

TECHNIQUES FOR PHYSIOLOGY

Pressure-dependent regulation of Ca²⁺ signalling in the vascular endothelium

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Key points

- Increased pressure suppresses endothelial control of vascular tone but it remains uncertain (1) how pressure is sensed by the endothelium and (2) how the vascular response is inhibited.
- This study used a novel imaging method to study large numbers of endothelial cells in arteries that were in a physiological configuration and held at normal blood pressures.
- Increased pressure suppressed endothelial IP₃-mediated Ca²⁺ signals.
- Pressure modulated endothelial cell shape.
- The changes in cell shape may alter endothelial Ca²⁺ signals by modulating the diffusive environment for Ca²⁺ near IP₃ receptors.
- Endothelial pressure-dependent mechanosensing may occur without a requirement for a conventional molecular mechanoreceptor.

Abstract The endothelium is an interconnected network upon which haemodynamic mechanical forces act to control vascular tone and remodelling in disease. Ca²⁺ signalling is central to the endothelium's mechanotransduction and networked activity. However, challenges in imaging Ca²⁺ in large numbers of endothelial cells under conditions that preserve the intact physical configuration of pressurized arteries have limited progress in understanding how pressure-dependent mechanical forces alter networked Ca²⁺ signalling. We developed a miniature wide-field, gradient-index (GRIN) optical probe designed to fit inside an intact pressurized artery that permitted Ca²⁺ signals to be imaged with subcellular resolution in a large number (~200) of naturally connected endothelial cells at various pressures. Chemical (acetylcholine) activation triggered spatiotemporally complex, propagating inositol trisphosphate (IP₃)-mediated Ca²⁺ waves that originated in clusters of cells and progressed from there across the endothelium. Mechanical stimulation of the artery, by increased intraluminal pressure, flattened the endothelial cells and suppressed IP₃-mediated Ca²⁺ signals in all activated cells. By computationally modelling Ca²⁺ release, endothelial shape changes were shown to alter the geometry of the Ca²⁺ diffusive environment near IP₃ receptor microdomains to limit IP₃-mediated Ca²⁺ signals as pressure increased. Changes in cell shape produce a geometric microdomain regulation of IP₃-mediated Ca²⁺ signalling to explain macroscopic pressure-dependent, endothelial mechanosensing without the need for a conventional mechanoreceptor. The suppression of IP₃-mediated Ca²⁺ signalling may explain the decrease in endothelial activity as pressure increases. GRIN imaging provides a convenient method that gives access to hundreds of endothelial cells in intact arteries in physiological configuration.

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Abbreviations $[Ca^{2+}]_c$, cytosolic Ca^{2+} concentration; GRIN, gradient index; IP_3 , inositol trisphosphate; IP_3R , inositol trisphosphate receptor; RyR, ryanodine receptor; TRP, transient receptor potential.

Introduction

The vascular endothelium is a one-cell-thick layer that directs the formation of new blood vessels (angiogenesis), prevents blood clotting, regulates vascular permeability, controls arterial tone and determines the extent of smooth muscle proliferation. The endothelium's control of each of these functions arises from the cells acting as a sensitive signal processing centre that detects and interprets multiple simultaneous messages such as those derived from mechanical stimuli (hydrostatic pressure, luminal shear stress, circumferential strain) and local and blood borne signals (autocrine, paracrine and electrical signals and neurotransmitters). Endothelial stimuli are transduced to changes in the endothelial Ca^{2+} concentration to coordinate the endothelium's control of vascular activity (Behringer & Segal, 2012; Sonkusare *et al.* 2012; Billaud *et al.* 2014). Ca^{2+} regulates the synthesis and release of various vasoactive agents such as nitric oxide, prostacyclin and endothelium-derived hyperpolarizing factor. Through these Ca^{2+} -dependent mediators the endothelium's control of vascular contraction, permeability, cell proliferation and angiogenesis is achieved. Therefore, central to an understanding of endothelial signal processing is an appreciation of the control of Ca^{2+} .

Two main sources of endothelial Ca^{2+} are recognized, the extracellular fluid and the intracellular stores of the endoplasmic reticulum (Moccia *et al.* 2012). Ca^{2+} entry from the extracellular fluid may occur via a large number of ion channels on the outside (plasma) membrane. The other main cytoplasmic Ca^{2+} concentration ($[Ca^{2+}]_c$) source is the internal endoplasmic reticulum store from which release proceeds mainly via the inositol 1,4,5-trisphosphate (IP_3) receptor (IP_3R). While Ca^{2+} release via IP_3R is well established the contribution of the ryanodine receptor (RyR) to the control of endothelial Ca^{2+} (if any) is less clear (Socha *et al.* 2012a). The sources of Ca^{2+} are not independent: Ca^{2+} influx regulates Ca^{2+} release and Ca^{2+} release regulates Ca^{2+} influx. For example, Ca^{2+} release from the endoplasmic reticulum may alter the activity of ion channels present on the plasma membrane to regulate Ca^{2+} entry either via Ca^{2+} -gated ion channels (Strotmann *et al.* 2003) or via membrane potential changes altering passive fluxes of the ion (Behringer & Segal, 2015). Alternatively Ca^{2+} influx may alter endoplasmic reticulum Ca^{2+} content or activity of IP_3R in a Ca^{2+} -induced Ca^{2+} release-like process (Earley & Brayden, 2015). Thus the local change in

$[Ca^{2+}]_c$ arising from the activity of channels in the plasma membrane or endoplasmic reticulum itself regulates the activity of ion channels to provide a feedback control of Ca^{2+} signals and modulate vascular function.

