtering with bash tools) → convert, as reflected in the updated use case section of the manuscript.

Competing Interests: No competing interests were disclosed.

Reviewer Report 09 October 2023

<https://doi.org/10.5256/f1000research.147881.r210929>

© 2023 Read T et al. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Timothy Read

¹ Emory University, Atlanta, Georgia, USA

² Emory University, Atlanta, Georgia, USA

Robert Petit III

¹ Wyoming Public Health Laboratory, Cheyenne, Wyoming, USA

² Wyoming Public Health Laboratory, Cheyenne, Wyoming, USA

This paper describes the 'AMDirT' open source software for interaction with ancient metagenome sequence sample metadata tables. The tables have been produced through a massive communal curation effort by the international SPAAM (The Standards, Precautions, and Advances in Ancient Metagenomics community) group to systematically improve on what is available from the public repositories of archived short read sequence data. AMDirT allows viewing and searching of the tables using the Python streamlit library and facilitates download of data and integration with open source nextflow analysis pipelines. AMDirT also helps with initial creation and validation of new metadata tables.

I successfully installed AMDirT version 1.4.6 using conda on my Intel MacBook following instructions in the manuscript. I was able to replicate the commands described. There is also a public server <https://www.spaam-community.org/AMDirT/> for the AMDirT viewer. The documentation for the software is very good and includes video tutorials.

The manuscript is well-written and clearly outlines the functions of the software. The development of AMDirT represents a significant effort, not least because of the community-wide consultation. The need for community-based metadata addresses a well-known problem with the SRA/ENA/DDBJ databases and the discussion provides some nice examples of the type of issues faced when downloading. AMDirT is a valuable tool for both the ancient metagenomics research community and those outside the community interested in browsing and accessing the data.

Specific Points

- The streamlit-based viewer is pretty slick but I feel the importance of command line interface (CLI) is a little underplayed here. The viewer can be used by people without CLI proficiency but to actually take actions like making new sample tables, or download and process data, CLI is essential.
- The instructions in the text and on github for conda install should guide users to install into a fresh conda environment rather than into the base.
- There should be a list of computational environments that the software has been tested on (e.g do M1-3 macs work?)
- I did not see a mention in the text that there is actually a public facing server <https://www.spaam-community.org/AMDirT/>
- “Newly added library information columns include the library name (how data are typically reported in original publications), the aDNA library generation method (e.g., double-stranded or single-stranded libraries), the library indexing polymerase (e.g., proof-reading or non-proofreading), and the library pretreatment method (e.g., non-Uracil-DNA Glycosylase (UDG), full-UDG, or half-UDG treatments). The latter three fields represent information about the sequencing library construction that influence the presence of aDNA damage, a factor that is critical for the processing of aDNA NGS data.^{8,18} Sequencing metadata columns include instrument model, library layout (single- or paired-end), library strategy (whole genome sequencing, targeted capture, etc.), and read count. “I could not work out how to search these fields through the viewer. These are accessible as downloads post validation but it seems that users would want to search through samples based on these fields?

Further questions

Finally, I have some three questions about the project that it would be great to get comments on.

1. How sustainable is the community effort to maintain these databases moving forward into the future with the probability of the number of samples increasing each year?
2. Have you tried sending the improved metadata back to SRA/ENA?
3. How difficult would it be to adapt the software for a different community of researchers that wanted to improve annotation but use fields that would be different from the ancient metagenomes community?

Is the rationale for developing the new software tool clearly explained?

Yes

Is the description of the software tool technically sound?

Yes

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?

Yes

Is sufficient information provided to allow interpretation of the expected output datasets and any results generated using the tool?

Yes

Are the conclusions about the tool and its performance adequately supported by the findings presented in the article?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Bacterial genomics, metagenomics, bacterial genetics, antibiotic resistance

We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 06 May 2024

James Fellows Yates

Thank you for your positive review. We apologies for the long time between your review and our revision, this was due to parental leave and extensive traveling of the corresponding authors.

Our responses to your specific points and further questions are as follows. Line numbers should correspond to the submitted revised word document (required by F1000), once track changes are accepted. Please note depending on which software we opened it, the line numbers were not indicating the same content. The line numbers should correspond to what is opened in Microsoft Office for Mac Word v16.78.3.

Specific Points

- *The streamlit-based viewer is pretty slick but I feel the importance of command line interface (CLI) is a little underplayed here. The viewer can be used by people without CLI proficiency but to actually take actions like making new sample tables, or download and process data, CLI is essential.*

Response: We have made more prominent references to the command line interface functionalities in the manuscript by further specifying in the introduction on lines 75-88, 101-140, and 151-155.

We also extended the 'Use case' section of the manuscript to further guide readers on how they could use AMDirT command line tools to perform the same workflow in the GUI as shown in Figure 3. Note that during the improvement of this use case, we also added an additional command called 'download' to further make it easier for CLI power-users to retrieve the tables for a more complete CLI based workflow.

