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Total syntheses of natural products containing spirocarbocycles

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The structures of natural products from a variety of sources contain spirocycles, two rings that share a common atom. The spiro motif is finding increasing inclusion in drug candidates, and as a structural component in several promising classes of chiral ligands used in asymmetric synthesis. Total syntheses of products containing all-carbon spirocycles feature several common methods of ring closure which we examine in this review.

1. Introduction

Natural products have been an invaluable source of medicines and other resources throughout history. In recent years, the search for novel compounds has moved from the land to the sea, concentrating on marine organisms rather than terrestrial species. There are several ways of classifying such compounds: by genus of their source, by physiological effect, or by a common structural element such as their carbon skeleton. This review will focus on natural products containing the spiro

motif. Spiro compounds possess two or more rings which share only one common atom, with the rings not linked by a bridge.¹

Spiro-containing compounds have been isolated from a wide range of biological sources, from plants to frogs to marine sponges. The shift from earth to water in search of compounds is mirrored here. Selected examples of such compounds are shown below (Fig. 1). Spirocycles containing heteroatoms, such as the spirooxindoles, are well-known in nature, and there have been several reviews on these species.^{2,3} One of the earliest isolated spiro natural products was β -vetivone, extracted from vetiver oil in 1939 by Pfau and Plattner.⁴ However, for many years its structure was believed to be hydroazulenic (**1.6**) rather than a spiro[4.5]decane (**1.7**), an error

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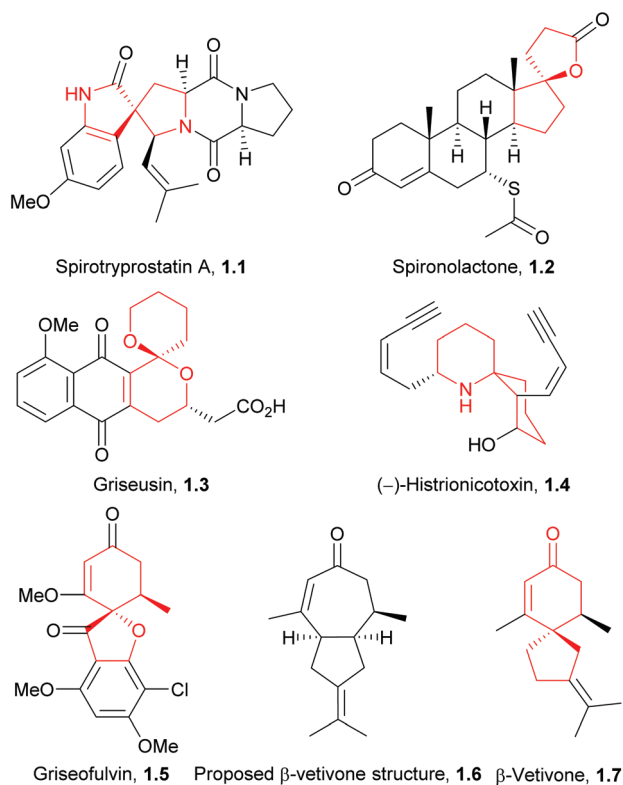


Fig. 1 Natural products and drugs containing spirocycles.

which was only rectified by its total synthesis by Marshall *et al.* in 1968.^{5,6}

The spiro motif is becoming more prevalent as a template in drug discovery, and has been the subject of a recent review.⁷ Spiro rings of different sizes convey both increased three-dimensionality for potential improved activity, and novelty for patenting purposes. Two examples of marketed drugs containing spirocycles are spironolactone (**1.2**) and griseofulvin (**1.5**, Fig. 1). Both are on the World Health Organisation's list of essential medicines; spironolactone is a diuretic and anti-hypertensive drug, whilst griseofulvin is an anti-fungal agent.

An additional important application of spiro compounds is in asymmetric synthesis. Enantioselectivity in organic reactions can be achieved through the use of chiral ligands, such as BINOL (**1.8**) or cinchona alkaloids (**1.9**, Fig. 2). Some spiro-

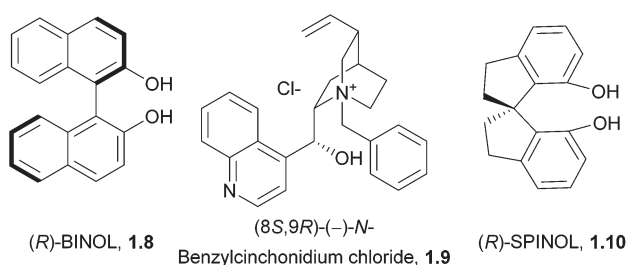


Fig. 2 Selected privileged ligand classes.

cycles, such as the spirobiindane SPINOL (**1.10**), belong to a class of privileged chiral ligands which show particular promise.⁸ The increased rigidity and restricted rotation at the spiro centre can assist in improving enantiomeric excess in asymmetric reactions.

The asymmetric synthesis of quaternary carbon centres is extremely challenging,⁹ with the stereocontrolled preparation of products containing such spiroatoms^{10–12} conveying additional difficulty. Enantioselective syntheses of spirocyclic ligands utilise several construction methods, such as alkylation, radical cyclisation, rearrangements, cycloaddition reactions, cleavage of bridged ring systems and the use of transition metal catalysts.¹³ Several ring-closing strategies are common in the synthesis of spirocarbocycle-containing natural products, and these will be discussed in this review. Examination of the syntheses of these compounds will be first by reaction type of the spirocycle-forming step, followed by increasing ring size, from [2.5] through to [5.5] (Table 1). Not all ring sizes are represented in the natural world, nor have all spiro-containing natural products been the target of organic synthesis. Some reaction types have not been included in this article, for example the Wittig reaction.

Some of the earliest-discovered and best-studied compounds such as the aforementioned β -vetivone have been synthesised by several of these methods. The field has matured significantly since the 1970s, when many of the structures of these spiro compounds were elucidated. Syntheses of these compounds span the last fifty years, and so not all of the original articles were published in English, but where relevant, have been translated and summarised here. In general, newer spirocyclisation methods tend to be both more efficient and are conducted enantioselectively. For example, Diels–Alder spirocyclisations of the acoranes were first attempted in 1975 and re-examined in 2013, where one isomer was selectively synthesised rather than the full range of four isomers (Scheme 1).

Reports from the last five years indicate that new spiro natural products are still being isolated and synthesised. Total syntheses of some compounds isolated many years ago were achieved using the less sophisticated methodologies available at the time, but there has been much improvement of these older methods by the application of more modern processing techniques and understanding. It may also be beneficial to apply some of the approaches recently developed in the enantioselective synthesis of chiral ligands to the synthesis of natural products. Several new natural product compounds have been recently identified that would benefit from an efficient preparative synthesis. Cyanthiwigin AC (Fig. 3), isolated from the marine sponge *Myrmekioderma styx*,¹⁴ is such an example. The cyanthiwigins exhibit a broad range of biological activity, including inhibition of human immunodeficiency virus and *Mycobacterium tuberculosis*, and cytotoxicity against human primary tumour cells.¹⁵ The first and only total synthesis of (+)-cyanthiwigin AC was achieved in 13 steps and 2% overall yield.¹⁶ Other compounds have also shown interesting preliminary biological activity ranging from anti-bacterial to anti-fouling activity but further investigation is hampered



Table 1 List of the natural products covered in this review, in order of increasing ring size then alphabetically

Natural product	Structure	Spiro	Section	Natural product	Structure	Spiro	Section
Illudin M		[2.5]	4	Anhydro- β -rotunol (2005)		[4.5]	2
Illudin S		[2.5]	4	Axenol (2002, 2007)		[4.5]	4, 6
(-)-Acutumine (synthesised in 2009)		[4.4]	7	Axisonitrile-3 (2009)		[4.5]	6
Ainsliadimer A (2010)		[4.4]	5	Bilinderone (2011)		[4.5]	7
Vannusal A (R = Ac) Vannusal B (R = H) (2009)		[4.4]	2	Cannabispirenone A: C=C, R ₁ = Me, R ₂ = H; B: C=C, R ₁ = H, R ₂ = Me Cannabispirone (C-C, R ₁ = Me, R ₂ = H) (1981, 1982, 1984)		[4.5]	2
Acorenone (1978, 1982, 2011)		[4.5]	3, 4, 7	Colletoic acid (2013)		[4.5]	5
Acorenone B (1975, 1978)		[4.5]	2, 3	Erythrodiene (1997, 1998)		[4.5]	2, 6
Acorone (1975, 1977, 1978, 1996, 2007)		[4.5]	2, 4, 5, 6	Gleenol (2002, 2007, 2009)		[4.5]	4, 6
Agarospinol (1970, 1975, 1980, 1995, 2006)		[4.5]	2, 3, 6, 7	Gochnatilide A (R = α -OH) Gochnatilide B (R = β -OH) Gochnatilide C (R = α -H) (2013)		[4.5]	5
Ainsliadimer B (2013)		[4.5]	5	Hinesene (2R,5S,10S) (2004)		[4.5]	3

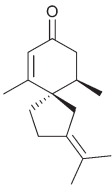
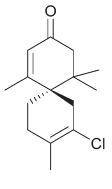
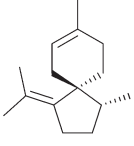
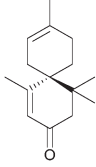
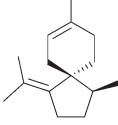
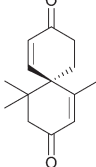
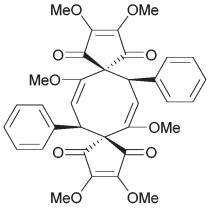
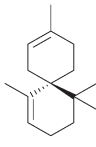
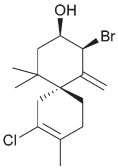
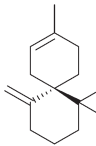


Table 1 (Contd.)

Natural product	Structure	Spiro	Section	Natural product	Structure	Spiro	Section
Hinesol (1970, 1973, 1980, 1995, 2004, 2006, 2007)		[4.5]	3, 6, 7	Spirojatamol (1997, 1998)		[4.5]	2, 6
Isoacorone (1975, 2000, 2007, 2008)		[4.5]	2, 4, 5, 6	Spirolaurenone (1982)		[4.5]	3
Lubimin (1984)		[4.5]	3	Trichoacorenol (2011)		[4.5]	4
Lubiminol (1996, 1998)		[4.5]	7	(+)-Vitrenal (1982, 1986)		[4.5]	2, 3
Oxylubimin (1984)		[4.5]	3	α -Acoradiene: L (1 <i>R</i> ,4 <i>S</i> ,5 <i>S</i>) R (1 <i>S</i> ,4 <i>R</i> ,5 <i>R</i>) (1983, 2001, 2004)		[4.5]	2, 4, 7
Premnaspirodiene (2 <i>R</i> ,5 <i>S</i> ,10 <i>R</i>) (2004)		[4.5]	3	α -Acorenol (2007)		[4.5]	4, 6
Proaporphines: Pronuciferine (<i>R</i> ₁ = Me, <i>R</i> ₂ = H) Orientalinone (<i>R</i> ₁ = Me, <i>R</i> ₂ = OMe) Kreysiginone (<i>R</i> ₁ = H, <i>R</i> ₂ = OMe) (1971, 1972, 1978) Sequosempervirin A (2007)		[4.5]	7	α -Vetispirene (1973, 1976, 1985, 2004, 2006, 2007)		[4.5]	2, 3, 6, 7
		[4.5]	4	β -Acoradiene (1975)		[4.5]	5
Solavetivone (1984)		[4.5]	3	β -Acorenol (2007)		[4.5]	4, 6
Spirocurcasone (2013, 2014)		[4.5]	3, 4	β -Vetispirene (1973)		[4.5]	3



Table 1 (Contd.)

Natural product	Structure	Spiro	Section	Natural product	Structure	Spiro	Section
β -Vetivone (1968, 1973, 1984, 2006, 2007)		[4.5]	2, 3, 6, 7	Laurencenone B (2008)		[5.5]	4
γ -Acoradiene (1973)		[4.5]	5	Laurencenone C (2010)		[5.5]	2, 4
δ -Acoradiene (1973)		[4.5]	5	Majusculone (2007, 2011)		[5.5]	2, 3
Linderaspirone A (2011)		[4.3.4.3]	7	α -Chamigrene (1991, 2006)		[5.5]	3, 4
Elatol (2008)		[5.5]	4	β -Chamigrene (2006, 2008)		[5.5]	4

by the inability to isolate or synthesise sufficient quantities. The ritterazines are such a class of compounds: they are a family of 26 steroidal alkaloids of which 11 contain a spirocarbocycle in addition to the spiroketal shared by all ritterazines.¹⁷ They possess potent cytotoxic activity against apoptosis-resistant malignant cell lines but research has been limited by the small amounts of material successfully isolated from naturally occurring sources, so efforts have focussed on their total synthesis. There have been several published syntheses of the other 15 ritterazines and derivatives, but at the time of writing there has not yet been a published total synthesis of the spirocarbocycle-containing ritterazines.

Taber *et al.* have come the closest to synthesising ritterazine N (**1.20**, Scheme 2) and related analogues^{18,19} by assembling a ritterazine N precursor. The final planned aldol cyclocondensation failed, but an alternative route *via* ozonolysis of an alkene followed by treatment with base successfully gave the related compound bis-18,18'-desmethylritterazine N (**1.23**).^{19,20}

As this review aims to summarise the progress in the field, but also to identify areas for improvement, where natural pro-

ducts have been synthesised by more than one strategy, the methods will be compared.

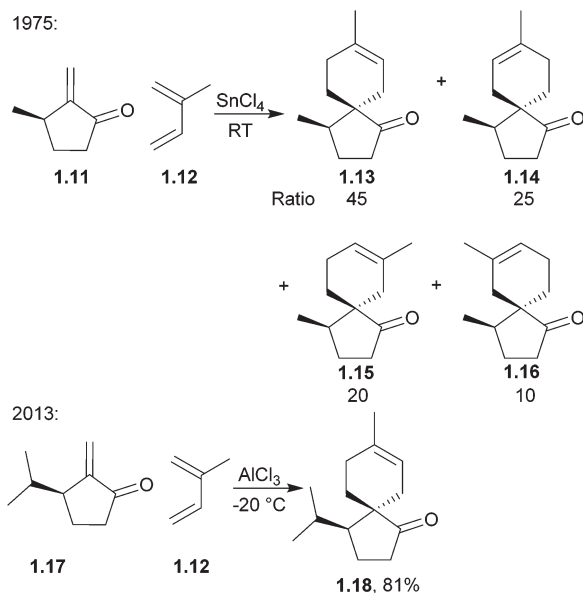
2. Aldol reaction

The aldol reaction is a powerful carbon-carbon bond forming reaction.²¹ Stereochemical control is possible through the use of chiral aldehyde starting materials, or chiral auxiliaries. The asymmetric reaction can also be performed catalytically through the use of chiral Lewis acids.²² This approach has been particularly effective in the synthesis of various natural product spirocarbocycles as illustrated below.

2.1 Vannusals A and B

Vannusals A and B are triterpenes with an unusual C₃₀ backbone, comprising seven rings and thirteen chiral centres, three of which are quaternary. They were isolated from the tropical zooplankton *Euplotes vannus*, and their biosynthesis is thought





Scheme 1 Comparing the synthesis of acoranes by the Diels–Alder reaction.

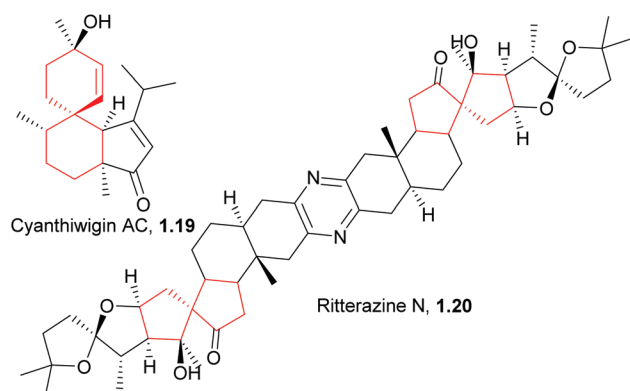


Fig. 3 Structures of cyanthiwigin AC and ritterazine N.

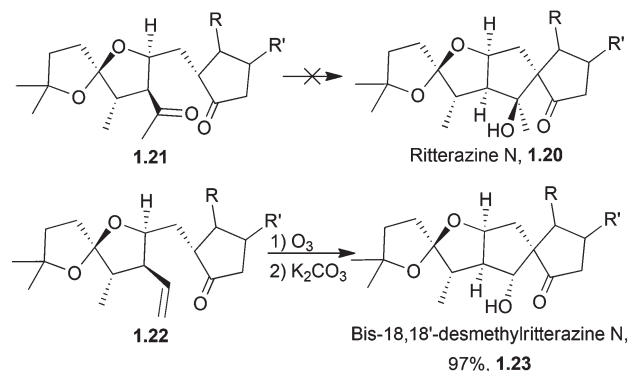
to proceed from a squalene-type precursor (Fig. 4), *via* several possible routes.²³

The originally assigned structures for the vannusals were incorrect, and the true structures were only established following the total synthesis efforts of the Nicolaou group.²⁴ The fused polycyclic carbon framework was assembled by an intramolecular spirocyclisation *via* a Mukaiyama-type aldol reaction^{25,26} (Scheme 3).

The above methodology was used in the total syntheses of the original incorrect vannusal B structure,²⁷ as well as the resyntheses of the true vannusals A and B.^{24,28–30}

2.2 Acoranes: α -acoradiene, acorone, isoacorone and acorenone B

The acoranes are a group of highly-oxygenated sesquiterpenes containing a spiro[4.5] core. They have been isolated from a variety of sources,^{31–33} including sweet flag *Acorus calamus*, the



Scheme 2 Attempted syntheses of ritterazine N. Bis-18,18'-desmethyrritterazine N was obtained as a single diastereomer in high yield.