The response to mechanical stimuli involves changes in $[Ca^{2+}]_c$ and must be integrated with signals from other sensors to converge on a physiological response. Two major mechanical stimuli are shear stress and pressure (Falcone *et al.* 1993; Huang *et al.* 1998; Popp *et al.* 1998; Muller *et al.* 1999; Marchenko & Sage, 2000; Paniagua *et al.* 2000; Sun *et al.* 2001; Duza & Sarelius, 2004). The endothelial response to shear stress is well characterized and several types of activity may occur. Vascular smooth muscle relaxation may be evoked, cell migration induced and endothelial gene expression changed (Falcone *et al.* 1993; Muller *et al.* 1999; Shiu *et al.* 2004; Chien, 2007). The increases in endothelial Ca^{2+} changes underlying these responses may involve several types of mechanically sensitive ion channels forces such as Piezo1 (Li *et al.* 2014; Ranade *et al.* 2014), ENaC (Kusche-Vihrog *et al.* 2014), ATP-gated P2X4 (Yamamoto *et al.* 2006), TREK-1 (Dedman *et al.* 2009) and various members of the TRP grouping of channels (Corey *et al.* 2004; Maroto *et al.* 2005; Spassova *et al.* 2006; Janssen *et al.* 2011). Alternatively, shear stress-evoked activation of ion channels may be indirect and force sensed by the cytoskeleton, apical glycocalyx, mechanosensitive or membrane curvature-sensitive protein complexes or G-protein-coupled receptors (Knudsen & Frangos, 1997; Thi *et al.* 2004; Tzima *et al.* 2005; Zimmerberg & Kozlov, 2006; Mederos y Schnitzler *et al.* 2008; Zhao *et al.* 2011). The diversity of responses highlights the complexity of the response to shear stress.

The response of the endothelium to pressure differs significantly from that of shear stress. Rather than being activated, a decrease in activity may occur as mechanical stimulation (pressure) is increased (Hishikawa *et al.* 1992; Gunduz *et al.* 2008). The decrease in endothelial activity may suppress smooth muscle relaxation (De Bruyn *et al.* 1994; Huang *et al.* 1998; Paniagua *et al.* 2000; Zhao *et al.* 2015). In healthy human volunteers, short-term increases in arterial blood pressure cause long-lasting inhibition of endothelium-dependent dilatation (Jurva *et al.* 2006; Phillips *et al.* 2011). In isolated arteries, acute exposure to increases in transmural pressure may impair endothelium-dependent relaxation (Hishikawa *et al.* 1992; Huang *et al.* 1998; Paniagua *et al.* 2000; Zhao *et al.* 2015). Relatively little is known on how pressure is sensed

by the endothelium, specifically when the arteries are in their natural 'tubular' configuration.

The biological response of arteries to pressure depends critically on the complex, cylindrical, three-dimensional arrangement of cells and on the interactions with other cell types, and may change with artery configuration. For example, in many intact arteries in their cylindrical configuration, myogenic contraction occurs when the artery is subjected to circumferential stretch by increases in pressure. However, this contraction does not occur in arteries stretched, with equivalent forces, on wires (Dunn *et al.* 1994). Increased mechanical stimulation by stretch, hypotonic cell swelling or shear stress activates TRPV4 channels (Strotmann *et al.* 2000; Alessandri-Haber *et al.* 2003; Loukin *et al.* 2010), a mechanism that may contribute to flow-induced dilatation (Kohler *et al.* 2006; Mendoza *et al.* 2010; Bubolz *et al.* 2012). However, TRPV4 channels are also deactivated by increased stretch when transmural pressure difference is increased in pressurized arteries (Bagher *et al.* 2012) in a cylindrical configuration. The cell shape and operative mechanical forces in a pressurized artery are quite different from experiments on isolated or cultured cells.

The carotid artery contributes to the control of cerebrovascular blood flow and cerebral vascular resistance (Mchedlishvili, 1986; Faraci & Heistad, 1990). The endothelium regulates carotid artery tone. Vascular relaxation is evoked by several endothelial activators including flow (shear stress), adrenomedullin and acetylcholine (Faraci *et al.* 1994; Plane *et al.* 1998; Chataigneau *et al.* 1998*a,b*; Ohashi *et al.* 2005) and the endothelium may also attenuate the vascular contractile response to vasoconstrictors (Lamping & Faraci, 2001). Many of the most serious forms of cardiovascular diseases (e.g. atherosclerosis) reside in larger arteries like the carotid artery and begin with endothelial dysfunction (Deanfield *et al.* 2007). However, studying the effects of mechanical forces like pressure on endothelial function (and dysfunction) in larger arteries in a physiological configuration has been exceptionally difficult. Assessment of endothelial function in large arteries has been largely indirect. The majority of papers in the past decade involved only the measurement of endothelium-dependent dilatation (e.g. Craig & Martin, 2012).

To study the function of endothelial cells in intact arteries, some investigations have used either wide-field or point-scanning fluorescence microscopes to visualize the endothelium through the wall of the artery (Bagher *et al.* 2012; Sonkusare *et al.* 2012; Tran *et al.* 2012). However, movement of the artery in pressure myograph systems is almost unavoidable and results in the vessel moving in and out of the focal plane, changing light levels and altering image quality to present a significant challenge to data analysis. Light scattering by the artery wall also reduces contrast and the curvature of the artery limits

the number of cells that may be visualized within a single optical plane. To overcome each of these difficulties and allow the endothelium to be studied in intact arteries, we have developed a miniature optical probe to record Ca²⁺ signalling from *inside* pressurized arteries. The probe has a field of view of 0.5 mm diameter, which allowed a large number (~200) of naturally connected endothelial cells to be imaged with subcellular resolution and has a high depth of field (141 μm) sufficient to maintain good focus across the highly curved, intact endothelial layer of a large artery. To handle the data from such a large number of cells a largely automated image processing routine was also developed. We show in native endothelial cells in their physiological configuration, that acetylcholine-evoked Ca²⁺ rises originate as IP₃-mediated Ca²⁺ signals in particular regions of the endothelium from which they progress to other cells as Ca²⁺ waves. Detection of pressure-dependent mechanical forces by the endothelium is integrated effortlessly into the same signalling pathway by geometric modulation of IP₃-evoked Ca²⁺ release brought about by changes in endothelial cell shape. We suggest the suppression of IP₃-mediated Ca²⁺ signals may underlie the inhibition of endothelial responses with increased pressure and, significantly, may not require a conventional mechanosensor for mechanotransduction to occur.

Methods

Ethical approval

All experiments employed tissue obtained from male Sprague–Dawley rats (10–12 weeks old; 250–350 g). Rats were humanely killed by overdose of pentobarbital sodium with the approval of the University of Strathclyde Local Ethical Review Panel (200 mg kg⁻¹; Schedule 1 procedure; Animals (Scientific Procedures) Act 1986, UK), under UK Home Office Project and Personal Licence authority.