- *The instructions in the text and on github for conda install should guide users to install into a fresh conda environment rather than into the base.*

Response: We have updated the documentation to reflect the new instructions in the code

repository README, documentation pages (both can be seen in

<https://amdir.readthedocs.io/en/master/README.html>), and the manuscript line 263

- *There should be a list of computational environments that the software has been tested on (e.g do M1-3 macs work?)*

Response: We have added a brief section about hardware and OSs that have been tested in the github repository README installation page of the documentation (<https://amdir.readthedocs.io/en/master/README.html>).

- *I did not see a mention in the text that there is actually a public facing server*
<https://www.spaam-community.org/AMDirT/>

Response: There was already a mention of the viewer interface at the line 369, but we have added more details lines 76, and 759

- *"Newly added library information columns include the library name (how data are typically reported in original publications), the aDNA library generation method (e.g., double-stranded or single-stranded libraries), the library indexing polymerase (e.g., proof-reading or non-proofreading), and the library pretreatment method (e.g., non-Uracil-DNA Glycosylase (UDG), full-UDG, or half-UDG treatments). The latter three fields represent information about the sequencing library construction that influence the presence of aDNA damage, a factor that is critical for the processing of aDNA NGS data.^{8,18} Sequencing metadata columns include instrument model, library layout (single- or paired-end), library strategy (whole genome sequencing, targeted capture, etc.), and read count. "I could not work out how to search these fields through the viewer. These are accessible as downloads post validation but it seems that users would want to search through samples based on these fields?"*

Response: We have now extended the AMDirT viewer and convert commands to support library level filtering. In the AMDirT viewer this is supported via a secondary table that is loaded after the sample selection has been made by the user. We've updated the use case example and Figure 2 to represent these changes.

Further questions

Finally, I have some three questions about the project that it would be great to get comments on.

- *How sustainable is the community effort to maintain these databases moving forward into the future with the probability of the number of samples increasing each year?*

Response: Ancient metagenomics as a field is still very young and therefore has indeed been very manageable in terms of numbers. But of course as it grows it could become more difficult.

In passive terms - our feeling so far is as we have started as the field is 'young' has benefited from the fact we will have good visibility and awareness of the resource within the community, something that would continue to spread as the research area expands through knowledge transfer. With this, we believe that we will have both a) more potential and willing contributors to cope with the increased number of samples and b) as they are already aware, will maybe already format and prepare their own sample and library metadata in a form easy to add to AncientMetagenomeDir.

More proactively, we will continue to survey the community to find out what are good motivating factors to continue contributing. We further plan to continue to refine the tooling and automation to make it as easy as possible for contributors to add and review

new contributors. The SPAAM community has also recently become affiliated with the ISBA society (<https://isbarch.org>), which can provide financial support for the previously 'volunteer' held hackathons. We also have various ideas for new publications that are typically the biggest motivating factor for researchers (e.g. extending the database to include more precise radiocarbon dates which are critical e.g. for phylogenetic dating analyses). Finally we have also expanded our aim of better metadata reporting of ancient DNA samples and sequencing data to a larger project covering the whole of palaeogenomics (<https://www.mixs-minas.org/>) that will also help further formalise such thinking in the community, and feed-back into the way AncientMetagenomeDir functions, as well as push such standardisation into the INDSC consortium databases (who use MIXS checklists themselves).

- *Have you tried sending the improved metadata back to SRA/ENA?*

Response: We have not (yet) attempted this. We have not actively recorded all the corrections between the reported information in the publications and as changed for the AncientMetagenomeDir repository. These corrections have been also derived from a mixture of 'obvious' mistakes in the publication (e.g. single-end sequencing reported, but paired FASTQ files uploaded to the ENA or SRA), to personal communications with the authors of the publication. In some cases the original authors have themselves made the corrections on ENA/SRA themselves. In any case, requesting changes on SRA/ENA would require permission from the original authors of the publications, which would require a large and extensive effort from the volunteers of SPAAM, with likely highly mixed chances of success (students move on, labs have changed/retired etc). In this vein we feel it would be more beneficial to invest time and effort in the MInAS project to correct future submissions at the source rather than historical data.

- *How difficult would it be to adapt the software for a different community of researchers that wanted to improve annotation but use fields that would be different from the ancient metagenomes community?*

Response: AMDirT is mostly based around the JSON schemas that define and enforce the structure of the underlying data. Furthermore, because the validate subpackage is designed using a domain-driven design software architecture, the base class holding the different methods responsible for testing the data are generic and can be applied to any dataframe object, as long as they come with their associated JSON schema. In summary, adapting it to a different community would require some modifications (as some part of the code is specific to ancient metagenomics data), but most of AMDirT core functionalities could be adapted without too much effort.

Competing Interests: We declare no competing interests.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research