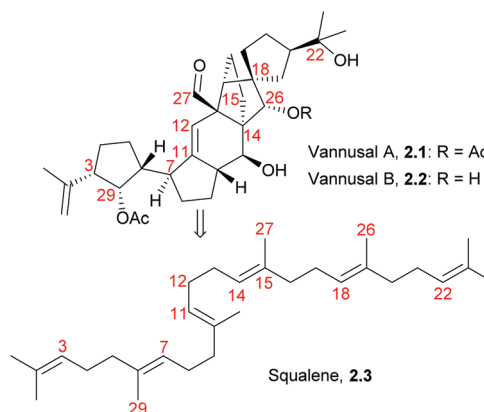
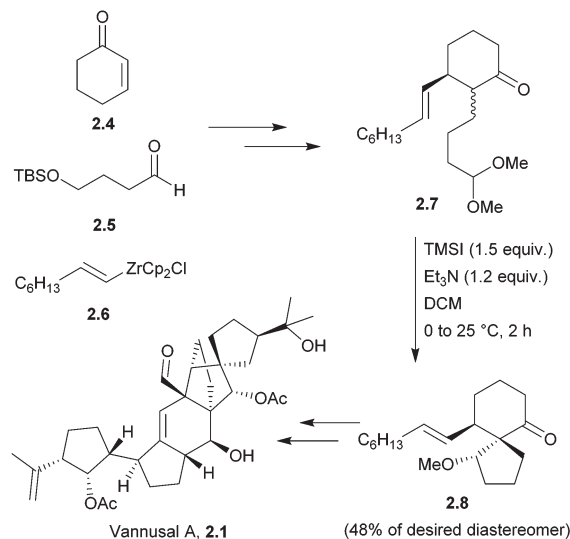
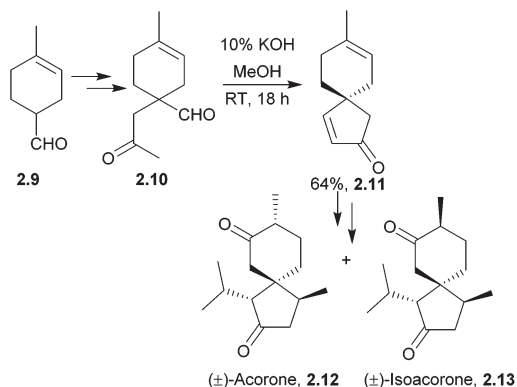


Fig. 4 Proposed link between the structures of vannusals and squalene.

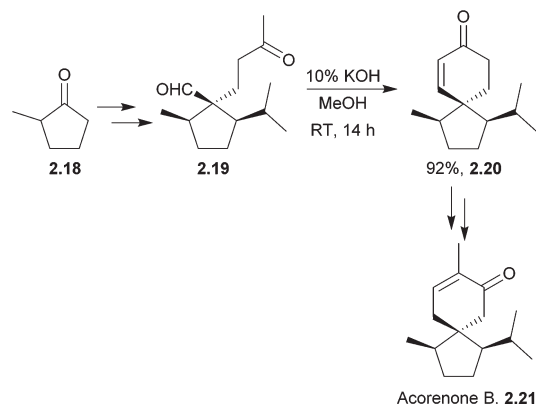


Scheme 3 Mukaiyama aldol reaction in the synthesis of vannusal A.





Scheme 4 Base-catalysed cyclisation in the synthesis of (±)-acorone and (±)-isoacorone. No mention was made of the dr of intermediate 2.11.



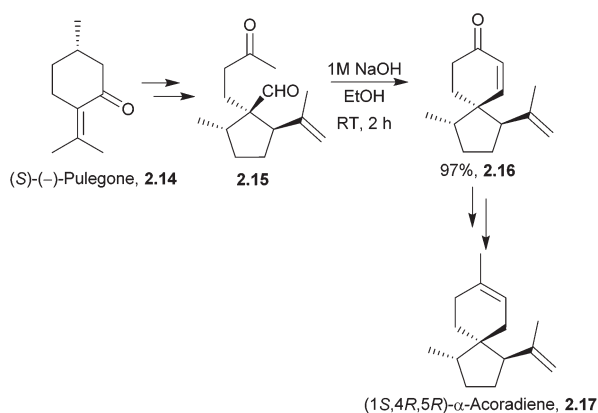
Scheme 6 Aldol reaction in the synthesis of acorenone B.

essential oils of the woods of *Juniperus rigida* and *J. chinensis*, and Australian sandalwood oil from *Santalum spicatum*.

(±)-Acorone was synthesised in 1977 by the Dolby group, starting from 4-methyl-3-cyclohexene carboxaldehyde and using an intramolecular base-catalysed aldol cyclisation³⁴ (NaOEt/EtOH). A similar method reported by Martin *et al.* in 1978 used an aldol cyclocondensation to also synthesise racemic acorone³⁵ (Scheme 4).

In addition, in 1996 Srikrishna *et al.* performed a formal total synthesis of (±)-acorone³⁶ using an intramolecular aldol condensation of 1,4-cyclohexanedione. (±)-Acorone and (±)-isoacorone were later formally synthesised by the same group using a Claisen rearrangement-based methodology.³⁷

Two enantiomers of the sesquiterpenoid α -acoradiene occur naturally: one is found in juniper wood, and the other is a pheromone of the broad-horned flour beetle. The structure of the beetle α -acoradiene was originally thought to be the (1*R*,4*R*,5*S*) enantiomer, which has been synthesised by ring-closing metathesis (see below). (1*S*,4*R*,5*R*)- α -Acoradiene was synthesised from (*S*)-(-)-pulegone in 10 steps and 16% overall yield³⁸ (Scheme 5) by Mori *et al.* in 2004. The key step was the



Scheme 5 Aldol condensation of 1,5-diketone 2.15 to form the spiro intermediate 2.16 in the synthesis of α -acoradiene.

aldol condensation of a 1,5-diketone to form the 6-membered ring.

Acorenone B has been synthesised under basic conditions to form the spirocycle³⁹ in high yield (Scheme 6) by Trost *et al.* in 1975.

2.3 Spirovetivanes: agarospirol, β -vetivone, α -vetispirene and anhydro- β -rotunol

The spirovetivanes are sesquiterpenes with a spiro[4.5] core. They include metabolites and phytoalexins, compounds produced when a species is subjected to stress such as pathogenic attack.⁴⁰ Vetiver oil from *Vetiveria zizanioides* contains several spirovetivanes, including β -vetivone,⁴ α -vetispirene and β -vetispirene.⁴¹ As stated above, for many years, the structure of β -vetivone was thought to be hydroazulenenic but this was revised to the spiro[4.5]decane in 1968.^{5,6}

Following on from this work by Marshall and Johnson, Wenkert *et al.* used methanolic KOH to effect a basic aldol cyclisation of a key intermediate in the synthesis of β -vetivone.⁴² The reaction proceeded in 95% yield, as part of a formal total synthesis requiring 8 steps to reach the preliminary compound.

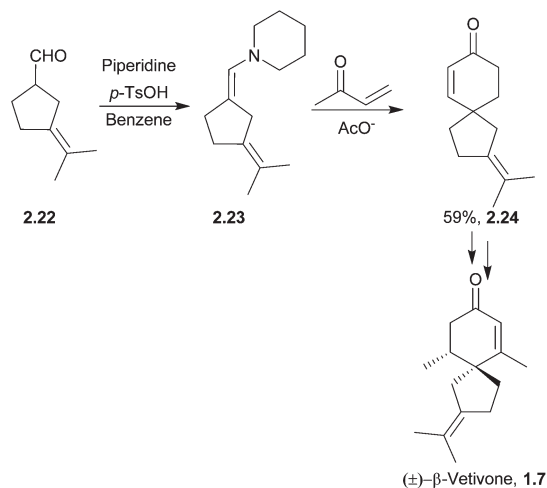
A tandem Michael-aldol condensation was used by Hutchins *et al.* in 1984 to synthesise racemic β -vetivone⁴³ (Scheme 7) in a very short and efficient synthesis.

Starting from a spiro[5.5]acetal, (-)-agarospirol, a metabolite isolated from agarwood oil,⁴⁴ was synthesised in 1975 *via* a 1,6-dicarbonyl and base to generate the α,β -unsaturated ketone.⁴⁵

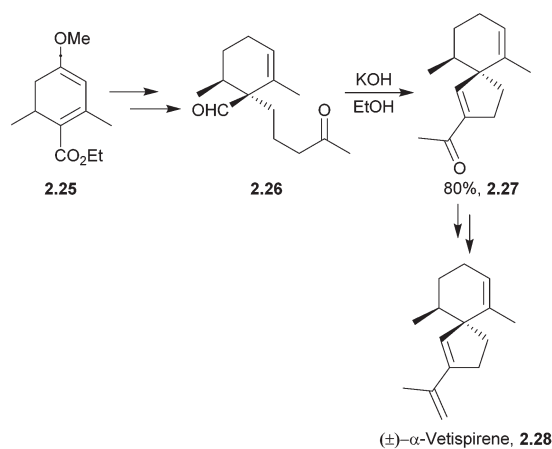
Ten years later, α -vetispirene was synthesised by Balme using a related intramolecular spirocondensation with a 1,5-dicarbonyl under basic conditions⁴⁶ (Scheme 8).

Anhydro- β -rotunol is a phytoalexin spirovetivadiene isolated from infected potato tubers.⁴⁷ In 2005 Srikrishna *et al.* performed an enantioselective synthesis of (+)-anhydro- β -rotunol starting from (*R*)-carvone in 13 steps and 15% overall yield⁴⁸ (Scheme 9). The spirocyclisation step involved a regioselective intramolecular aldol condensation of a keto-aldehyde, reminiscent of Balme's synthesis of α -vetispirene. Other conditions were attempted, but piperidine in AcOH was found to be the most successful.

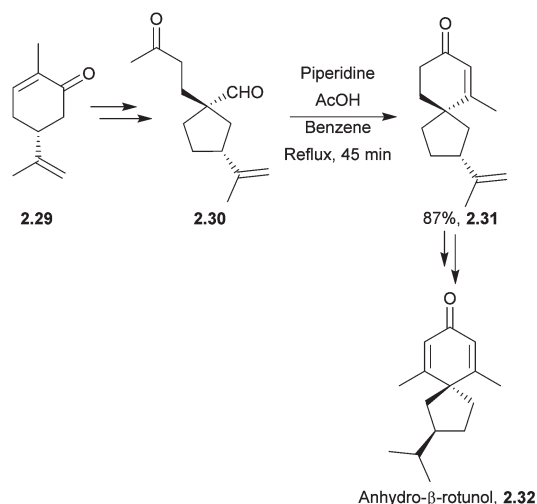




Scheme 7 Tandem Michael-aldol condensation in the synthesis of β -vetivone.



Scheme 8 Basic aldol condensation in the synthesis of α -vetispiene.



Scheme 9 Aldol condensation in the synthesis of anhydro- β -rotunol.

2.4 Erythrodiene and spirojatamol

Erythrodiene and spirojatamol are sesquiterpenes that share a common spirobicyclic [4.5]decane skeleton. (–)-Erythrodiene was isolated from a Caribbean encrusting coral, whereas (+)-spirojatamol was isolated from a Himalayan flowering plant. Erythrodiene and spirojatamol have been formally synthesised from *N*-cyclohexyl-4-isopropylcyclohexanimine⁴⁹ by Ihara *et al.* in 1997. The key spirocyclisation step was a tandem intramolecular Michael-aldol reaction of a ketoacetal with $\text{Me}_3\text{SiI}/(\text{Me}_3\text{Si})_2\text{NH}$ (Scheme 10).

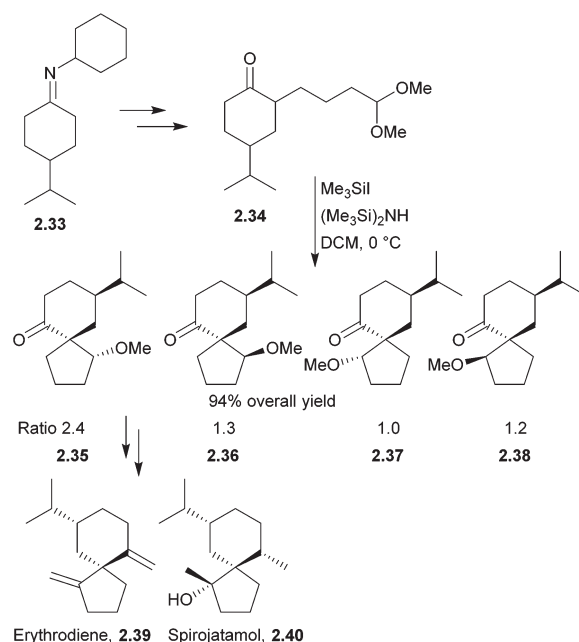
Forsyth *et al.* have used spirocarbomercuration to synthesise erythrodiene and spirojatamol;^{50,51} more recently, Oppolzer *et al.* have used a Pd-catalysed allylzincation⁵² and Renaud *et al.* a thiophenol-mediated spirocyclisation.⁵³

2.5 Vitrenal

(+)-Vitrenal is a sesquiterpenoid and a potent plant growth inhibitor.⁵⁴ The unnatural enantiomer, (–)-vitrenal, has been synthesised from (+)-3-carene⁵⁵ by Ito *et al.* in 1986. The key sequence is a one-pot deacetalisation-aldol condensation, which is preceded by a stereoselective Claisen rearrangement (Scheme 11). It showed weaker plant growth inhibition than the natural enantiomer.

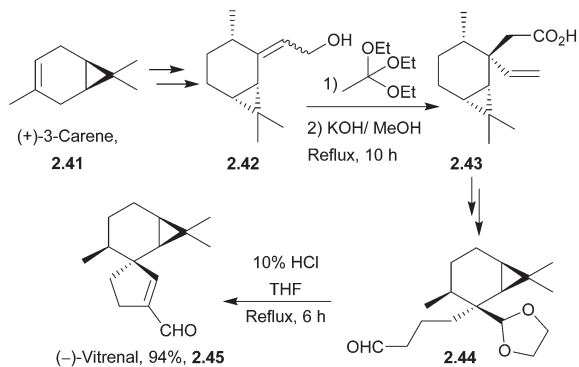
2.6 Cannabispirans: cannabispirenes A and B and cannabispirene

A variety of spiroindane products have been isolated from the marijuana plant, *Cannabis sativa*.^{56–62} They are structurally related to some synthetic oestrogenic-potentiating agents and

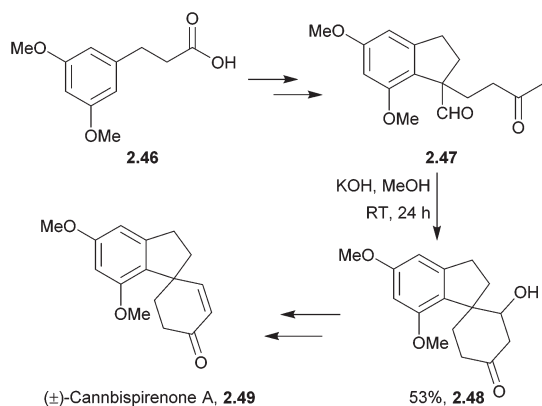


Scheme 10 Aldol reaction during the synthesis of erythrodiene and spirojatamol. The diastereomers 2.35–2.38 were separable by column chromatography.





Scheme 11 One-pot deacetalisation-Aldol condensation in the synthesis of (–)-vitrenal. Intermediate **2.42** was present as a 1 : 1 mixture of geometrical isomers, which were only separable by gas chromatography. Both isomers gave the same acid (**2.43**).



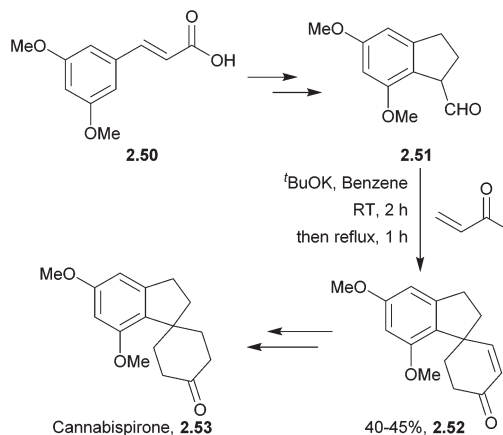
Scheme 12 Basic aldol reaction in the spirocyclisation of cannabispirenone A.

so may show similar oestrogenic activity to marijuana.⁵⁹ Crombie *et al.* used KOH to intramolecularly close the spiro ring of several cannabispirens^{63,64} (Scheme 12).

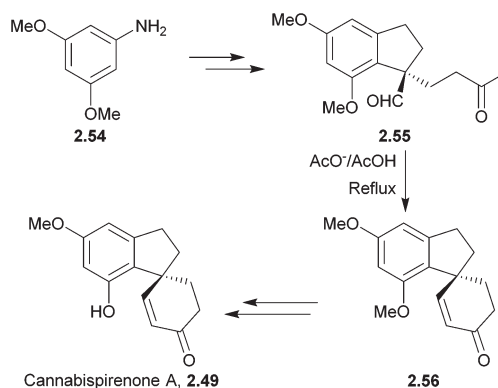
Cannabispirenone was synthesised by treating an aldehyde and methyl vinyl ketone with ^tBuOK⁶⁵ (Scheme 13) by El-Feraly *et al.* in 1981.

Cannabispirenone A was asymmetrically synthesised by Natale *et al.* in 1984 from 3,5-dimethoxyaniline in 11 steps, using an enamine and methyl vinyl ketone followed by hydrolysis to form the spiro centre⁶⁶ (Scheme 14).

Similarly, cannabispirenone B was synthesised by Novak *et al.* in 1982.⁶⁷ Aldehyde **2.51** was transformed to the piperidine enamine, before it was reacted with methyl vinyl ketone followed by hydrolysis and aldol cyclisation. The increased difficulty in the synthesis of cannabispirenone B over A lay in the selective ether cleavage of the less sterically hindered aryl methoxy group. Previously this had only been possible using BBr₃, which gave low yields and led to the formation of insoluble tars. Ten reagents were considered by the authors, of which only LiI in 2,4,6-trimethylpyridine was successful in 56% conversion and 82% yield.