Tissue preparation

The left and right common carotid arteries were exposed by blunt dissection. To prevent collapse of the artery upon removal, the rostral and caudal ends of the exposed carotid arteries were ligated with 8-0 suture and the arteries were then rapidly excised. Arteries were then cleaned of connective tissue under a dissection microscope, and visually checked for the presence of side branches. Subsequently, arteries without side branches were mounted onto two blunted and deburred 22-gauge cannula in a custom-designed imaging bath using two lengths of suture thread. Blood was removed from the arteries by flushing the lumen with physiological saline solution (PSS) for 10 min (150 μl min⁻¹) before the arteries were pressurized to 60 mmHg, gently heated to 37°C

and allowed to equilibrate at 37°C for 30 min. Flow was provided by a peristaltic pump (PS-200, Living Systems Instrumentation, At Albans City, VT, USA) connected to the proximal cannula. The endothelium of mounted arteries was then selectively loaded with a fluorescent Ca^{2+} indicator by intraluminal perfusion of PSS containing the membrane-permeant form of Oregon Green BAPTA-1 (20 μM , OGB-1/AM) and 0.04% Pluronic acid. Flow was stopped for 30 min to permit sufficient loading. Arteries were continuously superfused with PSS during this time. Following removal of excess dye, the distal cannula was removed from the artery and the artery was mounted on a gradient index (GRIN) microprobe (described below) and secured with suture thread. Transmural pressure was then incrementally increased to 160 mmHg, whilst stretching the artery to remove any buckle. Following this procedure, the pressure was decreased to 60 mmHg and the artery left for a further 30 min to equilibrate. During equilibration arteries were tested for leaks (which may indicate side branches, tears in the vessel wall or insufficiently tied sutures) by switching off the feedback on the pressure servo. Arteries that showed signs of leakage, identified as a reduction in pressure whilst isolated from the servo system, which could not be stopped by re-tying the sutures were discarded at this point.

Micro-endoscope GRIN imaging probe

The GRIN microprobe was developed to fit inside pressurized arteries (Fig. 1A) and consisted of a 0.5 mm diameter, 30.2 mm long, single pitch GRIN relay lens (SRL-050; Nippon Sheet Glass, USA) with a 0.5 mm \times 0.5 mm \times 0.5 mm aluminium-coated micro-prism (66-771; Edmund Optics, USA) attached to the distal surface with ultraviolet curing optical epoxy (NAO 68; Norland Products, USA). The lens was sheathed in a surgical stainless steel tube (0.71 mm outer diameter) for mechanical protection. In this single lens configuration, the GRIN rod acts to re-conjugate the image plane of detection optics through the length of the cylinder (Fig. 1D). By focusing a conventional microscope objective to a sufficient depth inside the GRIN rod, the image plane can be extended beyond the front surface of the prism and into tissue (Fig. 1C–E). This property renders the probe suitable for the replacement of a cannula in a custom-made pressure myograph, where the vessel must be tied to the probe to maintain pressure (Fig. 1B). The focal plane of the optical system can be varied in response to diameter variations in the vessel (Fig. 1C), without moving the probe, by focusing the microscope objective (Fig. 1B) further into the GRIN lens (see also Flusberg *et al.* 2008; Kim *et al.* 2010).

A 0.1 NA GRIN probe was used to ensure a good depth of focus (141 μm) across the entire endothelial surface of curved artery. However, the use of a low NA GRIN relay

lens imposes two opposing constraints on the transmission of light through the system. First, to maximize the delivery of excitation light through the GRIN rod, the NA of the excitation light should not be greater than the NA of the GRIN lens (Fig. 1D). Second, to maximize fluorescence detection, the NA of the collection lens should be higher than that of the GRIN rod. The solution to these opposing constraints is to illuminate the proximal end of the GRIN lens with collimated light, which is re-collimated at the output (Fig. 1E) (Saunter *et al.* 2012) evenly illuminating the entire field of view whilst keeping the illumination constant despite probe refocusing.

The assembled GRIN probe was held in place in a custom-designed arteriograph (Fig. 1G) that was mounted on the microscope stage of a standard inverted microscope (Fig. 1H). The fluorescence excitation and delivery system was constructed using 30 mm cage system components (Thorlabs, UK), and was also mounted onto the microscope stage via a three-axis translation stage. Two axes of the translation stage permitted the external optical system to be coupled to the GRIN microprobe, whilst the third enabled the focus of the system to be altered. Mounting both the arteriograph and the external optical system on the stage ensured that movement of the microscope stage did not decouple the probe from the system. Fluorescence excitation was provided by a fibre-coupled diode-pumped solid-state laser operating at 488 nm, which was collimated (Fig. 1B and H) before being focused by another lens and guided to the back of a 20 \times 0.5 NA infinity-corrected microscope objective (Plan Fluor; Nikon, UK) via a dichroic mirror. Optical excitation power density, measured at the output of the GRIN probe was 1 nW μm^2 . The microscope objective and a 65 mm focal length tube lens imaged fluorescence emission, returning through the probe, through an emission filter and onto a sCMOS camera (Zyla 3-Tap; Andor, UK) controlled by $\mu\text{Manager}$ (Edelstein *et al.* 2010), providing an effective pixel size of 1 μm at the object plane and permitting up to 200 endothelial cells to be imaged with subcellular resolution (see below, Optical characterization).

Optical characterization

To demonstrate the fluorescence signal detected by the microendoscope camera, we recorded images of a diffuse fluorescein solution (1 μM ; Fig. 2A), and of large $15.45 \pm 0.04 \mu\text{m}$ (mean \pm standard deviation; dimensions provided by manufacturer) diameter fluorescent beads (FS07F; Bang Laboratories Inc., USA; Fig. 2B). To image fluorescein fluorescence, the bath chamber was filled with the fluorescent solution. Large-diameter fluorescent beads were diluted in water (100 \times dilution in water) and left to settle on the microprobe prism surface. Figure 2A shows the normalized intensity across the centre of the circular field of view (raw image of fluorescein fluorescence

inset). Due to vignetting of the illumination light and of the fluorescence excitation by the cylindrically shaped GRIN lens, the efficiency drops to 50% at a distance approximately $150 \mu\text{m}$ from the centre of the probe. Translating the focal plane $500 \mu\text{m}$ along the z -axis, beyond the distal prism surface resulted in a slight increase in fluorescence signal, relative to maxima. Note that due to the curvature of pressurized arteries, this intensity profile does not necessarily reflect that obtained in imaging experiments performed in intact arteries. Average optical excitation power density ($1 \text{ nW } \mu\text{m}^{-2}$) was calculated across the full $500 \mu\text{m}$ field of view from power measurements made at the output of the GRIN microprobe using a photodiode power sensor and power

meter (200–1100 nm; S120VC and PM100A; Thorlabs, UK).

To study the resolving power of the micro-endoscope, we imaged subresolution $1 \mu\text{m}$ fluorescent microspheres (F-8823; Invitrogen, UK). The mean fluorescent microsphere diameter (as provided by the manufacturer) was $1.1 \mu\text{m}$ with a coefficient of variation (standard deviation/mean) of 4%. A small droplet of a fluorescent microsphere suspension (1×10^6 dilution in water) was manually pipetted onto the distal prism surface. Fluorescent beads were left to settle on the distal prism surface before being brought into focus and imaged (Fig. 2C). Line intensity profiles of the fluorescence emitted from individual beads were taken (Fig. 2D).

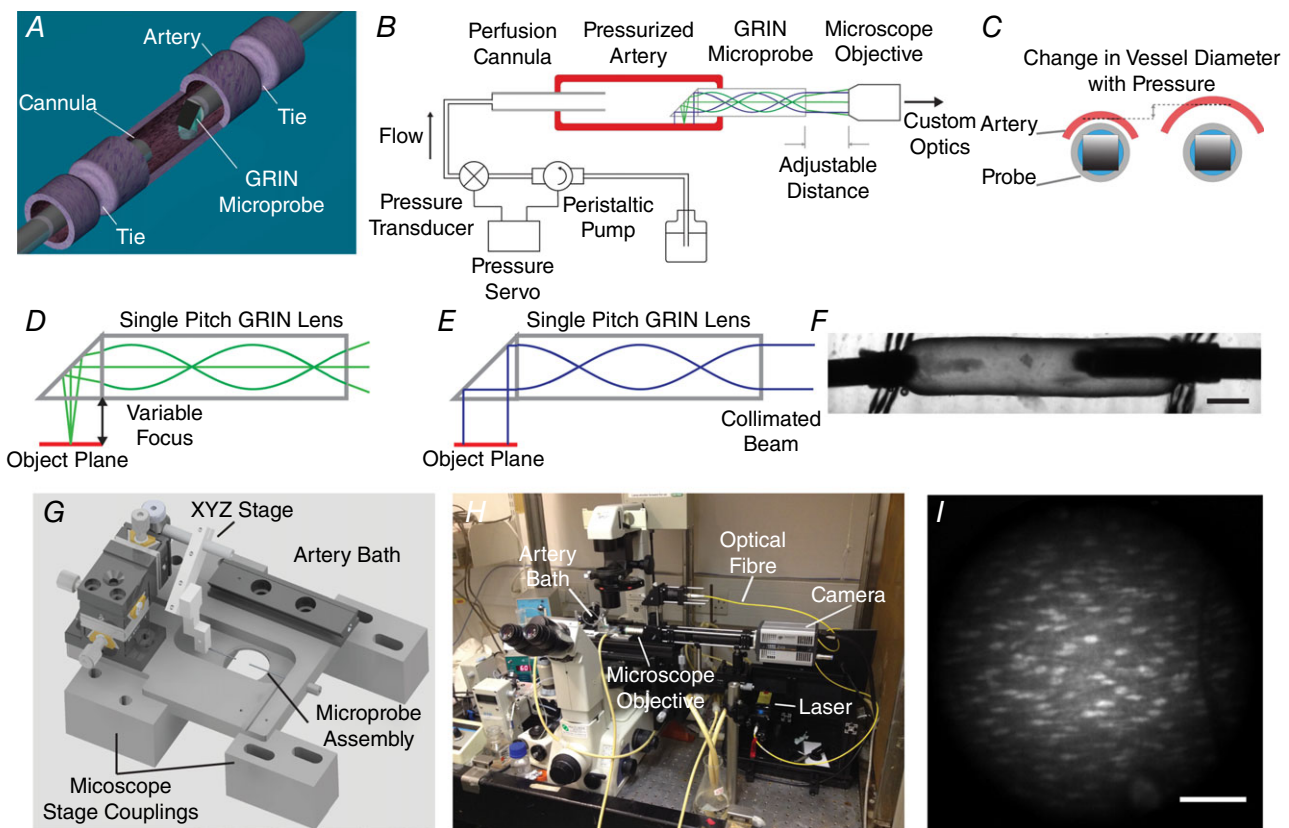


Figure 1. Endothelial GRIN imaging system

A, cartoon showing the GRIN microprobe assembly inside a pressurized artery and endothelium. B, a simplified schematic diagram of the GRIN imaging system and the cannulated, pressurized artery. Focusing into the GRIN microprobe with a conventional microscope objective extended the image plane beyond the front surface of the prism and onto the endothelium. The connecting perfusion cannula (B) and pressure servo system permit intraluminal pressure to be controlled and the probe can be refocused if the artery position moves (C). D, schematic diagram illustrating the optical emission path through the GRIN microprobe. The GRIN lens reconjugates the image plane of a conventional microscope through the length of the cylinder. E, schematic diagram illustrating the optical excitation path through the GRIN microprobe. The collimated input excitation light is re-collimated at the output of the GRIN lens. F, a transmission image of a cannulated artery shows the GRIN microprobe on the right-hand side. G, 3-dimensional rendering of the custom pressure arteriograph showing the position of the GRIN microprobe assembly and XYZ translation stage for positioning the artery. H, picture of set-up during live imaging showing the camera and laser arrangement. I, fluorescence image of the endothelium visualized using the GRIN imaging system showing some activated cells (see online Supporting information, Movie S1). Note the field of view is circular due to the cross-sectional shape of the GRIN lens. Scale bar $100 \mu\text{m}$.

Gaussian fits of these line intensity profiles yielded a lateral resolution of $4.51 \pm 0.04 \mu\text{m}$ ($n = 6$ beads). This measured value may be considered to be the actual resolution of our system because it is much larger than the actual size of the beads (Kim *et al.* 2010). All optical characterization experiments were performed using the same set-up and external optics as that used for intraluminal imaging of the endothelium.

Ca²⁺ imaging

Endothelial Ca²⁺ signals were recorded (5 Hz) from ~ 200 endothelial cells in each artery by GRIN microendoscopy.