Scheme 13 Spirocyclisation step in the synthesis of cannabispirenone.



Scheme 14 Aldol condensation in the synthesis of cannabispirenone A.

2.7 Chamigrenes: majusculone and laurencenone C

The chamigrene family is a group of over 100 sesquiterpene natural products that contain a spiro[5.5]undecane core. Examples include majusculone and laurencenone C.^{68,69} Some chamigrenes are halogenated, and the group shows diverse biological activity.

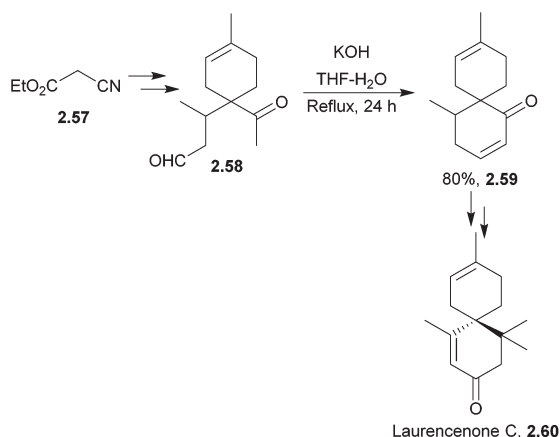
Zhu *et al.* synthesised racemic laurencenone C in 2010, and the key step was a base-catalysed aldol condensation⁷⁰ (Scheme 15). The full sequence involved 11 steps with an overall yield of 17%.

Racemic majusculone also was synthesised from ethyl cyanoacetate in 15 steps⁷¹ (Scheme 16) by Zhu *et al.* in 2011. The key step in this particular synthesis was the tandem oxidation-aldol condensation of the diol to the keto aldehyde which underwent base-mediated cyclisation.

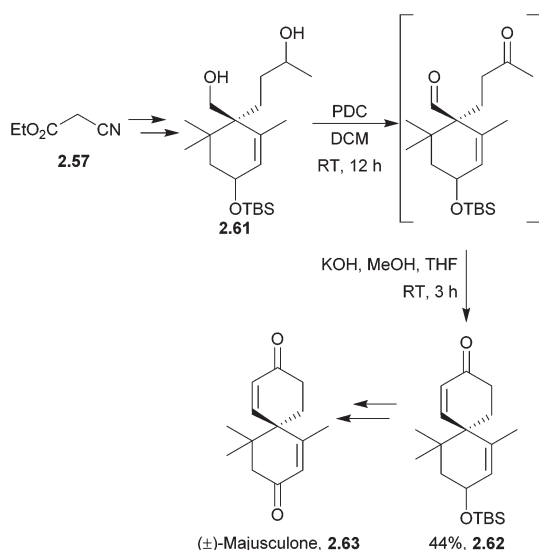
3. Acid and base promoted spirocyclisation

Acids or bases can be used in a wider range of cyclisation reactions than just to promote the aldol reaction. For example, the





Scheme 15 Aldol cyclisation in the synthesis of laurencenone C.



Scheme 16 Oxidation followed by cyclisation in the synthesis of (±)-majusculone.

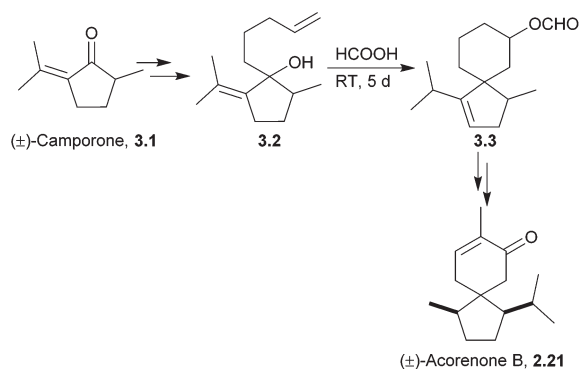
related Robinson annulation and Prins cyclisation have been used to synthesise natural products containing spirocarbocycles as described below.

3.1 Acoranes: acorenone, acorenone B and α -acorenone

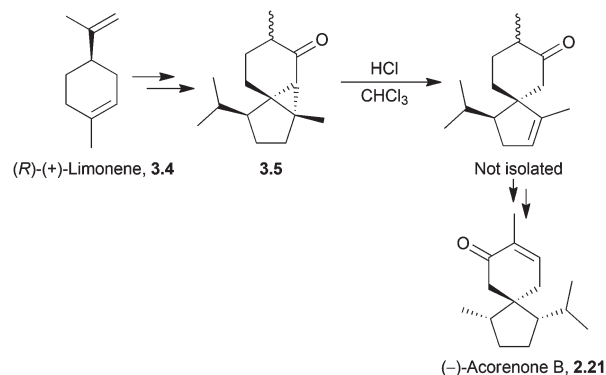
(±)-Acorenone B has been synthesised from (±)-camphorone.⁷² Formic acid was used to activate the allylic alcohol to attack by an alkene *via* an allylic cation to form the spiro centre (Scheme 17).

No mention was made of stereochemistry or diastereoselectivity in the original 1975 paper and thus this method may be less effective than the cyclisation in base which was described above.

In 1976, White *et al.* started from (*R*)-(+)-limonene to synthesise (–)-acorenone B in 11 steps.⁷³ The key step was an acid-



Scheme 17 Treatment of an intermediate with formic acid in the synthesis of acorenone B. The dr were not stated, but the product was isolated as a single diastereomer *via* preparative TLC.



Scheme 18 Ring-opening of an intermediate 3.5 in the synthesis of (–)-acorenone B.

catalysed ring-opening of a cyclopropane to reveal the spiro-[4.5]decanone (Scheme 18).

If lithium in liquid ammonia was used to perform a reductive scission of the cyclopropane bond, inversion of the stereochemistry of the methyl group was observed.

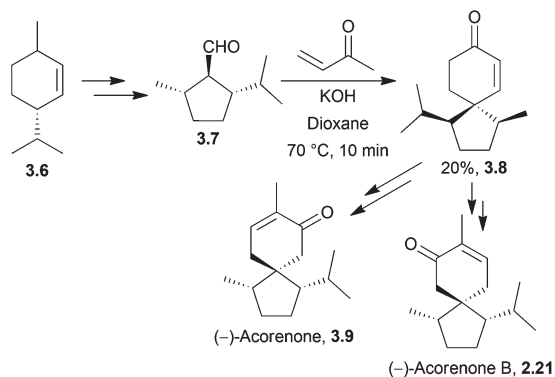
Pesaro *et al.* used a Robinson annulation in 1978 to synthesise both (–)-acorenone and (–)-acorenone B, in 4 steps starting from (+)-*p*-menth-1-ene⁷⁴ (Scheme 19).

The reaction gave the key spiro intermediate in low yield, with the stereoselectivity of the reaction controlled by the isopropyl and methyl groups, which direct the electrophilic addition of the methyl vinyl ketone to the less hindered face.

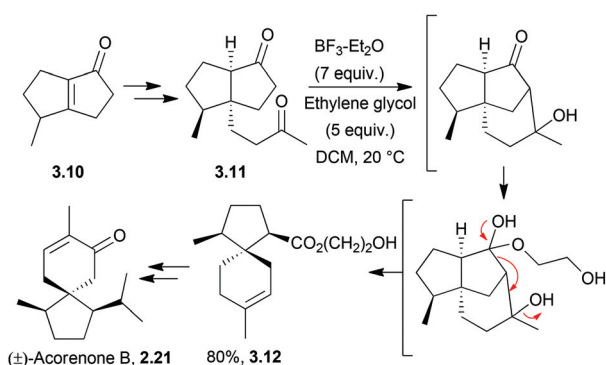
More recently, Sakai *et al.* used a Lewis acid to effect a one-pot transformation to a spiro compound en route to racemic acorenone B (Scheme 20).⁷⁵ The mechanism is thought to proceed *via* an aldol condensation, followed by hemiacetalisation and finally a Grob-type fragmentation to realise the intermediate.

In 2014, Chen *et al.* used the reduction of a cycloheptatriene to trigger an acid-induced rearrangement in the formal synthesis of α -acorenone (Scheme 21).⁷⁶





Scheme 19 Treatment of an aldehyde with methyl vinyl ketone in base in the synthesis of (-)-acorenone and (-)-acorenone B.



Scheme 20 Three-step procedure in the synthesis of acorenone B.

3.2 Spirovetivanes: agarospirol, hinesol, β-vetivone, α-vetispirene and β-vetispirene

Acetal formation followed by reduction and treatment with acid was used to form the molecules hinesol, β-vetivone, and α- and β-vetispires^{77,78} (Scheme 22). These syntheses were published in a series of articles from Yamada *et al.* in 1973.

A Lewis acid-catalysed Prins cyclisation was used by McCurry *et al.* in 1973 in the synthesis of racemic β-vetivone and β-vetispiene⁷⁹ (Scheme 23).

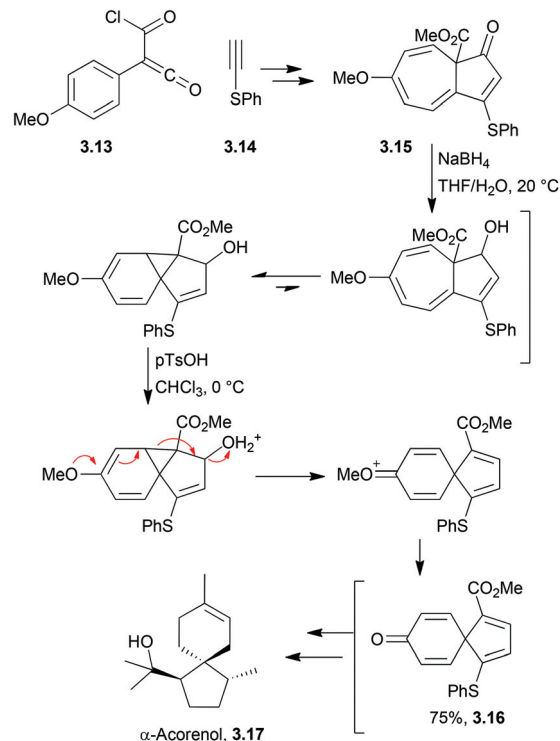
Of the four possible diastereomers, the two shown in Scheme 23 constituted 97% of the reaction mixture. The mechanism is thought to proceed *via* a 6-membered ring transition state.

Deslongchamps *et al.* have synthesised racemic agarospirol and hinesol from a keto ester.^{80,81} The spirocycle was formed by treatment of a cyclopropane intermediate with acid (Scheme 24).

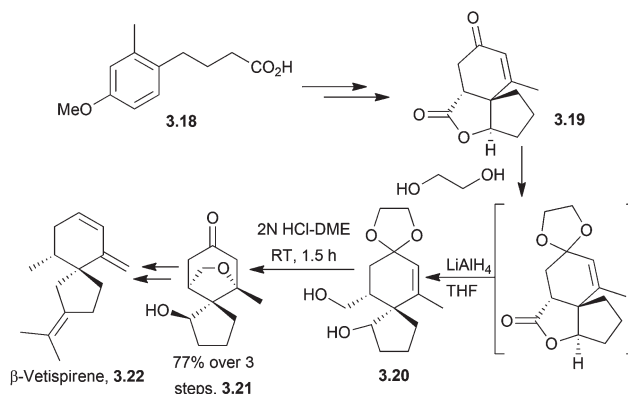
The intermediate is thought to undergo facile cyclopropane ring-opening under acid conditions *via* the enol.

Spiroannulation of 5-methyl-1,3-cyclohexanedione with an alkyl halide in base was used to synthesise β-vetivone⁸² (Scheme 25) by Stork *et al.* in 1973.

A similar method was used by Markó *et al.* in 2004 to form racemic agarospirol, hinesol and α-vetispiene. A spiro di-



Scheme 21 Reduction and acid-induced rearrangement in the synthesis of α-acorenone.



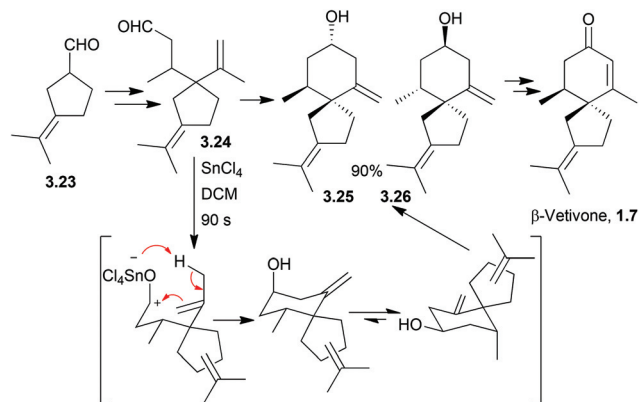
Scheme 22 Three-step sequence in the synthesis of β-vetispiene.

ketone was formed by condensation of a silyl enol ether and an *ortho* ester to form a β-ketoketal followed by base-catalysed intramolecular cyclisation⁸³ (Scheme 26).

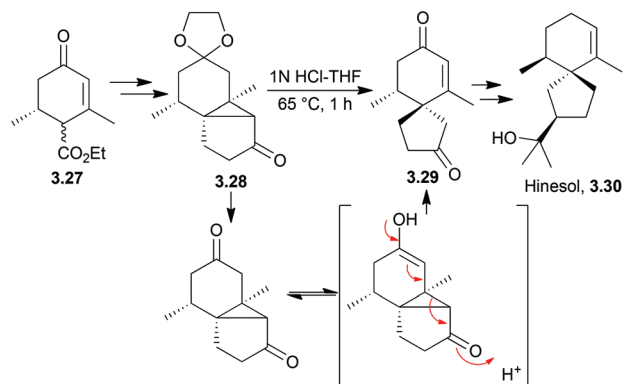
In 1988, Posner *et al.* used a tandem oxidation-acid-promoted cyclisation in the third asymmetric total synthesis of (-)-β-vetivone (Scheme 27).⁸⁴

Acidic or basic conditions are commonly employed in the key cyclisation step of (±)-hinesol.^{85,86} A base-catalysed reverse Prins reaction was used by Marshall *et al.* in 1970⁸⁷ (Scheme 28).

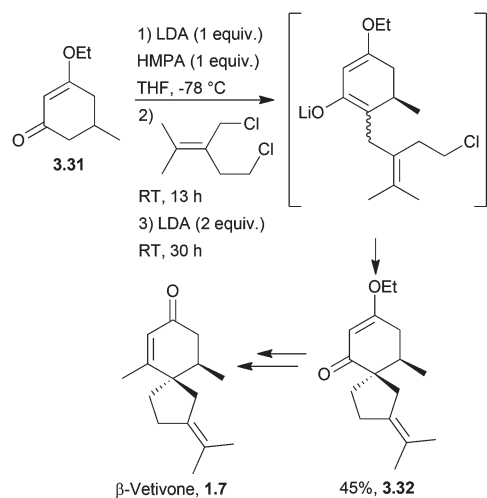




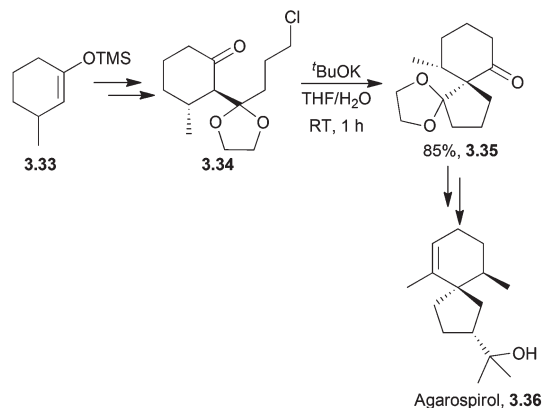
Scheme 23 Prins cyclisation followed by chair-chair interconversion in the synthesis of β -vetivone. Diastereomers **3.25** and **3.26** were separable, but the dr was not stated.



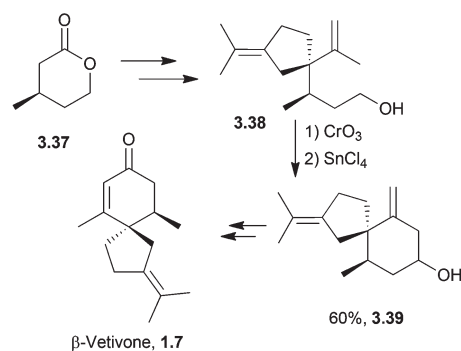
Scheme 24 Treatment of an intermediate with acid in the synthesis of hinesol.



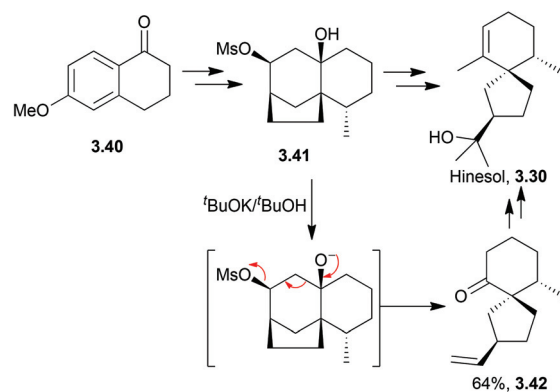
Scheme 25 Spiroannulation in base during the synthesis of β -vetivone.



Scheme 26 Base-catalysed intramolecular cyclisation during the synthesis of agarospirol. The de of intermediate **3.45** was >95%.



Scheme 27 Oxidation then cyclisation in the synthesis of β -vetivone.



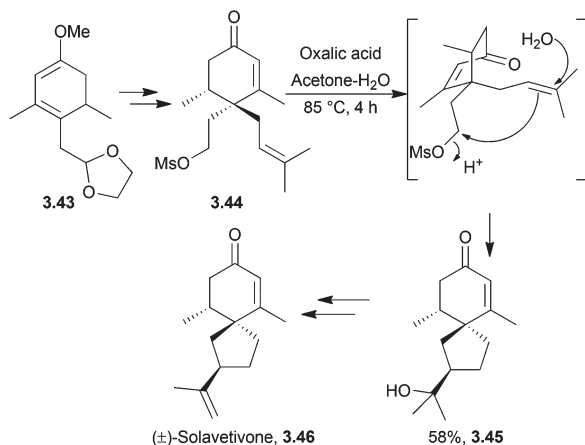
Scheme 28 Basic conditions for the spirocyclisation step during the synthesis of hinesol.