The plane of focus of the GRIN imaging system was set to the endothelial layer. Arteries at resting diameter were stimulated by application of ACh ($100 \mu\text{M}$), delivered by a handheld pipette to the outside of the pressurized artery to confirm endothelial viability. Preliminary experiments established that $\sim 80\%$ of the cells in the field of view responded to ACh ($100 \mu\text{M}$). Arteries in which the majority of endothelial cells did not exhibit a Ca²⁺ response to ACh were discarded. To examine the effect of increases in transmural pressure, endothelial Ca²⁺ signalling was recorded at pressures within the physiological range for the artery (60 mmHg, 110 mmHg and 160 mmHg). Following each change in pressure, the imaging system was refocused on

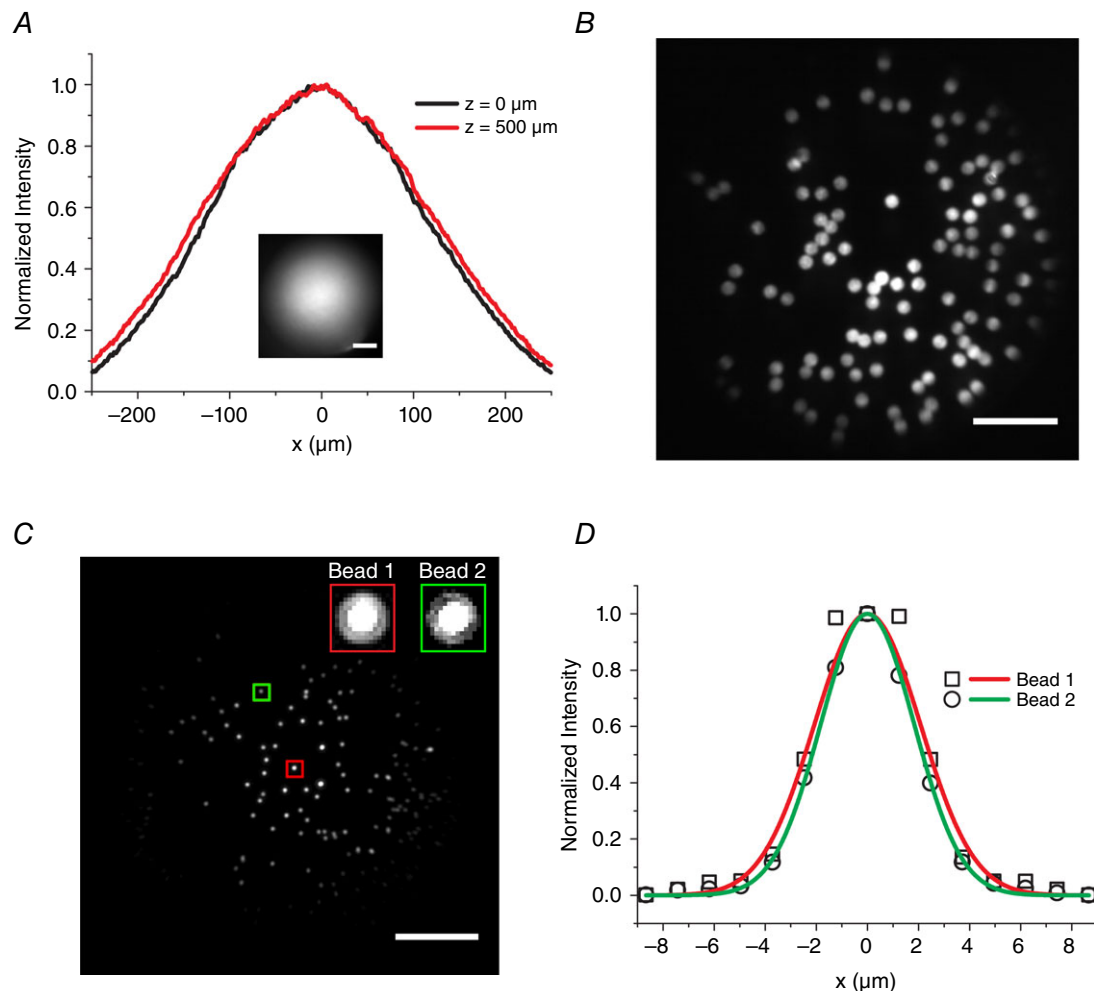


Figure 2. Optical characterization of the side-viewing GRIN imaging system

A, normalized intensity profiles, plotted as a function of radius (x) from the centre of the circular cross-section of the GRIN lens, of images of fluorescein solution ($1 \mu\text{m}$) taken with the GRIN microendoscope as the focus was translated along the z -axis. B, fluorescence image of $15 \mu\text{m}$ fluorescent spheres. C, raw image of sub-resolution ($1.0 \mu\text{m}$) fluorescent beads obtained with the microendoscopic imaging system. Vignetting and heterogeneous fluorescence emission from the beads themselves result in apparent variation in size. By rescaling the intensity of the image (insets), beads of apparently different diameter are shown to be approximately equal in size. D, normalized fluorescence intensity profiles, with Gaussian curves fitted, of the two beads highlighted in A. From such intensity profiles, we calculated the optical resolution of our system to be $4.51 \pm 0.04 \mu\text{m}$ ($n = 6$ beads). Scale bars: $100 \mu\text{m}$.

the endothelium and arteries were left to equilibrate for 20 min. The Ca^{2+} responses evoked by ACh at various transmural pressures were studied in single experiments and expressed relative to the control response (60 mmHg). There was significant overlap in the cells imaged at each pressure, but matching individual cells was not possible. In other experiments, the extraluminal PSS was exchanged for PSS containing various pharmacological inhibitors or Ca^{2+} -free PSS and left for 20 min, before activation again with 100 μM extraluminally applied ACh. The Ca^{2+} responses evoked by ACh in arteries treated with pharmacological inhibitors or Ca^{2+} -free bath solution were studied (control and treatment) in the same artery and expressed relative to the control response. Following each acquisition period, the bath solution was immediately exchanged and the arteries were left for at least 20 min to re-equilibrate. Smooth muscle cells were not loaded with OGB-1/AM in any of our preparations, as indicated by an absence of fluorescence staining orthogonal to the longitudinal vessel axis.

Signal analysis

Individual endothelial cells were segmented using a custom, semi-automated image processing procedure. Previous successful fully automated image processing of endothelial Ca^{2+} signals has been achieved by assigning regions of interest (ROIs), of predetermined size and shape to image sequences based on subcellular activity (Francis *et al.* 2012). Here, the activity of entire cells was used to determine the shape of individual cellular ROIs. In detail, a series of images was created to illustrate the active wavefronts by generating the forward differences of the cytoplasmic Ca^{2+} concentration ($[\text{Ca}^{2+}]_c$) change. First, to facilitate visual inspection of endothelial Ca^{2+} signals, the active Ca^{2+} wavefronts themselves were examined by generating the forward difference of $[\text{Ca}^{2+}]_c$ changes ($F_t - F_{t-1}$, obtained by sequential subtraction (SS); Bradley *et al.* 2003; McCarron *et al.* 2010). Then single images, illustrating all endothelial cells exhibiting ACh-evoked Ca^{2+} activity, were created by taking projections of the standard deviation (STDev) of intensity of SS image stacks. Unsharp masking of standard deviation projections was used to create sharpened, background-corrected STDev images, where ROIs encompassing individual cells could be easily obtained by intensity thresholding. ROIs corresponding to individual cell outlines were verified for each image series and erroneous ROIs were corrected manually. Cell outlines were stored as polygon descriptions within text (.txt) files for subsequent processing and analysis. Except for the creation of standard deviation projections (performed in FIJI; Schindelin *et al.* 2012) and for manually splitting joined cells, all processing was performed using batch-processing algorithms in ImagePro Plus.

Individual fluorescence signals were extracted for each polygonal region from the raw image stacks using a custom program in the Python language. Fluorescence signals are expressed as baseline corrected values (F/F_0), calculated by dividing the raw signals by the average value of using a user-defined period (typically 50 frames) preceding ACh-evoked Ca^{2+} activity. There was a significant variation in the time taken for each cell to respond to ACh. To aid analysis, individual F/F_0 traces were aligned with respect to their peak rate of change. The alignment provides a clear illustration of total Ca^{2+} activity. Baseline values of F/F_0 , peak amplitudes and the time of peak rate of change for each signal were calculated automatically and stored as data tables within .csv files. These .csv files were then imported into Origin 9.1 for calculation of peak changes in fluorescence intensity ($\Delta F/F_0$), and for plotting using custom analysis scripts. For presentation, a 6-point (1.2 s), third-order polynomial Savitzky–Golay filter was applied within Origin; all measurements were from unsmoothed traces.

Measurement of arterial diameter

At the end of some experiments, we recorded the diameter of arteries as pressure was increased, in 5 mmHg increments, from 0 mmHg to 200 mmHg. For videomicroscopy-based diameter measurements, arteries were illuminated with bright field illumination, which was guided to a CCD camera (Sony XC-77; Sony, Japan) mounted on the side-port of the inverted microscope on which our GRIN imaging system was mounted. Images were captured from the CCD camera using Micromanager software (Edelstein *et al.* 2010), and a USB video capture device (Dazzle; Pinnacle Systems, USA) and stored on a computer for subsequent analysis. Due to light scattering by the artery wall, the luminal diameter could not be assessed. Thus, outer artery diameter was measured using the Vessel Diameter plugin for ImageJ (Fischer *et al.* 2010).

Histological analysis

Rat carotid arteries were fixed at pressure after length adjustment (to remove buckle) at 160 mmHg. Following length adjustment, luminal PSS was replaced with Zenker's fixative. Once the lumen was filled with fixative, the artery was sealed and the pressure was immediately raised to 60 mmHg or 160 mmHg, and the extraluminal PSS was replaced with Zenker's solution. Arteries were left to fix for at least 2 h – a time determined, in preliminary experiments to be required to prevent a reduction in arterial dimensions upon removal of pressure. Following fixation, arteries were removed from the myography chamber, washed overnight in tap water and stored in 70% ethanol at 4°C until use. Arteries were paired for

analysis (i.e. two arteries from each animal were fixed at the two different pressures (60 and 160 mmHg).

Following fixation, arteries were dehydrated in alcohol series (70% ethanol, 4 h, 4 changes; 95% ethanol, 2 h, 2 changes; 100% ethanol, 2 h, 4 changes), cleared (1:1 mixture of 100% ethanol, 1 h; Histo-Clear (National Diagnostics, Atlanta, GA, USA), overnight, 4 changes), infiltrated and embedded (paraffin wax, 4 h). Wax blocks were cut into 5 μm thick sections and mounted onto slides. Slides were rehydrated in Histo-Clear (10 min, 1 change), then alcohol series (100% ethanol, 10 min, 1 change; 95% ethanol, 2 min; 70% ethanol, 2 min), before being washed in distilled water. Rehydrated slides were stained with Harris's haematoxylin solution (10 min), washed with warm running tap water (10 min) then Scotts tap water (1 min), differentiated in acid alcohol (0.3%, 5 s), stained with eosin (5 min), washed with warm running tap water (10 min), then dehydrated in alcohol series (70% ethanol, 2 min; 95% ethanol, 2 min; 100% ethanol, 10 min, 1 change), cleared in Histo-Clear (10 min, 1 change), before being mounted with Histo-Clear. Stained artery cross-sections were imaged using a Leica DM LB2 microscope with a Leica DFC320 camera (Leica Microsystems, UK). The thickness of endothelial cell nuclei, used as an indication of height of the endothelial cell layer, were measured using Image Pro Plus.

Solutions and drugs

PSS consisted of (mM): NaCl (145), KCl (4.7), Mops (2.0), NaH_2PO_4 (1.2), glucose (5.0), ethylenediamine-tetraacetic acid (EDTA, 0.02), sodium pyruvate (2.0), MgCl_2 (1.17), CaCl_2 (2.0) (pH adjusted to 7.4 with NaOH). In Ca^{2+} -free PSS, no Ca^{2+} was added, the concentration of MgCl_2 was increased to 3.17 mM, and ethylene glycol tetraacetic acid (EGTA, 1 mM) was included.