3.3 Spirovetivadienes: solavetivone, lubimin, oxylubimin, premnaspirodiene and hinesene

Solavetivone is a spirovetivadiene phytoalexin produced by tobacco plants infected with the tobacco mosaic virus.^{88–91}

Lubimin and oxylubimin are two of several stress metabolites





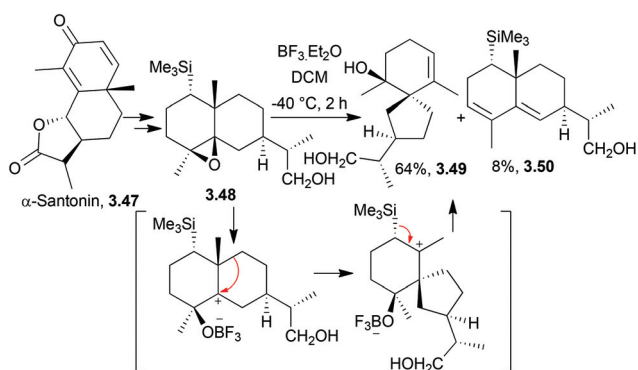
Scheme 29 Oxalic acid-induced cyclisation in the synthesis of (±)-solavetivone.

produced by fungally-infested white potato tubers.^{92,93} Premnaspirodiene and hinesene are diastereomers: (–)-premnaspirodiene is the (2*R*,5*S*,10*R*)-isomer whereas hinesene has the (2*R*,5*S*,10*S*) configuration.

Racemic solavetivone was synthesised through a Diels–Alder reaction followed by π -cyclisation⁸⁵ (Scheme 29) by Murai *et al.* in 1984. The starting material was 3,5-dimethylanisole and the procedure required 12 steps to give the product in 3% overall yield. The Murai group used this methodology to synthesise lubimin and oxylubimin in a series of follow-up articles.^{94,95}

Other conditions attempted for the π -cyclisation were unsuccessful, including using SnCl₄ in DCM at –78 °C, acetic acid at 110 °C and formic acid at 50 °C.

Premnaspirodiene and hinesene have both been synthesised from α -santonin by Pedro *et al.* in 2004.⁹⁶ The key step was an acid-promoted rearrangement of the carbon skeleton (Scheme 30) *via* a tertiary carbocation formed by ring-opening of the epoxide.



Scheme 30 Acid-promoted rearrangement of the carbon skeleton in the synthesis of premnaspirodiene.

3.4 Vitrenal

Takahashi synthesised racemic vitrenal in 12 steps and 7% overall yield from the monoterpene piperitenone⁹⁷ in 1982. The key step was an acid promoted acetal hydrolysis and enol ether trapping (Scheme 31).

3.5 Spirolaurenone

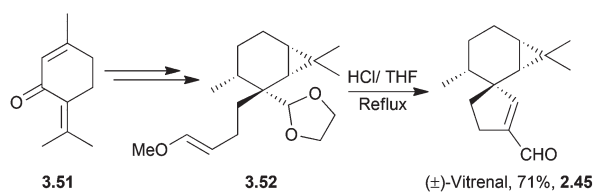
Spirolaurenone is a bromine-containing sesquiterpenoid⁹⁸ based upon the spiroaurane skeleton.⁹⁹

Total synthesis of (±)-spirolaurenone proceeded from a bromohydrin of homogeranitrile in 13 steps and 2% overall yield¹⁰⁰ as described in 1982 by Murai *et al.* The key step was the cleavage of a cyclopropane ring under acidic conditions to give a mixture of *endo* and *exo* alkenes which were separated by preparative HPLC (Scheme 32). The *endo* isomer was used to reach the natural product.

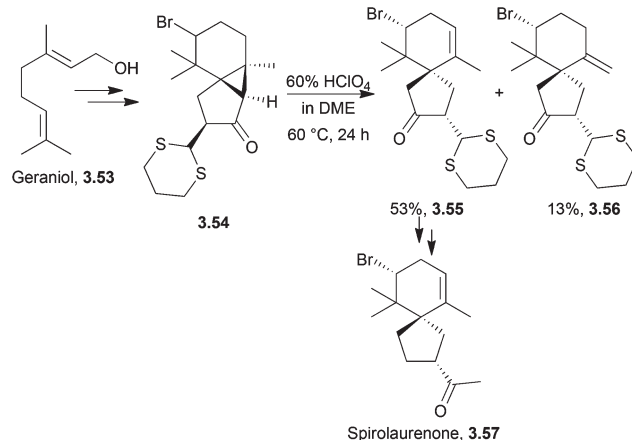
3.6 Chamigrenes: α -chamigrene and majusculone

Unlike its halogenated derivatives, α -chamigrene was isolated from the five-flavour berry, *Schisandra chinensis*, rather than a marine organism. Racemic α -chamigrene was synthesised by Canonne *et al.* in 1991 by intramolecular enolate alkylation promoted by KH¹⁰¹ (Scheme 33).

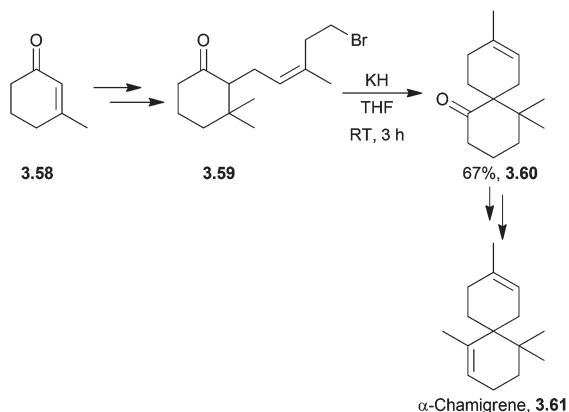
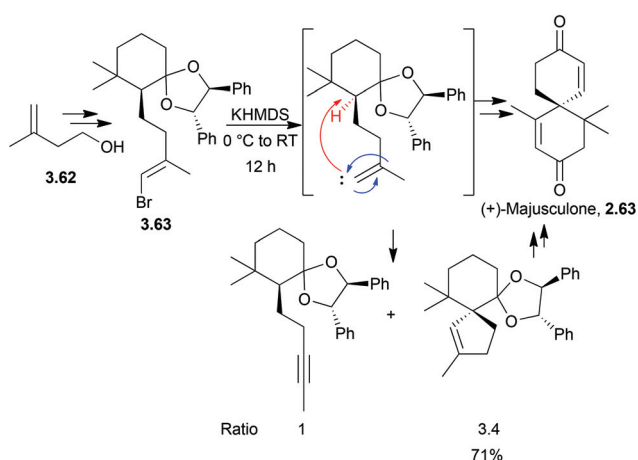
The first enantioselective synthesis of (+)-majusculone was achieved by alkylidene carbene insertion promoted by KHMDS¹⁰² (Scheme 34) as described by Taber *et al.* in 2007.



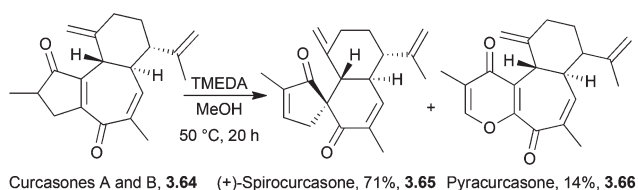
Scheme 31 Acid-catalysed hydrolysis in the synthesis of (±)-vitrenal.



Scheme 32 Spirocyclisation step in the synthesis of spirolaurenone.

Scheme 33 Spirocyclisation step in the synthesis of α -chamigrene.

Scheme 34 Spirocyclisation in the synthesis of (+)-majusculone.



Scheme 35 Use of TMEDA in the synthesis of spirocurcasone.

α -Elimination generated the carbene which inserted into a C–H bond shown in red to give the spirocycle. An alkyne by-product from the 1,2-rearrangement was also observed, shown in blue. Concerns over the ability of the carbene to insert into the deactivated, congested methine were not realised.

3.7 Spirocurcasone

Spirocurcasone is a diterpenoid with a novel carbon skeleton, isolated from *Jatropha curcas*.¹⁰³ A high-yielding one-pot semi-synthesis of (+)-spirocurcasone from the natural products curcasones A and B has been reported very recently by Qin *et al.* in 2014 (Scheme 35).¹⁰⁴

In the absence of a metal, the TMEDA is thought to act as a base. Various conditions were screened, and MeOH was found to give higher conversions of the starting material over toluene or DCM, and the relatively mild temperature of 50 °C was more effective than 90 °C or reflux.

4. Ring-closing metathesis

Spirocarbocycles can be synthesised using a variety of metal complexes.¹⁰⁵ Ring-closing metathesis^{106,107} typically uses a ruthenium-based catalyst such as Grubbs' I or Grubbs' II to react two terminal alkenes intramolecularly to give an alkene and ethene. Kotha *et al.* have comprehensively reviewed the synthesis of spirocyclics by ring-closing metathesis in 2003.¹⁰⁸

4.1 Illudins

The illudins are sesquiterpenoids isolated from fungi, including the highly poisonous Jack-o' lantern mushroom, *Omphalotus illudens*.^{109–111} The illudins themselves are extremely toxic. Illudins M and S show potent cytotoxic activity *in vitro*, but are much less effective *in vivo*.^{112,113} There has been considerable work towards developing anti-cancer derivatives, which are now in clinical trials. These include bicyclic and isomeric analogues of illudin M,¹¹⁴ acylfulvene and hydroxymethylacylfulvene^{115,116} (Fig. 5). The latter is now in Phase II clinical trials against ovarian, prostate, and gastrointestinal cancers.¹¹⁷

The illudin core has been synthesised by Movassaghi *et al.* using an enyne ring-closing metathesis cascade¹¹⁸ (Scheme 36) in 2009.

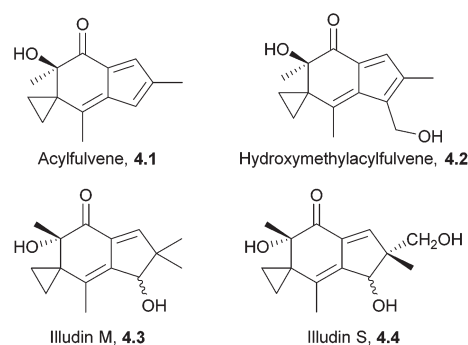
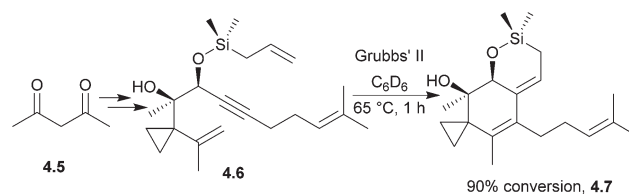
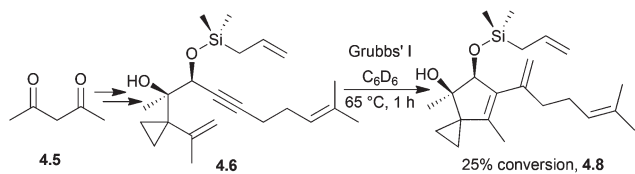


Fig. 5 Acylfulvene, hydroxymethylacylfulvene and illudins M and S.



Scheme 36 Enyne ring-closing metathesis in the synthesis of the illudin core.





Scheme 37 Enyne ring-closing metathesis in the synthesis of the illudin core to give an unwanted side-product.

With Grubbs' II catalyst, conversion to the desired product was 90% by ^1H NMR spectroscopy, whereas with Grubbs' I catalyst the conversion was only 25% to an unwanted side-product that contained a five-membered ring (Scheme 37).

Illudins M and S have also been previously synthesised by Matsumoto *et al.*, using a basic aldol ring-closing reaction.^{119,120}

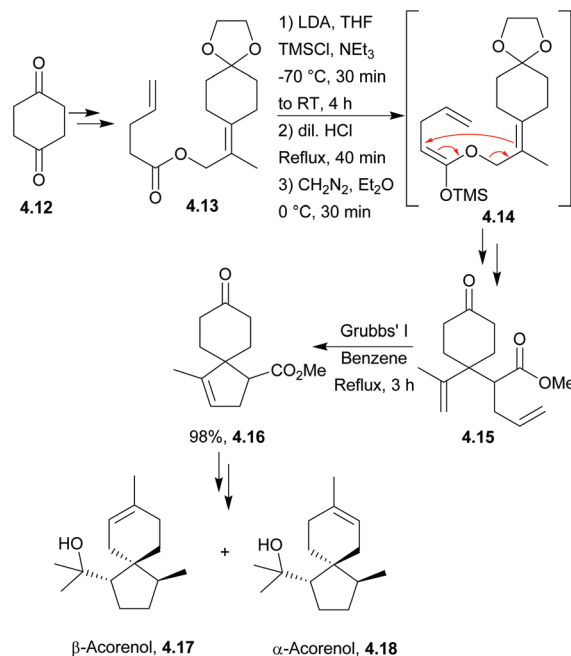
4.2 Acoranes: trichoacorenol, α -acorenol, β -acorenol, acorone, isoacorone, acorenone and α -acoradiene

Trichoacorenol is a sesquiterpenoid¹²¹ produced by the fungi *Trichoderma koningii* and *T. harzianum*.^{121,122} The unnatural enantiomers (–)-trichoacorenol and (–)-acorenone have both been synthesised from (+)-(*R*)-pulegone¹²² by Dickschat *et al.* in 2011. The key step was a ring-closing metathesis which occurred in moderate yield for such a transformation (Scheme 38).

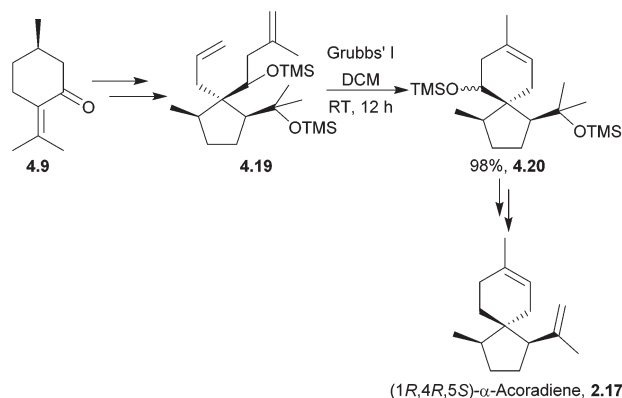
α -Acorenol and β -acorenol were synthesised starting from cyclohexan-1,4-dione in 7 steps and 67% overall yield³³ by Srikrishna *et al.* in 2007. The key steps were an Ireland–Claisen rearrangement and ring-closing metathesis (Scheme 39). The same methodology had previously been applied to the formal syntheses of ($\pm$)-acorone and ($\pm$)-isoacorone.¹²³

The (1*R*,4*R*,5*S*) enantiomer of α -acoradiene has been synthesised by Mori *et al.* in a 14 step sequence, using Grubbs' I catalyst in the key spirocyclisation (Scheme 40).¹²⁴

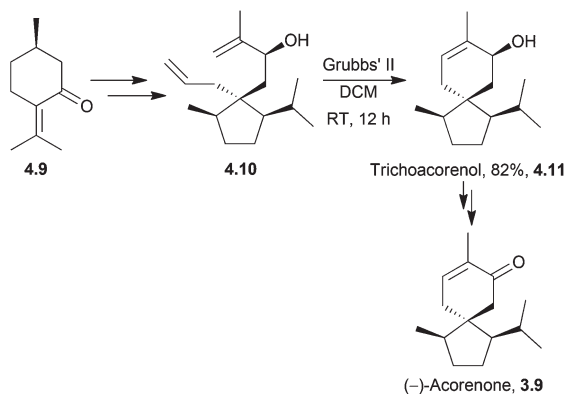
The stereochemistry of the product formed did not match the stereochemistry of the broad-horned beetle pheromone,



Scheme 39 Ring-closing metathesis in the synthesis of ($\pm$)- α -acorenol and ($\pm$)- β -acorenol.



Scheme 40 Ring-closing metathesis in the synthesis of α -acoradiene.



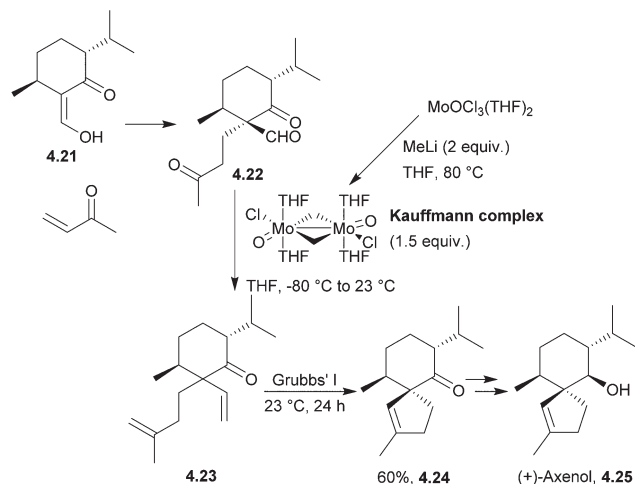
Scheme 38 Ring-closing metathesis during the synthesis of (–)-trichoacorenol and (–)-acorenone.

thus disproving the originally proposed structure and identifying the pheromone as (1*S*,4*R*,5*R*)- α -acoradiene.³⁸

4.3 Spiroaxanes: axenol and gleenol

Axenol and gleenol are enantiomeric sesquiterpenes sharing the spiroaxane skeleton. They have both been isolated from the Eurypon marine sponges, but (–)-gleenol has also been found in a variety of coniferous trees, including *Picea glehnii*, *P. koraiensis*, *Cryptomeria japonica* and *Juniperus oxycedrus*. Axenol has not been found to exhibit biological activity, but (+)-gleenol possesses termiticidal and anti-helminthic activities, as well as regulating the growth of plant seeds.





Scheme 41 Ring-closing metathesis in the synthesis of (+)-axenol.

(+)-Axenol and (–)-gleenol have been synthesised by Spitzner *et al.*, using an organometallic domino reaction, with an unusual combination of methylenation followed by ring-closing metathesis^{125,126} (Scheme 41). The first step in the synthesis was a Michael addition with methyl vinyl ketone,¹²⁷ followed by selective methylenation with the Kauffmann molybdenum complex which was generated *in situ* from $\text{MoOCl}_3(\text{THF})_2$.¹²⁸ A mixture of inseparable isomers was formed in the ratio 2 : 1, of which the major (*R*)-isomer (4.22) was the desired product. Grubbs' I catalyst was added to the crude reaction mixture in a one-pot reaction to give epimers of the spiroketone 4.23.

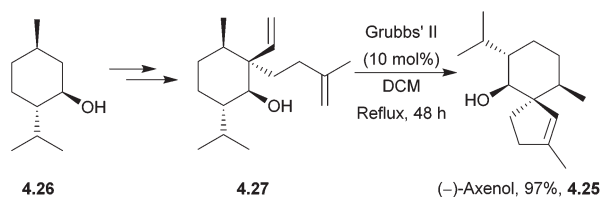
A McMurry coupling was also directly attempted on the tri-carbonyl, but the spiroketone product was not observed by GC-MS analysis.

Kobayashi *et al.* synthesised (–)-axenol in 2015 using ring-closing metathesis (Scheme 42).¹²⁹ The key stereochemistry of the quaternary carbon in the intermediate was built using allylic substitution starting from a picolinate derived from menthol.

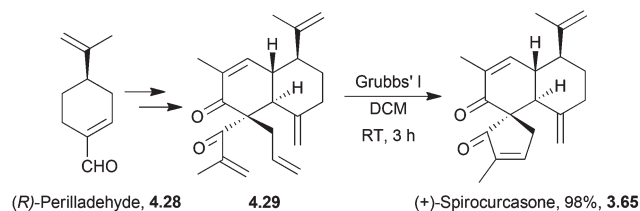
The TBS-protected alcohol was also considered, but gave lower conversions of substrate.

4.4 Spirocurcasone

The total synthesis of (–)-spirocurcasone was achieved by ring-closing metathesis of a pentaene¹³⁰ by Ito *et al.* in 2013



Scheme 42 Ring-closing metathesis in the synthesis of (–)-axenol.



Scheme 43 Ring-closing metathesis in the synthesis of spirocurcasone.

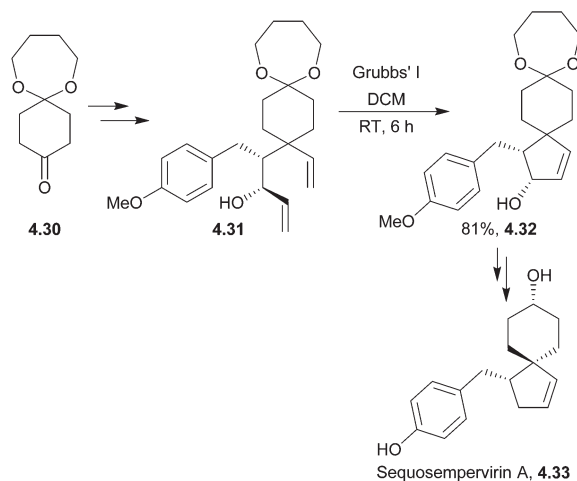
(Scheme 43). No protecting groups were required in the 9-step procedure starting from (*R*)-perillaldehyde.

4.5 Sequoempervirin A

Sequoempervirin A is a novel spiro-norlignan compound isolated in 2004 from the branches and leaves of the California redwood *Sequoia sempervirens*.¹³¹ Three years later, the first total synthesis of sequoempervirin A used ring-closing metathesis as the spirocyclisation step leading to a cyclic allylic alcohol en route to the final product¹³² (Scheme 44).

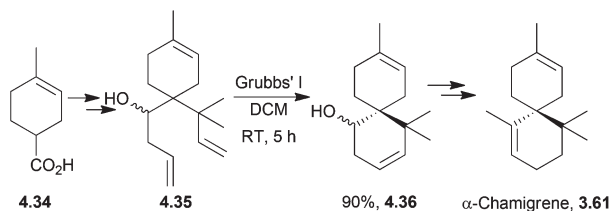
4.6 Chamigrenes: α-chamigrene, β-chamigrene, laurencenones B and C and elatol

Other members of the chamigrene natural product family not previously mentioned include β-chamigrene, laurencenone B and elatol. The potent and varied biological activity of elatol makes it a compound of significant interest. It has shown anti-biofouling activity,¹³³ anti-bacterial activity,^{134–136} anti-fungal activity¹³⁷ and cytotoxicity.¹³⁸ It was isolated from red algae *Laurencia* sp., as were laurencenones B and C. Like α-chamigrene, β-chamigrene has been isolated from *Schisandra chinensis*, although it was first isolated from the leaf oil of the cypress *Chamaecyparis taiwanensis*.¹³⁹



Scheme 44 Ring-closing metathesis during the synthesis of sequoempervirin A.





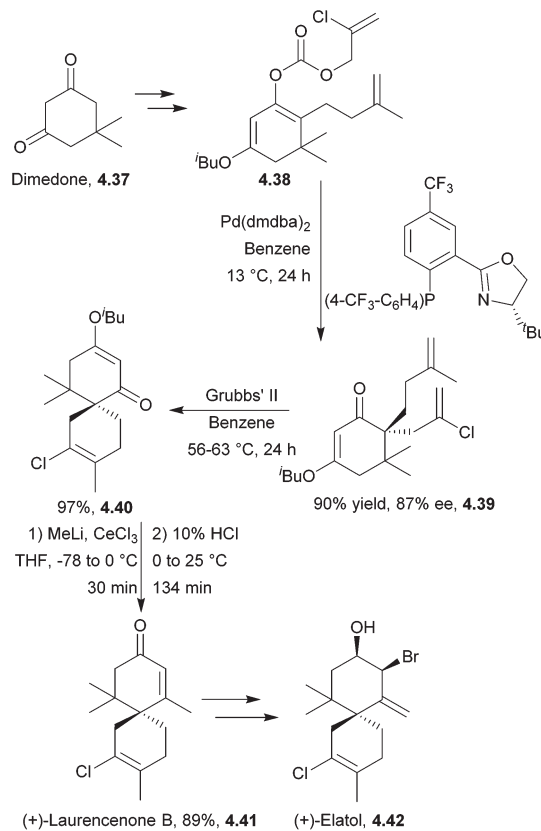
Scheme 45 Ring-closing metathesis in the synthesis of racemic α -chamigrene.

($\pm$)- α -Chamigrene was synthesised from cyclohexenecarboxylic acid¹⁴⁰ by Srikrishna *et al.* in 2006. The procedure used an Ireland–Claisen rearrangement and ring-closing metathesis as previously demonstrated by the same group in the synthesis of ($\pm$)-isoacorone to give the product in 11 steps and 32% overall yield (Scheme 45).

Racemic β -chamigrene and laurencenone C have been synthesised by an Ireland–Claisen rearrangement, followed by either ring-closing metathesis¹⁴⁰ or an intramolecular type II carbonyl ene reaction¹⁴¹ by Srikrishna *et al.* The metathesis method required 11 steps to give the product in 32% overall yield starting from a cyclohexenecarboxylic acid. For laurencenone C, the initial method required 10 steps to give a 17% overall yield.

(+)-Laurencenone B, (+)-elatol¹⁴² and (–)-laurencenone C¹⁴³ have all been synthesised by ring-closing metathesis by Stoltz *et al.* The starting material for both laurencenones was dimedone. The synthesis required 7 steps to give the products in 34% and 31% overall yield respectively (Scheme 46). (+)-Elatol required 9 steps to give the product in 11% overall yield.

This catalytic asymmetric method was generalised to the chamigrene product family, using a palladium-based enantioselective decarboxylative allylation followed by ring-closing metathesis.¹⁴³



Scheme 46 Ring-closing metathesis in the synthesis of (+)-laurencenone B and (+)-elatol.

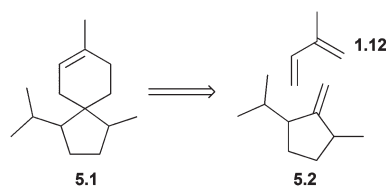


Fig. 6 Retrosynthetic analysis of the carbon skeleton of collettoic acid.

5. Diels–Alder reaction

The Diels–Alder reaction is a valuable tool in total synthesis,¹⁴⁴ since it forms two carbon–carbon bonds and up to four stereocentres. Its application to the synthesis of spirocarbocycle-containing natural products is described below.

5.1 Acoranes: collettoic acid, β -, γ -, δ -acoradienes, acorone and isoacorone

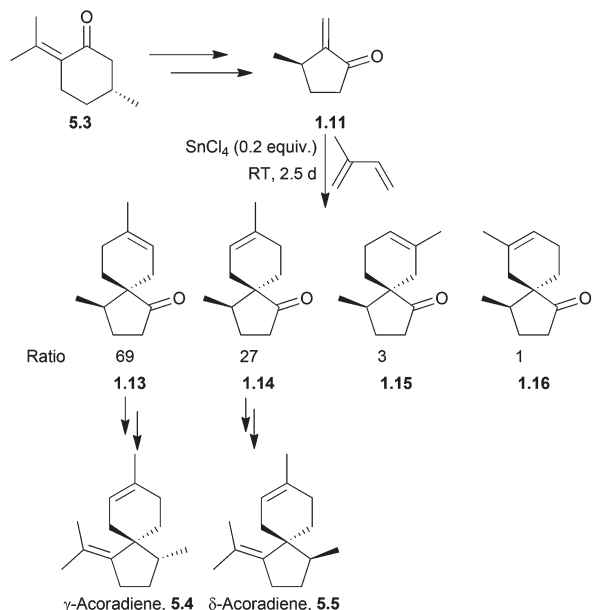
For the avoidance of confusion, it should be noted that γ - and δ -acoradiene are also known as α - and β -alaskene respectively, due to their isolation from the Alaskan cedar, *Chamaecyparis nootkatensis*.^{145,146}

Retrosynthetic analysis of the acorane carbon skeleton reveals that the cyclohexene can be broken into two units, of which one is isoprene (Fig. 6).

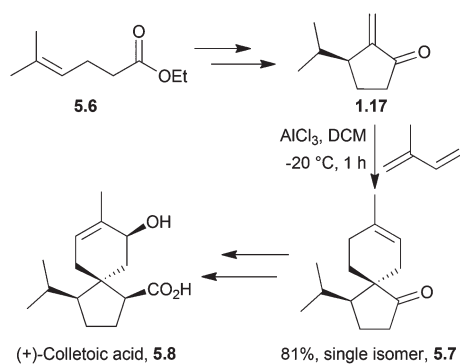
γ - and δ -Acoradiene were synthesised by Marx *et al.* as early as 1973¹⁴⁷ using a method which was extended to β -acora-

diene, acorone, and isoacorone two years later¹⁴⁸ (Scheme 47). The starting material was (*R*)-pulegone, and a SnCl_4 -catalysed Diels–Alder reaction was used. The major products were those formed from the electronically favourable *para* orientation of the reagents, whilst the minor products were formed from the *meta* orientation. The Lewis acid increased selectivity for the products of *para* cycloaddition. The reaction had been performed without SnCl_4 , with heating to 100 °C and gave the same products, in the ratio 45 : 25 : 20 : 10. The transition state of the major isomer was formed by attack of the isoprene unit on the opposite face to the methyl group, reducing unfavourable steric interactions, and eventually leading to the formation of γ -acoradiene. The second isomer was formed by attack of the isoprene on the same face as the methyl group, which was then through further functionalisation converted to δ -acoradiene.





Scheme 47 Lewis-acid catalysed Diels–Alder reaction with isoprene in the synthesis of γ - and δ -acoriadienes.

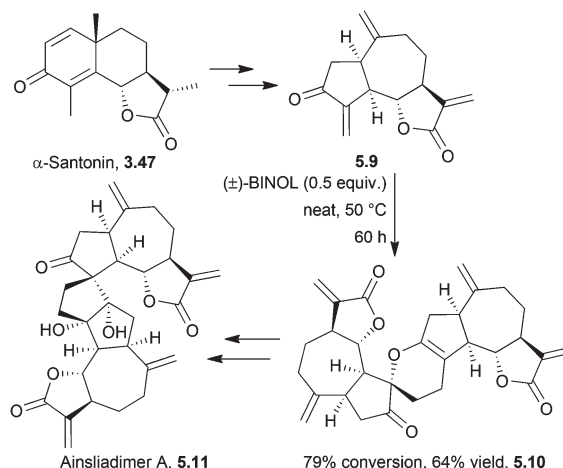


Scheme 48 Lewis-acid catalysed Diels–Alder reaction in the synthesis of (+)-colletoic acid.

(+)-Colletoic acid is a sesquiterpenoid also from the acorane family and a potent inhibitor of the 11β -hydroxysteroid type 1 enzyme, making it a therapeutic target for metabolic syndrome.¹⁴⁹ This compound was generated as a single enantiomer in 2013 by a diastereoselective intramolecular 5-*exo*-Heck reaction.¹⁵⁰ At the same time, Nakada *et al.* used a stereoselective Lewis-acid catalysed Diels–Alder reaction with isoprene to create the spirocentre^{151,152} (Scheme 48). It was found that the starting material for the Diels–Alder reaction decomposed at high temperatures, but when run at $-20\text{ }^{\circ}\text{C}$ in the presence of the Lewis acid AlCl_3 , the reaction gave the desired product as a single isomer.

5.2 Ainsliadimers A and B and gochnatiolides A–C

The ainsliadimers and gochnatiolides are a related set of dimeric sesquiterpene lactones, biosynthesised from the guaianolide dehydrozalanin C.



Scheme 49 Hetero-[4 + 2] Diels–Alder reaction in the synthesis of ainsliadimer A.

A biomimetic total synthesis of ainsliadimer A in 2010 by Lei *et al.* proceeded in 14 steps, starting from α -santonin¹⁵³ (Scheme 49). The key step was a hydrogen bond-promoted [4 + 2]-hetero Diels–Alder dimerisation.

During an investigation into hydrogen bond donor catalysis, hydrogen-bonding solvents were used but only chloroform gave a trace amount of the dimer when the monomer was heated at $35\text{ }^{\circ}\text{C}$ for 12 h. Additives including triethylamine or hydrochloric acid did not improve conversion, and longer reaction times and higher temperatures led to increased decomposition. Hydrogen bond donor catalysts were studied, and racemic BINOL was found to be particularly effective at catalysing the dimerisation.

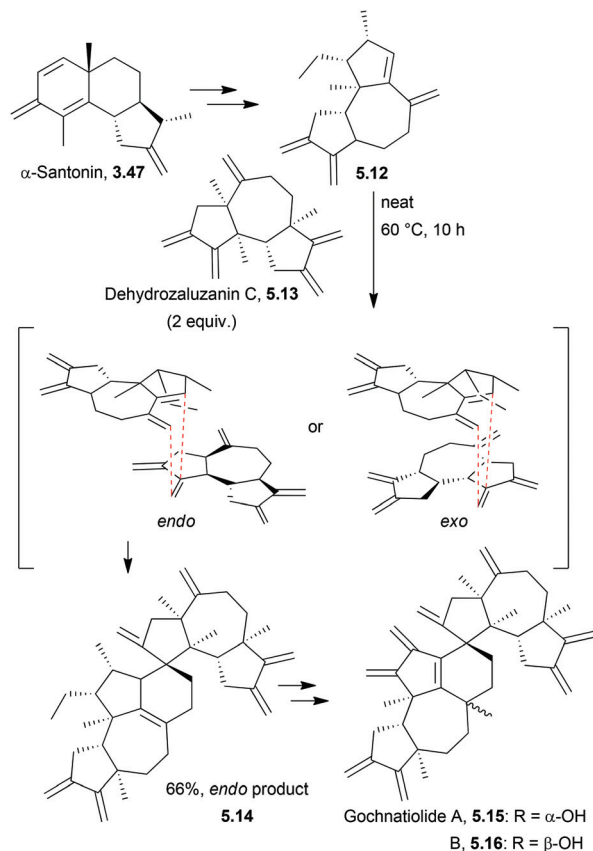
More recently, Qin *et al.* reported the total syntheses of ainsliadimer B and gochnatiolides A and B which proceeded in 25 steps and 1% overall yield using a cross Diels–Alder cycloaddition¹⁵⁴ (Scheme 50).

The Diels–Alder reaction could proceed *via* either an *endo* or *exo* transition state, but only the product arising from the *endo* transition state was observed. The *exo* transition state would experience a high degree of steric interaction between the silyl groups on the bottom face of the diene and the seven-membered ring of the dehydrozalanin C. When attempted in a variety of solvents, the reaction proceeded slowly to give the Diels–Alder product in less than 15% yield and so the reaction was run neat. Addition of acids such as ceric ammonium sulfate, pyridinium *p*-toluenesulfonate, tosic acid and acetic acid failed to catalyse the reaction, instead causing the decomposition of the dehydrozalanin C product.

6. Claisen rearrangement

The Claisen rearrangement is a [3,3]-sigmatropic rearrangement where an allyl vinyl ether is converted to an unsaturated





Scheme 50 Cross Diels–Alder *via* an *endo* intermediate in the synthesis of ainsliadimer B and gochnatiolides A and B.

carbonyl.^{155,156} Its use in the synthesis of spiro sesquiterpenoids has been recently reviewed by Kobayashi *et al.*¹⁵⁷

6.1 Acoranes: acorone, isoacorone, α -acorenel and β -acorenel

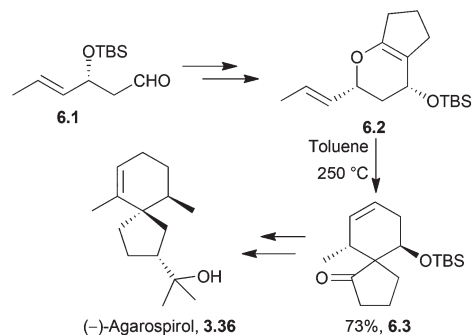
As previously mentioned, ($\pm$)-acorone and ($\pm$)-isoacorone have been formally synthesised by the Claisen rearrangement followed by ring-closing metathesis. The Srikrishna group synthesised racemic α -acorenel and β -acorenel employing this strategy (see above).