Although a relatively high extraluminal concentration of ACh was required to activate the majority of endothelial cells, it is likely that the endothelium was not exposed to this concentration due to the presence of an adventitial barrier to diffusion. The concentration of ACh in the lumen in the present experiments was estimated to be ~ 100 -fold less than the bath concentration at the time of Ca^{2+} measurement. Two experiments support this conclusion. First, in arteries surgically opened, ACh (1 μM) produced approximately equivalent responses (i.e. $\sim 80\%$ cells responding) to ACh (100 μM) applied to the bath in intact artery. Secondly, the concentration of ACh was also estimated from the time course of diffusion of fluorescein across the vascular wall in the same experimental conditions as the pressurized artery. The fractional fluorescence change (relative to the final steady-state value) at the time (10 s after addition) that ACh evoked Ca^{2+} responses was used to estimate the fraction of the ACh present in the lumen (~ 1 μM). In

other studies, the potency of extraluminally applied ACh was reported to be $\sim 1/50$ of intraluminally applied ACh in the dog mesenteric artery (Toda *et al.* 1990) and 50–100 times less potent in femoral artery (Angus *et al.* 1983; Toda *et al.* 1988). Additionally bradykinin is unable to evoke relaxant responses in isolated porcine coronary arteries when applied extraluminally, independent of enzymatic degradation and luminal pressure, but is able to evoke responses when applied intraluminally (Tanko *et al.* 1999).

Drugs were all obtained from Sigma except for OGB-1/AM and Pluronic F-127, which were obtained from Invitrogen. All drugs were dissolved in DMSO and diluted to working concentration in PSS such that the total volume of DMSO was less than or equal to 0.1%.

Harris's haematoxylin, Scotts tap water and Eosin were obtained from and Sigma (UK). Zenker's fixative was obtained from Fisher Scientific.

Statistics

Summarized data are expressed as means \pm standard error of the mean (SEM). One-way ANOVA (with Tukey's *post hoc* test as appropriate) was used for comparisons between groups, and biological replicate (animal) was treated as a random factor. Statistical analyses were performed using Minitab 17 (Minitab Inc., USA). A *P*-value less than 0.05 was considered significant and *n* is number of animals.

Model description

To understand how changes in cell geometry alter IP_3 -evoked Ca^{2+} release, local $[\text{Ca}^{2+}]$ in the IP_3 receptor (IP_3R) microdomain was determined computationally in the time period encompassing ion channel opening. The cytosolic concentrations of ionic Ca^{2+} , buffer and buffered Ca^{2+} are represented by $C_{\text{Ca,cyt}}$, $C_{\text{B,cyt}}$ and $C_{\text{CaB,cyt}}$ respectively. The partial differential equation governing the concentration, C_s , of a species, *s*, is given by:

$$\frac{\partial C_s}{\partial t} = D_s \nabla^2 C_s + \phi_s + J_s \quad (1)$$

where C_s is the concentration, D_s is the diffusion coefficient, ϕ_s is the source term derived from chemical reactions and J_s is the source term resulting from trans-membrane flux, which was taken as zero for all species except Ca^{2+} , for which it comprises IP_3R -mediated Ca^{2+} currents.

Buffering. We employed the first order mass action reaction kinetic:

$$\phi_{\text{Ca,cyt}} = -K_{\text{on}} C_{\text{Ca,cyt}} C_{\text{B,cyt}} + K_{\text{off}} C_{\text{CaB,cyt}} \quad (2)$$

$$\phi_{\text{B,cyt}} = \phi_{\text{CaB,cyt}} = -\phi_{\text{Ca,cyt}} \quad (3)$$

where K_{on} and K_{off} are the rate constants for the buffer.

Trans-membrane flux. Ca²⁺ pumps (SERCAs) and membrane (plasma and ER) leakage currents are typically continuous, low magnitude processes that function to maintain specific concentrations of Ca²⁺ in the cytosol (typically < 100 nM) and the ER (typically > 0.5 mM) in the long term (Table 1). This is in contrast to IP₃Rs, which are reported to open for durations of between 2 ms and 20 ms, with a transient current that is far higher than those of the aforementioned long-term processes. We therefore omitted the slow acting sources and sinks, as their effects over the brief temporal and spatial scales of an individual microdomain, with which we are concerned, are limited. The transport of Ca²⁺ from the ER to the cytosol, through an open IP₃R, is a purely diffusive process, and is therefore driven in linear proportion to the ionic concentration gradient between the two partitions. Therefore individual IP₃R are represented by the source term:

$$J_s = \alpha J_0 \frac{C_{Ca,er} - C_{Ca,cyt}}{C_{Ca,er}} \quad (4)$$

where $C_{Ca,er}$ is the concentration of Ca²⁺ in the ER, taken as a constant, J_0 is the experimentally measured maximal ion current of an isolated IP₃R and α is a conversion factor from a current (in moles per second) to a molar concentration for the voxel to which the current is applied.

Computation. Equation (1) was solved for a regular voxel grid in Cartesian space (Table 1) using a first order finite difference solver. A sufficiently large volume was used such that edge effects are negligible, with the condition being that $C_{Ca,cyt}$ did not exceed a level 1% above baseline at the edges (excluding the edge containing the IP₃Rs), except where we intentionally simulated compressed cells. The Courant–Freidrichs–Lewy condition:

$$\frac{2D_s dt}{dx^2} < 1 \quad (5)$$

was obeyed for each species, where dt is the simulation time step and dx is the linear size of a voxel. Simulation was conducted using bespoke C code employing Intel AVX-2 vector extensions. The Python language was used to process and analyse simulation results.

Boundary conditions. Our model was initialized with homogeneous Ca²⁺ and buffer concentrations as given in Table 1, with $C_{CaB,cyt}$ being initialized to equilibrium values. A zero flux boundary condition was applied to the edges of the cuboidal simulation volume.

Simulation verification. Our simulation code was compared with analytical expressions for simplified test cases of 3D diffusion from a point source and for the

microdomain profile of an isolated Ca²⁺ source in a strongly buffered environment.

Analytical diffusion. Equation (6) gives the analytical form of the concentration profile for a slug of mass M released at position $r = 0$ and time $t = 0$, where r is scalar radius from the origin (Balluffi *et al.* 2005).

$$C(r, t) = \frac{M}{(4 D \pi t)^{3/2}} \exp\left(-\frac{r^2}{4Dt}\right) \quad (6)$$

The Ca²⁺ diffusion component of our simulation was initialized with a volume of 1.68 μm on a side, with Ca²⁺ parameters as per Table 1, and with zero Ca²⁺ concentration except in the central voxel, which was set to 1 M. Figure 3A and B illustrates the good agreement between our simulation and the analytical case.