6.2 Spirovetivanes: agarospirol, hinesol, β -vetivone and α -vetispirene

(–)-Agarospirol,^{158,159} (+)- α -vetispirene, hinesol (an enantiomer of agarospirol) and β -vetivone have all been synthesised by the Kobayashi group using Claisen rearrangement methodologies^{160,161} (Scheme 51).

6.3 Spiroaxanes: axenol, axisonitrile-3 and gleenol

Axisonitrile-3 is a nitrogenous sesquiterpenoid of interest due to its significant biological activity, including anti-fouling activity against barnacle larvae,¹⁶² cytotoxicity¹⁶³ and potent



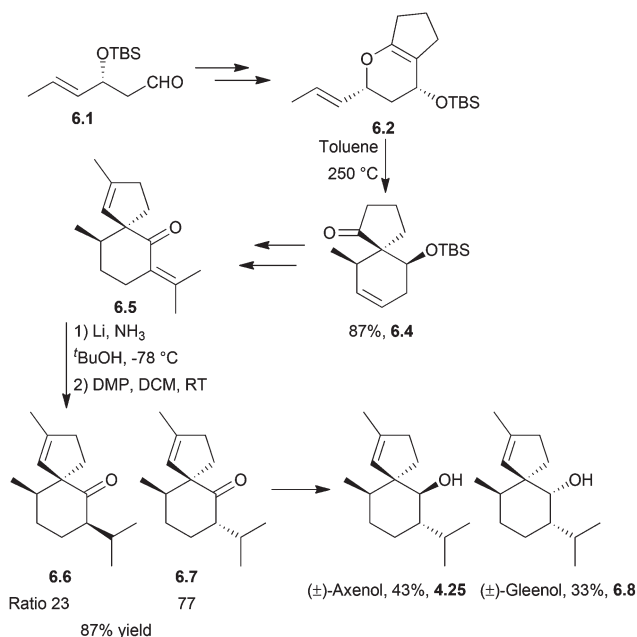
Scheme 51 Claisen rearrangement in the synthesis of agarospirol. The intermediate **6.3** was present in >95% dr.

anti-malarial activity.^{164,165} It possesses an unusual isonitrile group. Axenol is a key intermediate in its synthesis.

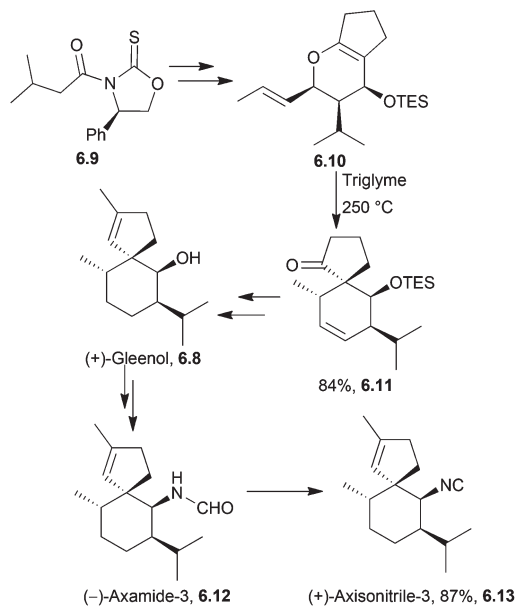
Racemic axenol and gleenol have been synthesised by the Kobayashi group in 2007,^{166,167} applying the same methodology as for the synthesis of other spiro[4.5]decenes described above.

Reduction and re-oxidation of the spiroketone gave a mixture of enantiomers in the ratio 77:23, which were epimerised by treatment with ethanolic KOH and then separated by column chromatography. Reduction with LiAlH_4 gave a mixture of racemic gleenol and axenol (Scheme 52).

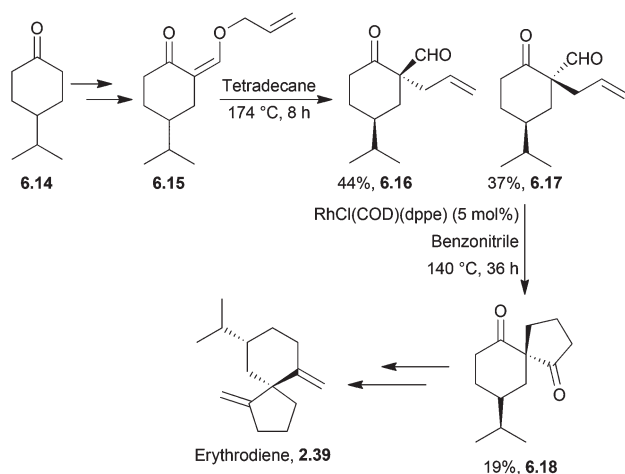
A Claisen rearrangement was used by the same group to synthesise (+)-axisonitrile-3 *via* (+)-gleenol and (–)-axamide-3¹⁶⁸ in 2009 (Scheme 53).



Scheme 52 Claisen rearrangement in the synthesis of ($\pm$)-gleenol and ($\pm$)-axenol.



Scheme 53 Claisen rearrangement in the synthesis of (+)-axisonitrile-3. The dr of the intermediate **6.11** was >95%.



Scheme 54 Rhodium-catalysed Claisen rearrangement in the synthesis of erythrodiene.

6.4 Erythrodiene and spirojatamol

Eilbracht *et al.* reported a formal synthesis of erythrodiene and spirojatamol which used a rhodium-based catalyst for a tandem Claisen rearrangement-hydroacylation of an allyl vinyl ether¹⁶⁹ (Scheme 54). This method was also applied in 1996 for the synthesis of acoradienes and spirovetivanes as they share the spiro[4.5]decanone skeleton.¹⁷⁰

The acoradiene and spirovetivane procedure was performed in one pot,¹⁷¹ and the particular rhodium catalyst chosen was selected because it did not decompose at the required higher temperatures and thus could catalyse the Claisen and hydroacylation reactions.¹⁷²

7. Photochemical reaction

Photochemistry is a powerful tool used in total synthesis to access products that can be otherwise difficult to form. Its use in natural product synthesis has been extensively reviewed.^{173–175}

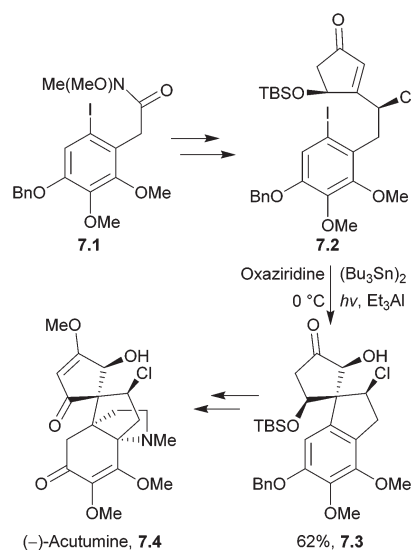
7.1 Acutumine

Acutumine is a tetracyclic alkaloid which shows selective T-cell cytotoxicity and anti-amnesic properties.¹⁷⁶

Castle *et al.* reported the first total synthesis of (–)-acutumine in 2009, which utilised a photochemical radical-polar crossover reaction to form the spirocycle¹⁷⁷ (Scheme 55).

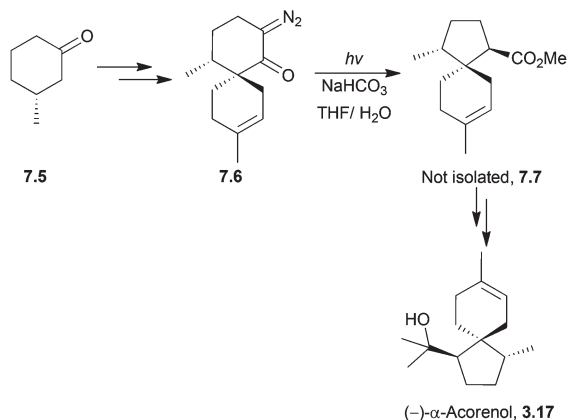
The cascade process was observed at 0 °C with a sun-lamp. Optimisation experiments identified Et₃Al as a more effective promoter than Et₃B or Et₂Zn, and 3-phenyl-2-(phenylsulfonyl)oxaziridine as a better hydroxylating agent than oxygen, DMDO or ^tBuOOH. Side-products were observed, with the alcohol group replaced by an iodide or hydrogen. Attempts were made to convert the iodide into the desired product by treatment with diethyl zinc, oxygen and the oxaziridine gave the desired product in 62% yield, thus increasing the overall yield to 66%. Samarium diiodide/HMPA was considered as a replacement for (Bu₃Sn)₂, but only gave a low yield of the desired product, and led to the formation of many side-products. This was attributed to the presence of functional groups in the starting material that could react with SmI₂. The mechanism still requires further work to develop a better understanding.

Reisman *et al.* have attempted the synthesis of (–)-acutumine *via* a photochemical [2 + 2]-cycloaddition, but were only successful in constructing the aza-propellane core.¹⁷⁸ (–)-Acutumine has also been synthesised through a Hosomi-Sakurai allylation in 17 steps¹⁷⁹ by Herzon *et al.* in 2013.



Scheme 55 Photochemical reaction in the synthesis of (–)-acutumine. No diastereomers were observed.





Scheme 56 Photolysis in the synthesis of α-acorenol.

7.2 Acoranes: α-acorenol, β-acorenol, (–)-acorenone and (±)-α-acoradiene

Photolysis was used in 1973 by Ramage *et al.* to stereospecifically synthesise (–)-α- and (+)-β-acorenol (Scheme 56).¹⁸⁰

An enone [2 + 2] photocycloaddition was used in 1982 by Baldwin *et al.* to synthesise (–)-acorenone¹⁸¹ (Scheme 57).

The key photochemical step was highly regio- and stereo-specific, with the furanone reacting only in a head-to-tail manner. (–)-Acorenone was synthesised in a total of 13 steps in approximately 8% overall yield.

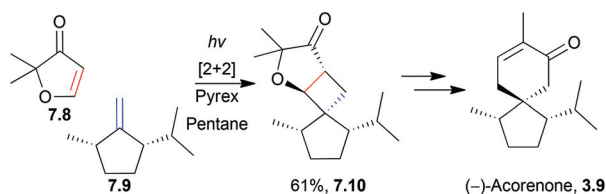
Irradiation of an enone was also used as the key step by Oppolzer *et al.* in 1983 in the synthesis of racemic α-acoradiene¹⁸² (Scheme 58).

The low yield was ascribed to the difficulty in separating the various isomers produced by the radical reaction. Formation of the diradical led to a loss of stereochemical information, and resulted in the formation of two enantiomers, of which reductive fragmentation of only one gave the desired product.

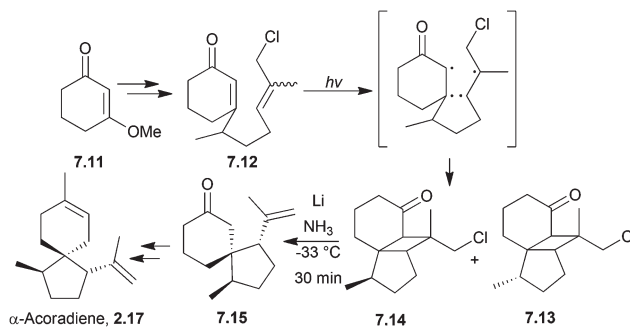
7.3 Spirovetivanes: lubiminol, agarospirol, hinesol, β-vetivone and α-vetispirene

Lubiminol is a spirovetivadiene phytoalexin isolated from infected potatoes^{183,184} and the red pepper *Capsicum annuum*.¹⁸⁵

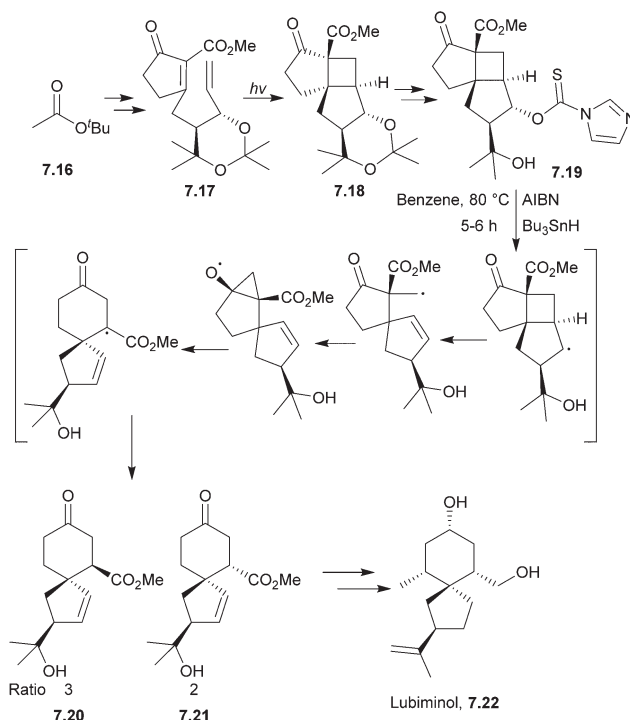
A double diastereoselective intramolecular photocycloaddition has been used to synthesise the spiro centre of lubiminol (Scheme 59), followed by a radical fragmentation–rearrangement reaction, on which Crimmins *et al.* have published a series of articles.^{186–188}



Scheme 57 Enone [2 + 2] photocycloaddition in the synthesis of (–)-acorenone.



Scheme 58 Photochemical cyclobutane formation followed by reductive fragmentation in the synthesis of racemic α-acoradiene. The configurations of 7.13 and 7.14 were not determined because this chirality was lost by reduction.



Scheme 59 Photocycloaddition during the synthesis of lubiminol.

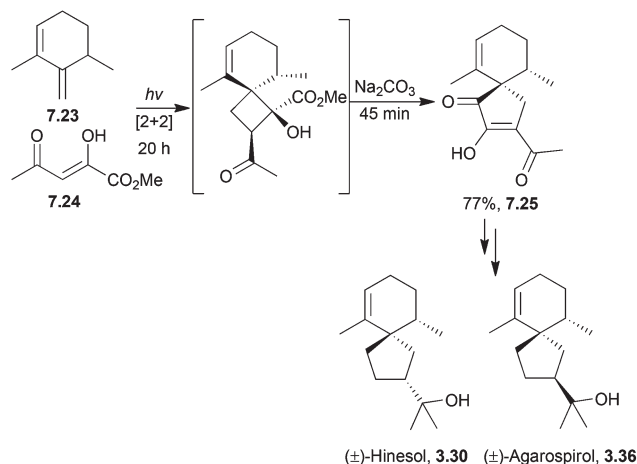
The reaction proceeded *via* a Dowd–Beckwith type ring expansion to give the spirocyclic intermediate.

Takeshita *et al.* used photocycloaddition followed by a base-catalysed benzilic acid rearrangement in 1995 to synthesise racemic hinesol and agarospirol¹⁸⁹ (Scheme 60).

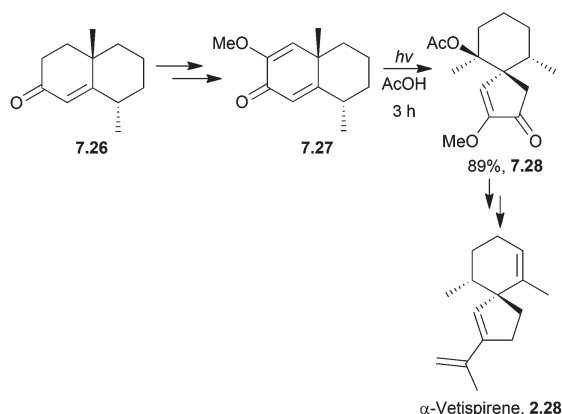
As the rearrangement was not triggered using triethylamine as a base, it was postulated that formation of a sodium chelated intermediate could assist in the rearrangement.

(–)-β-Vetivone has been synthesised by a dienone ring contraction of a spiro starting material with irradiation in acid to form a spiroketone.⁴⁵ Photolysis of a cross-conjugated diene was used by Caine *et al.* to synthesise α-vetispirene¹⁹⁰ (Scheme 61) in 1976.





Scheme 60 Photochemical reaction in the synthesis of racemic hinesol and agarospirol.



Scheme 61 Photolysis in the synthesis of α -vetispirene.

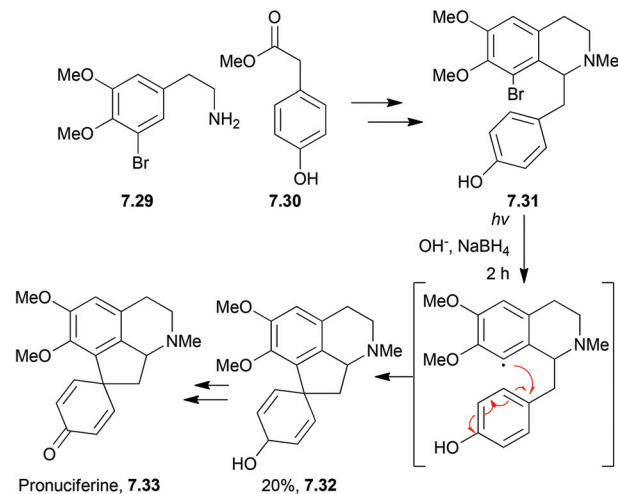
Glacial rather than aqueous acetic acid gave a dramatic improvement in yield (28% to 89%), and a reduction in side-product formation. The absence of water prevented hydrolysis of the enol ether.

7.4 Proaporphines

The proaporphines are a major group of isoquinoline alkaloids. They are biosynthetic precursors to aporphine alkaloids which exhibit a broad range of biological activity.

Photolysis has been used by the Iwata and Fukumoto groups to synthesise pronuciferine,^{191,192} orientalinone,¹⁹³ *O*-methyl-orientalinone,¹⁹⁴ and *O*-methyl-kreysiginone¹⁹⁵ (Scheme 62).

Pronuciferine has been synthesised using methyl vinyl ketone in base,¹⁹⁶ and more recently pronuciferine and stepharine have been synthesised by aromatic oxidation with a hypervalent iodine reagent.¹⁹⁷ This led to the unprecedented C–C bond formation between the *para* position of a phenol and an enamide carbon.

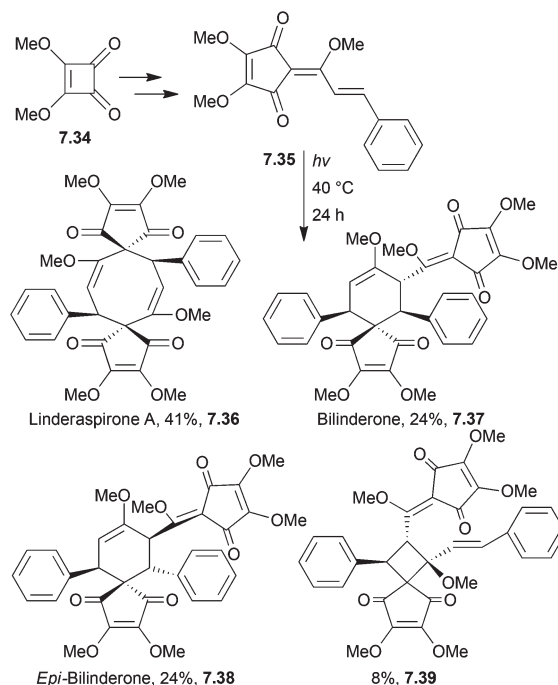


Scheme 62 Photolysis of an intermediate in the synthesis of pronuciferine.

7.5 Bilinderone and linderaspirone A

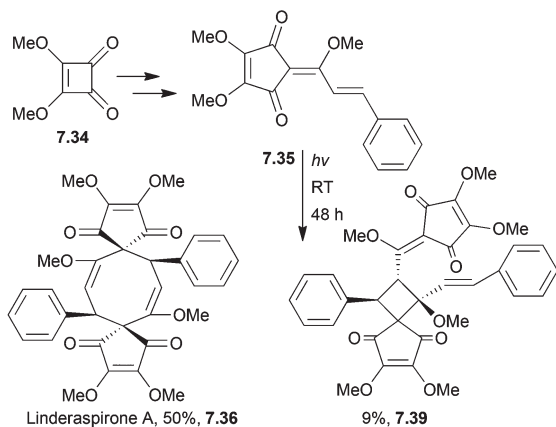
(±)-Bilinderone¹⁹⁸ and (±)-linderaspirone A¹⁹⁹ show promising activity in the improvement of insulin sensitivity. Linderaspirone A is a highly symmetrical compound with a [4.3.4.3] bispiro core, whereas bilinderone has a [4.5]spiro core.

Linderones have been synthesised biomimetically by Wang *et al.* in 2011.²⁰⁰ The key step was a mild photochemical dimerisation. (±)-Linderaspirone A was synthesised by a [2 + 2]-cycloaddition-Cope rearrangement (Scheme 63), whilst



Scheme 63 Photochemical dimerisation in the synthesis of linderaspirone A and bilinderone.





Scheme 64 Photochemical reaction in the synthesis of linderaspirones A.

($\pm$)-bilinderone was synthesised by a [2 + 2]-cycloaddition-radical rearrangement.

Irradiation of the neat starting material as a solid state thin-film produced a mixture of products, including linderaspirones A, bilinderone, *epi*-bilinderone and an additional side product containing an unusual spiro[3.4] system. Photolysis in various solvents, including methanol, acetone, THF, acetonitrile, toluene and benzene all gave yields of linderaspirones A and bilinderone in less than 20%.

In 2013, Hu *et al.* synthesised linderaspirones A in 3 steps as well as bilinderone using a Darzens-type cyclopentadione reaction (Scheme 64).²⁰¹ The starting material was the same as was used above.

When performed under oxygen rather than argon, the yield of linderaspirones A increased from 38% to 50%.

8. Conclusions

Spirocarbocycles are important structural motifs found in drugs, ligands and natural products, and so strategies towards their challenging synthesis are key. This review has examined several common reactions used to form these quaternary centres, including the use of acid and base, particularly in the aldol reaction. Ring-closing metathesis, the Diels–Alder reaction, Claisen rearrangements and photochemistry have also been used to synthesise these spirocarbocycles.

As seen above, the aldol reaction has been successful on a wide range of carbon skeletons, although it lacks stereoselectivity unless the substrate already contains the desired chirality. High yields are achievable with suitable starting materials but if a mixture of products are obtained, the efficiency decreases. This is similarly true for those reactions effected using acid or base, but since these do not operate under one mechanism, it is difficult to generalise. Stereoselectivity depends on both the reagents and the reaction conditions: for example, the possibility of a 6-membered chair transition state can increase selectivity. It is well-known

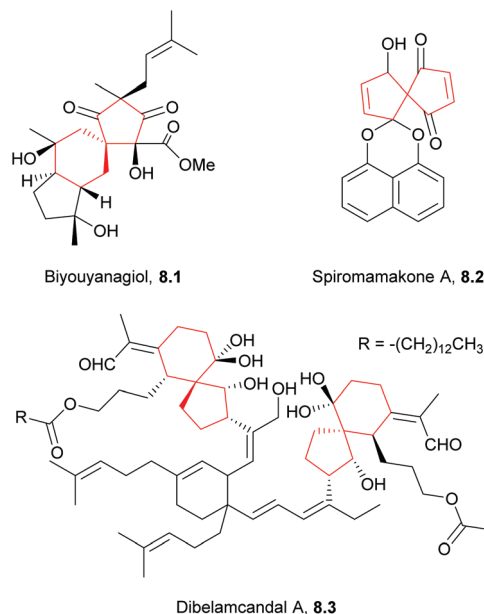


Fig. 7 Unsynthesised natural products containing spirocarbocycles.

that ring-closing metathesis typically proceeds in excellent yields and, like the Claisen rearrangement, is highly atom efficient. This also applies to the Diels–Alder reaction, which can be chemo-, regio-, diastereo- and enantioselective. It is important to note, however, that a substrate's suitability for these methods depends on its carbon skeleton and structure. Finally, photochemical reactions are often not the obvious first choice for many organic chemists, and as shown above, their use is again highly substrate-dependent. It can be possible to get complex mixtures of products, but on occasion high yields are achievable, as shown by several examples above.

More work is required to develop new and improved routes to these compounds and others in the literature that are yet to be synthesised. As highlighted above, some natural products with biological applications still require syntheses. Examples of compounds discovered in the last ten years with biological activity which are yet to be synthesised include biyouyanagiol,²⁰² spiromamakone A,²⁰³ and dibelmancandal A²⁰⁴ (Fig. 7).

Other natural products would benefit from more efficient routes, for example by the application of modern techniques to improve selectivity in older methods. Whilst progress over the last fifty years has been substantial, there is potential for further synthesis and exploitation of natural products containing spirocarbocycles.

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Notes and references

- G. Moss, *Pure Appl. Chem.*, 1999, **71**, 531–558.
- L. Hong and R. Wang, *Adv. Synth. Catal.*, 2013, **355**, 1023–1052.
- G. Singh and Z. Desta, *Chem. Rev.*, 2012, **112**, 6104–6155.
- A. Pfau and P. Plattner, *Helv. Chim. Acta*, 1939, **22**, 640–654.
- J. A. Marshall and P. Johnson, *J. Am. Chem. Soc.*, 1967, **89**, 2750–2751.
- P. Johnson and J. Marshall, *Chem. Commun.*, 1968, 391.
- Y. Zheng, C. M. Tice and S. B. Singh, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 3673–3682.
- K. Ding, *Chem. – Asian J.*, 2009, **4**, 32–41.
- K. W. Quasdorf and L. E. Overman, *Nature*, 2014, **516**, 181–191.
- M. Sannigrahi, *Tetrahedron*, 1999, **55**, 9007–9071.
- E. Corey and A. Guzman-Perez, *Angew. Chem., Int. Ed.*, 1998, **37**, 388–401.
- K. Fuji, *Chem. Rev.*, 1993, **93**, 2037–2066.
- R. Rios, *Chem. Soc. Rev.*, 2012, **41**, 1060–1074.
- J. Peng, M. A. Avery and M. T. Hamann, *Org. Lett.*, 2003, **5**, 4575–4578.
- J. Peng, K. Walsh, V. Weedman, J. D. Bergthold, J. Lynch, K. L. Lieu, I. A. Braude, M. Kelly and M. T. Hamann, *Tetrahedron*, 2002, **58**, 7809–7819.
- T. J. Reddy, G. Bordeau and L. Trimble, *Org. Lett.*, 2006, **8**, 5585–5588.
- B. R. Moser, *J. Nat. Prod.*, 2008, **71**, 487–491.
- D. F. Taber and K. V. Taluskie, *J. Org. Chem.*, 2006, **71**, 2797–2801.
- D. F. Taber, J.-M. Joerger and K. V. Taluskie, *Pure Appl. Chem.*, 2008, **80**, 1141–1148.
- D. F. Taber and J. Joerger, *J. Org. Chem.*, 2008, **73**, 4155–4159.
- B. Schetter and R. Mahrwald, *Angew. Chem., Int. Ed.*, 2006, **45**, 7506–7525.
- T. Machajewski and C. Wong, *Angew. Chem., Int. Ed.*, 2000, **39**, 1352–1375.
- G. Guella, F. Dini and F. Pietra, *Angew. Chem., Int. Ed.*, 1999, **38**, 1134–1136.
- K. C. Nicolaou, H. Zhang and A. Ortiz, *Angew. Chem., Int. Ed.*, 2009, **48**, 5642–5647.
- K. C. Nicolaou, M. P. Jennings and P. Dagneau, *Chem. Commun.*, 2002, 2480–2481.
- K. C. Nicolaou, W. Tang, P. Dagneau and R. Faraoni, *Angew. Chem., Int. Ed.*, 2005, **44**, 3874–3879.
- K. C. Nicolaou, H. Zhang, A. Ortiz and P. Dagneau, *Angew. Chem., Int. Ed.*, 2008, **47**, 8605–8610.
- K. C. Nicolaou, A. Ortiz and H. Zhang, *Angew. Chem., Int. Ed.*, 2009, **48**, 5648–5652.
- K. C. Nicolaou, A. Ortiz, H. Zhang, P. Dagneau, A. Lanver, M. P. Jennings, S. Arseniyadis, R. Faraoni and D. E. Lizos, *J. Am. Chem. Soc.*, 2010, **132**, 7138–7152.
- G. Saielli, K. C. Nicolaou, A. Ortiz, H. Zhang and A. Bagno, *J. Am. Chem. Soc.*, 2011, **133**, 6072–6077.
- J. Vrkoc, V. Herout and F. Sorm, *Collect. Czech Chem. Commun.*, 1961, **26**, 3183–3185.
- B. Tomita, T. Isono and Y. Hirose, *Tetrahedron Lett.*, 1970, **11**, 1371–1372.
- A. Srikrishna and R. Ramesh Babu, *Tetrahedron Lett.*, 2007, **48**, 6916–6919.
- D. McCrae and L. Dolby, *J. Org. Chem.*, 1977, **42**, 1607–1610.
- S. Martin and T. Chou, *J. Org. Chem.*, 1978, **43**, 1027–1031.
- A. Srikrishna, P. P. Kumar and R. Viswajanani, *Tetrahedron Lett.*, 1996, **37**, 1683–1686.
- A. Srikrishna and P. P. Kumar, *Tetrahedron*, 2000, **56**, 8189–8195.
- T. Tashiro, S. Kurosawa and K. Mori, *Biosci. Biotechnol. Biochem.*, 2004, **68**, 663–670.
- B. Trost, K. Hiroi and N. Holy, *J. Am. Chem. Soc.*, 1975, **97**, 5873–5877.
- I. Ahuja, R. Kissen and A. M. Bones, *Trends Plant Sci.*, 2012, **17**, 73–90.
- N. Andersen, M. Falcone and D. Syrdal, *Tetrahedron Lett.*, 1970, **11**, 1759–1762.
- E. Wenkert, B. L. Buckwalter, A. A. Craveiro, E. L. Sanchez and S. S. Sathe, *J. Am. Chem. Soc.*, 1978, **100**, 1267–1273.
- R. Hutchins, N. Natele, I. Taffer and R. Zipkin, *Synth. Commun.*, 1984, **14**, 445–451.
- M. L. Maheshwari, S. C. Bhattacharyya and K. Varma, *Tetrahedron*, 1965, **21**, 115–138.
- C. Hughes, M. Deighton and R. Ramage, *J. Chem. Soc., Chem. Commun.*, 1975, 662–663.
- G. Balme, *Tetrahedron Lett.*, 1985, **26**, 2309–2310.
- D. Coxon, K. Price and B. Howard, *Tetrahedron Lett.*, 1974, **15**, 2921–2924.
- A. Srikrishna and S. S. V. Ramasastry, *Tetrahedron Lett.*, 2005, **46**, 7373–7376.
- Y. Tokunaga, M. Yagihashi, M. Ihara and K. Fukumoto, *J. Chem. Soc., Perkin Trans. 1*, 1997, 189–190.
- H. Huang and C. J. Forsyth, *Tetrahedron Lett.*, 1993, **34**, 7889–7890.
- H. Huang and C. J. Forsyth, *J. Org. Chem.*, 1995, **60**, 2773–2779.
- W. Oppolzer and F. Flachsmann, *Helv. Chim. Acta*, 2001, **84**, 416–430.
- M. Lachia, F. Dénès, F. Beaufils and P. Renaud, *Org. Lett.*, 2005, **7**, 4103–4106.
- A. Matsuo, S. Uto, H. Nozaki and M. Nakayama, *J. Chem. Soc., Perkin Trans. 1*, 1984, 215–221.
- M. Kodama, U. Tambunan, T. Tsunoda and S. Ito, *Bull. Chem. Soc. Jpn.*, 1986, **59**, 1897–1900.
- E. Boeren, M. El-Sohly, C. Turner and A. Salemin, *Experientia*, 1976, **33**, 848.
- C. Bercht, J. van Dongen, W. Heerma, R. Lousberg and F. Kueppers, *Tetrahedron*, 1976, **32**, 2939–2943.
- T. Ottersen, A. Aasen, F. S. El-Ferally and C. Turner, *J. Chem. Soc., Chem. Commun.*, 1976, 580–581.



- 59 F. S. El-Feraly, M. Elsohly, E. Boeren and C. Turner, *Tetrahedron*, 1977, **33**, 2373–2378.
- 60 Y. Shoyama and I. Nishioka, *Chem. Pharm. Bull.*, 1978, **26**, 3641–3646.
- 61 L. Crombie, W. Crombie and S. Jamieson, *Tetrahedron Lett.*, 1979, **20**, 661–664.
- 62 J. Kettenes-van den Bosch and C. Salemink, *Recl. J. R. Neth. Chem. Soc.*, 1978, **97**, 221–222.
- 63 L. Crombie, P. Tuchinda and M. J. Powell, *J. Chem. Soc., Perkin Trans. 1*, 1982, 1477–1484.
- 64 L. Crombie, M. Powell and P. Tuchinda, *Tetrahedron Lett.*, 1980, **21**, 3603–3606.
- 65 F. El-Feraly and Y. Chan, *J. Nat. Prod.*, 1981, **44**, 557–561.
- 66 N. Natale, B. Marron, E. Evain and C. Dodson, *Synth. Commun.*, 1984, **14**, 599–603.
- 67 J. Novak and C. A. Salemink, *J. Chem. Soc., Perkin Trans. 1*, 1982, 2403–2405.
- 68 M. Suzuki, E. Kurosawa and K. Kurata, *Bull. Chem. Soc. Jpn.*, 1987, **60**, 3795–3796.
- 69 D. Kennedy, I. Selby and R. Thomson, *Phytochemistry*, 1988, **27**, 1761–1766.
- 70 K.-C. Chu, H.-J. Liu and J.-L. Zhu, *Synlett*, 2010, 3061–3064.
- 71 J. Zhu, P. Huang, R. You, F. Lee, S. Taso and I. Chen, *Synthesis*, 2011, 715–722.
- 72 H. Wolf and M. Kolleck, *Tetrahedron Lett.*, 1975, **16**, 451–454.
- 73 J. Ruppert, M. Avery and J. White, *J. Chem. Soc., Chem. Commun.*, 1976, 978.
- 74 M. Pesaro and J.-P. Bachmann, *J. Chem. Soc., Chem. Commun.*, 1978, 203–204.
- 75 S. Nagumo, H. Suemune and K. Sakai, *J. Chem. Soc., Chem. Commun.*, 1990, 1778–1779.
- 76 X. D. Yue, L. Chen and W. D. Z. Li, *Tetrahedron*, 2014, **70**, 5505–5512.
- 77 K. Aoki, K. Yamada, H. Nagase, Y. Hayakawa and Y. Hirata, *Tetrahedron Lett.*, 1973, **14**, 4967–4970.
- 78 K. Yamada, H. Nagase, Y. Hayakawa, K. Aoki and Y. Hirata, *Tetrahedron Lett.*, 1973, **14**, 4963–4966.
- 79 P. McCurry and R. Singh, *Tetrahedron Lett.*, 1973, **14**, 3325–3328.
- 80 M. Mongrain, J. Lafontaine, A. Bélanger and P. Deslongchamps, *Can. J. Chem.*, 1970, **48**, 3273–3274.
- 81 J. Lafontaine, M. Mongrain, M. Sergent-Guay, L. Ruest and P. Deslongchamps, *Can. J. Chem.*, 1980, **58**, 2460–2476.
- 82 G. Stork, R. Danheiser and B. Ganem, *J. Am. Chem. Soc.*, 1973, **95**, 3414–3415.
- 83 N. Maulide, J.-C. Vanherck and I. E. Markó, *Eur. J. Org. Chem.*, 2004, 3962–3967.
- 84 G. Posner and T. Hamill, *J. Org. Chem.*, 1988, **53**, 6031–6035.
- 85 A. Murai, S. Sato and T. Masamune, *Bull. Chem. Soc. Jpn.*, 1984, **57**, 2276–2281.
- 86 S. N. Janaki and G. S. R. Subba Rao, *J. Chem. Soc., Perkin Trans. 1*, 1997, 195–200.
- 87 J. A. Marshall and S. F. Brady, *J. Org. Chem.*, 1970, **35**, 4068–4077.
- 88 T. Ito and T. Takahashi, *Agric. Biol. Chem.*, 1979, **43**, 413–414.
- 89 T. Fujimori, R. Uegaki, Y. Takagi, S. Kubo and K. Kato, *Phytochemistry*, 1979, **18**, 2032.
- 90 R. Uegaki, T. Fujimori, S. Kubo and K. Kato, *Phytochemistry*, 1981, **20**, 1567–1568.
- 91 R. Uegaki, S. Kubo and T. Fujimori, *Phytochemistry*, 1988, **27**, 365–368.
- 92 G. Birnbaum, C. Huber and M. Post, *J. Chem. Soc., Chem. Commun.*, 1976, 330–331.
- 93 A. Stoessl, J. Stothers and E. Ward, *Can. J. Chem.*, 1978, **56**, 645–653.
- 94 A. Murai, S. Sato and T. Masamune, *Bull. Chem. Soc. Jpn.*, 1984, **57**, 2286–2290.
- 95 A. Murai, S. Sato and T. Masamune, *Bull. Chem. Soc. Jpn.*, 1984, **57**, 2291–2294.
- 96 G. Blay, L. Cardona, A. M. Collado, B. García, V. Morcillo and J. R. Pedro, *J. Org. Chem.*, 2004, **69**, 7294–7302.
- 97 H. Magari, H. Hirota, T. Takahashi, A. Matsuo, S. Uto, H. Nozaki, M. Nakayama and S. Hayashi, *Chem. Lett.*, 1982, 1143–1146.
- 98 M. Suzuki, N. Kowata and E. Kurosawa, *Tetrahedron*, 1980, **36**, 1551–1556.
- 99 M. Suzuki, E. Kurosawa and T. Irie, *Tetrahedron Lett.*, 1970, **11**, 4995–4998.
- 100 A. Murai, K. Kato and T. Masamune, *Tetrahedron Lett.*, 1982, **23**, 2887–2890.
- 101 J. Plamondon and P. Canonne, *Tetrahedron Lett.*, 1991, **32**, 589–592.
- 102 D. F. Taber, M. I. Sikkander and P. H. Storck, *J. Org. Chem.*, 2007, **72**, 4098–4101.
- 103 G. Chianese, E. Fattorusso, O. O. Aiyelaagbe, P. Luciano, H. C. Schröder, W. E. G. Müller and O. Taglialatela-Scafati, *Org. Lett.*, 2011, **13**, 316–319.
- 104 X. Li, Y. Yang, X. Peng, M. Li, L. Li, X. Deng, H. Qin, J. Liu and M. Qiu, *Org. Lett.*, 2014, **16**, 2196–2199.
- 105 V. A. D'yakonov, O. A. Trapeznikova, A. de Meijere and U. M. Dzhemilev, *Chem. Rev.*, 2014, **114**, 5775–5814.
- 106 K. C. Nicolaou, P. G. Bulger and D. Sarlah, *Angew. Chem., Int. Ed.*, 2005, **44**, 4490–4527.
- 107 A. Gradillas and J. Pérez-Castells, *Angew. Chem., Int. Ed.*, 2006, **45**, 6086–6101.
- 108 S. Kotha and E. Manivannan, *ARKIVOC*, 2003, 67–76.
- 109 M. Anchel, A. Hervey and W. Robbins, *Proc. Natl. Acad. Sci. U. S. A.*, 1950, **36**, 300–305.
- 110 T. McMorris and M. Anchel, *J. Am. Chem. Soc.*, 1963, **85**, 831–832.
- 111 T. McMorris and M. Anchel, *J. Am. Chem. Soc.*, 1965, **87**, 1594–1600.
- 112 M. Kelner, T. McMorris, W. Beck, J. Zamora and R. Taetle, *Cancer Res.*, 1987, **47**, 3186–3189.



- 113 T. McMorris, M. Kelner, W. Wang, L. Estes, M. Montoya and R. Taetle, *J. Org. Chem.*, 1992, **57**, 6876–6883.
- 114 F. Kinder, R. Wang, W. Bauta and K. Bair, *Bioorg. Med. Chem. Lett.*, 1996, **6**, 1029–1034.
- 115 M. Movassaghi, G. Piizzi, D. S. Siegel and G. Piersanti, *Angew. Chem., Int. Ed.*, 2006, **45**, 5859–5863.
- 116 D. S. Siegel, G. Piizzi, G. Piersanti and M. Movassaghi, *J. Org. Chem.*, 2009, **74**, 9292–9304.
- 117 T. C. McMorris, M. D. Staake and M. J. Kelner, *J. Org. Chem.*, 2004, **69**, 619–623.
- 118 M. Movassaghi, G. Piizzi, D. S. Siegel and G. Piersanti, *Tetrahedron Lett.*, 2009, **50**, 5489–5492.
- 119 T. Matsumoto, H. Shirahama, A. Ichihara, H. Shin, S. Kagaqa, F. Sakan, S. Matsumoto and S. Nishida, *J. Am. Chem. Soc.*, 1968, **90**, 3280–3281.
- 120 T. Matsumoto, H. Shirahama, A. Ichihara, H. Shin, S. Kagawa, F. Sakan and K. Miyano, *Tetrahedron Lett.*, 1971, 2049–2052.
- 121 Q. Huang, Y. Tezuka, Y. Hatanaka, T. Kikuchi, A. Nishi and K. Tubaki, *Chem. Pharm. Bull.*, 1995, **43**, 1035–1038.
- 122 N. L. Brock and J. S. Dickschat, *Eur. J. Org. Chem.*, 2011, 5167–5175.
- 123 A. Srikrishna and M. Rao, *Indian J. Chem., Sect. B*, 2008, **47**, 1423–1429.
- 124 S. Kurosawa, M. Bando and K. Mori, *Eur. J. Org. Chem.*, 2001, 4395–4399.
- 125 K. Oesterreich, I. Klein and D. Spitzner, *Synlett*, 2002, 1712–1714.
- 126 K. Oesterreich and D. Spitzner, *Tetrahedron*, 2002, **58**, 4331–4334.
- 127 E. J. Corey and S. Nozoe, *J. Am. Chem. Soc.*, 1965, **87**, 5728–5733.
- 128 T. Kauffmann, *Angew. Chem., Int. Ed.*, 1997, **36**, 1258–1275.
- 129 T. Ozaki and Y. Kobayashi, *Synlett*, 2015, 1085–1088.
- 130 H. Abe, A. Sato, T. Kobayashi and H. Ito, *Org. Lett.*, 2013, **15**, 1298–1301.
- 131 Y.-M. Zhang, N.-H. Tan, M. He, Y. Lu, S.-Q. Shang and Q.-T. Zheng, *Tetrahedron Lett.*, 2004, **45**, 4319–4321.
- 132 S. Maity and S. Ghosh, *Tetrahedron Lett.*, 2007, **48**, 3355–3358.
- 133 I. Granado and P. Caballero, *Sci. Mar.*, 1995, **59**, 21–39.
- 134 J. D. Martin, C. Perez and J. L. Ravelo, *J. Am. Chem. Soc.*, 1986, **108**, 7801–7811.
- 135 C. S. Vairappan, M. Daitoh, M. Suzuki, T. Abe and M. Masuda, *Phytochemistry*, 2001, **58**, 291–297.
- 136 C. S. Vairappan, *Biomol. Eng.*, 2003, **20**, 255–259.
- 137 G. M. König and A. D. Wright, *J. Nat. Prod.*, 1997, **60**, 967–970.
- 138 T. Dias, I. Brito, L. Moujir, N. Paiz, J. Darias and M. Cueto, *J. Nat. Prod.*, 2005, **68**, 1677–1679.
- 139 Y. Ohta and Y. Hirose, *Tetrahedron Lett.*, 1968, 2483–2485.
- 140 A. Srikrishna, B. V. Lakshmi and M. Mathews, *Tetrahedron Lett.*, 2006, **47**, 2103–2106.
- 141 A. Srikrishna and R. Ramesh Babu, *Tetrahedron*, 2008, **64**, 10501–10506.
- 142 D. E. White, I. C. Stewart, R. H. Grubbs and B. M. Stoltz, *J. Am. Chem. Soc.*, 2008, **130**, 810–811.
- 143 D. E. White, I. C. Stewart, B. A. Seashore-Ludlow, R. H. Grubbs and B. M. Stoltz, *Tetrahedron*, 2010, **66**, 4668–4686.
- 144 K. C. Nicolaou, S. A. Snyder, T. Montagnon and G. Vassilikogiannakis, *Angew. Chem., Int. Ed.*, 2002, **41**, 1668–1698.
- 145 N. H. Andersen and D. D. Syrdal, *Tetrahedron Lett.*, 1970, **26**, 2277–2280.
- 146 N. Andersen and D. Syrdal, *Tetrahedron Lett.*, 1972, **10**, 899–902.
- 147 J. N. Marx and L. R. Norman, *Tetrahedron Lett.*, 1973, 4375–4378.
- 148 J. N. Marx and L. Norman, *J. Org. Chem.*, 1975, **40**, 1602–1606.
- 149 A. Aoyagi, M. Ito-kobayashi, Y. Ono, Y. Furukawa, M. Takahashi, Y. Muramatsu, M. Umetani and T. Takatsu, *J. Antibiot.*, 2008, **61**, 136–141.
- 150 T. Ling, E. Griffith, K. Mitachi and F. Rivas, *Org. Lett.*, 2013, **15**, 5790–5793.
- 151 T. Sawada and M. Nakada, *Org. Lett.*, 2013, **15**, 1004–1007.
- 152 T. Sawada and M. Nakada, *Org. Lett.*, 2014, **16**, 1537.
- 153 C. Li, X. Yu and X. Lei, *Org. Lett.*, 2010, **12**, 4284–4287.
- 154 D. Xia, Y. Du, Z. Yi, H. Song and Y. Qin, *Chem. – Eur. J.*, 2013, **19**, 4423–4427.
- 155 F. E. Ziegler, *Acc. Chem. Res.*, 1977, **10**, 227–232.
- 156 A. M. M. Castro, *Chem. Rev.*, 2004, **104**, 2939–3002.
- 157 A. Nakazaki and S. Kobayashi, *Synlett*, 2012, 1427–1445.
- 158 I. Yosioka, H. Hikino and Y. Sasaki, *Chem. Pharm. Bull.*, 1961, **9**, 84–85.
- 159 I. Yosioka and T. Kimura, *Chem. Pharm. Bull.*, 1965, **13**, 1430–1434.
- 160 A. Nakazaki and S. Kobayashi, *Chem. Lett.*, 2007, **36**, 42–43.
- 161 A. Nakazaki, T. Era, Y. Numada and S. Kobayashi, *Tetrahedron*, 2006, **62**, 6264–6271.
- 162 T. Okino, E. Yoshimura, H. Hirota and N. Fusetani, *Tetrahedron*, 1996, **52**, 9447–9454.
- 163 H. Prawat, C. Mahidol, S. Wittayalai, P. Intachote, T. Kanchanapoom and S. Ruchirawat, *Tetrahedron*, 2011, **67**, 5651–5655.
- 164 A. Wright, H. Wang, M. Gurrath, G. M. König, G. Kocak, G. Neumann, P. Loria, M. Foley and L. Tilley, *J. Med. Chem.*, 2001, **44**, 873–885.
- 165 C. Angerhoffer and J. Pezzuto, *J. Nat. Prod.*, 1992, **55**, 1787–1789.
- 166 A. Nakazaki, H. Miyamoto, K. Henmi and S. Kobayashi, *Synlett*, 2005, 1417–1420.
- 167 A. Nakazaki, T. Era and S. Kobayashi, *Chem. Pharm. Bull.*, 2007, **55**, 1606–1609.
- 168 K. Tamura, A. Nakazaki and S. Kobayashi, *Synlett*, 2009, 2449–2452.
- 169 T. Sattelkau and P. Eilbracht, *Tetrahedron Lett.*, 1998, **39**, 9647–9648.
- 170 T. Sattelkau, C. Hollmann and P. Eilbracht, *Synlett*, 1996, 1221–1223.



- 171 T. Sattelkau and P. Eilbracht, *Tetrahedron Lett.*, 1998, **39**, 1905–1908.
- 172 P. Eilbracht, A. Gersmeier, D. Lennartz and T. Huber, *Synthesis*, 1995, 330–334.
- 173 *Handbook of Synthetic Photochemistry*, ed. A. Albini and M. Fagnoni, 2010.
- 174 T. Bach and J. P. Hehn, *Angew. Chem., Int. Ed.*, 2011, **50**, 1000–1045.
- 175 D. Ravelli, M. Fagnoni and A. Albini, *Chem. Soc. Rev.*, 2013, **42**, 97–113.
- 176 B.-W. Yu, J.-Y. Chen, Y.-P. Wang, K.-F. Cheng, X.-Y. Li and G.-W. Qin, *Phytochemistry*, 2002, **61**, 439–442.
- 177 F. Li, S. S. Tartakoff and S. L. Castle, *J. Org. Chem.*, 2009, **74**, 9082–9093.
- 178 R. Navarro and S. E. Reisman, *Org. Lett.*, 2012, **14**, 4354–4357.
- 179 S. M. King, N. A. Calandra and S. B. Herzon, *Angew. Chem., Int. Ed.*, 2013, **52**, 3642–3645.
- 180 G. Guest, C. R. Hughes, A. Sattar and R. Ramage, *J. Chem. Soc., Chem. Commun.*, 1973, 526–527.
- 181 S. Baldwin and J. Fredericks, *Tetrahedron Lett.*, 1982, **23**, 1235–1238.
- 182 W. Oppolzer, F. Zutterman and K. Baettig, *Helv. Chim. Acta*, 1983, **66**, 522–533.
- 183 A. Stoessl and E. Ward, *Tetrahedron Lett.*, 1976, **17**, 3271–3274.
- 184 N. Katsui, A. Matsunaga, H. Kitahara, F. Yagihashi, A. Murai, T. Masamune and N. Sato, *Bull. Chem. Soc. Jpn.*, 1977, **50**, 1217–1225.
- 185 Y. Kawaguchi, T. Ochi, Y. Takaishi, K. Kawazoe and K.-H. Lee, *J. Nat. Prod.*, 2004, **67**, 1893–1896.
- 186 M. T. Crimmins, C. M. Dudek and A. W. Cheung, *Tetrahedron Lett.*, 1992, **33**, 181–184.
- 187 M. T. Crimmins, Z. Wang and L. A. McKelvie, *Tetrahedron Lett.*, 1996, **37**, 8703–8706.
- 188 M. T. Crimmins, Z. Wang and L. A. McKelvie, *J. Am. Chem. Soc.*, 1998, **120**, 1747–1756.
- 189 T. Hatsui, J. Wang and H. Takeshita, *Bull. Chem. Soc. Jpn.*, 1995, **68**, 2393–2399.
- 190 D. Caine, A. Boucugnani, S. Chao, J. Byron Dawson and P. Ingwalson, *J. Org. Chem.*, 1976, **41**, 1539–1544.
- 191 Z. Horii, C. Iwata and Y. Nakashita, *Chem. Pharm. Bull.*, 1978, **26**, 481–483.
- 192 Z. Horii, Y. Nakashita and C. Iwata, *Tetrahedron Lett.*, 1971, **12**, 1167–1168.
- 193 T. Kametani, K. Fukumoto, S. Shibuya, H. Nemoto, T. Nakano, T. Sugahara, T. Takahashi, Y. Aizawa and M. Toriyama, *J. Chem. Soc., Perkin Trans. 1*, 1972, 1435–1441.
- 194 T. Kametani, T. Sugahara, H. Sugi, S. Shibuya and K. Fukumoto, *Tetrahedron*, 1971, **27**, 5993–5998.
- 195 T. Kametani, T. Sugahara, H. Sugi, S. Shibuya and K. Fukumoto, *J. Chem. Soc., Chem. Commun.*, 1971, 724.
- 196 K. Bernauer, *Experientia*, 1964, **15**, 380–381.
- 197 T. Honda and H. Shigehisa, *Org. Lett.*, 2006, **8**, 657–659.
- 198 F. Wang, Y. Gao, L. Zhang and J.-K. Liu, *Org. Lett.*, 2010, **12**, 2354–2357.
- 199 F. Wang, Y. Gao, L. Zhang, B. Bai, Y. Hu, Z. Donh, Q. Zhai, H. Zhu and J. Liu, *Org. Lett.*, 2010, **12**, 3196–3199.
- 200 H. Tan, C. Zheng, Z. Liu and D. Z. Wang, *Org. Lett.*, 2011, **13**, 2192–2195.
- 201 X. Hu, Y. Wang, F. Xiao, W. Liu, Q. Zhang and X. Li, *Asian J. Org. Chem.*, 2013, **2**, 216–219.
- 202 N. Tanaka, Y. Kashiwada, S. Y. Kim, W. Hashida, M. Sekiya, Y. Ikeshiro and Y. Takaishi, *J. Nat. Prod.*, 2009, **72**, 1447–1452.
- 203 S. A. Van Der Sar, J. W. Blunt and M. H. G. Munro, *Org. Lett.*, 2006, **8**, 2059–2061.
- 204 Z. Song, X. Xu, W. Deng and S. Peng, *Org. Lett.*, 2011, **13**, 462–465.