Analytical microdomain profile. An analytical solution for the equilibrium concentration profile, at distance r from an isolated source of constant current J_{Ca} in an isotropic, exists for an inexhaustible buffer that only forward binds (known as the excess buffer approximation; Smith, 1996):

$$c(r) = c_\infty + \frac{J_{Ca}}{2\pi D_{Ca}} \frac{e^{-r/\lambda}}{r} \quad (7)$$

where c_∞ is the Ca²⁺ concentration far from the source – that is the equilibrium level, $\gamma = (D\tau)^{0.5}$ is a space constant and $\tau = 1/(K_{on}B)$ is the mean capture time of the buffer, of concentration B . Our simulation was configured to match, as well as possible, this analytical case. Specifically parameters from Table 1 were used to simulate a single source, with the following modifications: K_{Off} was set to zero (forward binding only), $\phi_{B,cyt}$ from Eqn (2) was set to zero (inexhaustible buffer) and the concentration gradient was removed from Eqn (3) (constant current source). As a consequence of these changes any background Ca²⁺ was rapidly depleted in this model, so c_∞ was also set to zero to facilitate direct comparison. After running the model, the total quantity of free (unbuffered) cytosolic Ca²⁺ was examined (Fig. 3C). This level rose rapidly, stabilizing to equilibrium in ~ 0.1 ms. A comparison of Eqn (6) and the concentration profile of the model, once equilibrium is reached, is shown in Fig. 3. The responses were very similar, with the effect of quantized voxel sizes limiting the model concentration at small r and edge effects raising the concentration at high r .

Simulation parameters. Parameters for the simulation were chosen to be physiologically relevant, and are given in Table 1 along with references.

Table 1. Numerical parameters used in reaction diffusion simulations

Grid and solver			References and notes
Voxel Scale	dx	0.015 μm	
Solver time step	dt	0.120 μs	
Courant–Freidcrchs–Lewy condition $[\text{Ca}^{2+}]$	$2Ddt dx^{-2}$ must be < 1	0.23	Evaluated for Ca^{2+} as most diffusive
Cytosolic Ca^{2+} Equilibrium concentration	C_{inf}	70 nM	Huang <i>et al.</i> (2000); Means <i>et al.</i> (2006); Bortolozzi <i>et al.</i> (2008)
Diffusion coefficient Cytosolic buffer – parvalbumin	$D_{\text{Ca,cyt}}$	220 $\mu\text{m}^2 \text{s}^{-1}$	Means <i>et al.</i> (2006)
Diffusion coefficient	$D_{\text{B,cyt}}$	90 $\mu\text{m}^2 \text{s}^{-1}$	Bortolozzi <i>et al.</i> (2008)
Diffusion coefficient	$D_{\text{CaB, cyt}}$	90 $\mu\text{m}^2 \text{s}^{-1}$	As for buffer
Forward binding constant	K_{on}	18.5 $\mu\text{M}^{-1} \text{s}^{-1}$	Bortolozzi <i>et al.</i> (2008)
Backward binding constant	K_{off}	0.95 s^{-1}	Bortolozzi <i>et al.</i> (2008)
IP ₃ R			
ER Ca^{2+} concentration	$C_{\text{Ca,er}}$	0.25 mM	Means <i>et al.</i> (2006)
Maximum channel current	J_0	$3.3 \times 10^{-19} \text{ mol s}^{-1}$	Tu <i>et al.</i> (2005); Means <i>et al.</i> (2006)

Rate constants and diffusion coefficients were obtained from Means *et al.* (2006) and Bortolozzi *et al.* (2008) and reported as measured in the cytoplasm of cells.

Results

The GRIN microprobe inserted easily into the lumen of an artery (Fig. 1) delivered a field of view of 0.5 mm diameter and allowed a large number (~ 200) of naturally connected endothelial cells to be imaged (Fig. 1I) with subcellular resolution ($\sim 4.5 \mu\text{m}$; Fig. 2D). In arteries pressurized to 60 mmHg, activation of the endothelium by extra-luminal acetylcholine (ACh; 100 μM applied to the chamber, $\sim 1 \mu\text{M}$ estimated at the vessel lumen; 60 mmHg; online Supporting information, Movies S1 and S2) evoked reproducible rises in cytoplasmic Ca^{2+} concentration ($[\text{Ca}^{2+}]_c$) in the majority of cells in the field (Fig. 4A). The response was composed of temporally distinct components. Initially, small spatial groupings of cells (macrodomain) activated (Fig. 4A, Movie S2) in small regions of the endothelium and from which waves expanded (Fig. 4A) to recruit the remaining majority of cells in the field of view (Figs 4 and 5).

As a result of the wave progression there was a large spread of times to first Ca^{2+} response (latency) in various cells (Fig. 4B and C, left panels) which complicated summarizing and evaluating responses. Therefore, the temporal latency between individual Ca^{2+} traces (F/F_0) was removed by aligning the signals to each other (using the initial peak in the first derivative of each signal) (Fig. 4B and C, middle panels). This procedure removed heterogeneity resulting from the time to onset

of individual signals and provided a clearer illustration of total Ca^{2+} activity. Upon consecutive stimulation with ACh, the response, as measured from the number of cells activated and the average peak change in F/F_0 , was approximately reproducible (Fig. 4D, $n = 5$).

The Ca^{2+} rise evoked by ACh was approximately reproducible on successive application of ACh (Fig. 4D; $n = 5$). The ACh-evoked Ca^{2+} rise persisted in a Ca^{2+} -free bathing solution ($n = 4$), but was blocked by the SERCA inhibitor cyclopiazonic acid (10 μM ; $n = 3$) and the IP₃R blocker 2-aminoethoxydiphenyl borate (100 μM ; $n = 3$; Fig. 6). Caffeine (10 mM; $n = 3$) did not evoke a Ca^{2+} increase and the ACh-evoked Ca^{2+} rise was unaltered by ryanodine (10 μM ; $n = 3$). These results suggest that the Ca^{2+} rise originated from an IP₃-sensitive Ca^{2+} store and that RyRs play a minor role in Ca^{2+} signalling in the endothelium (Fig. 6). The TRPV4 antagonist RN1734 (30 μM ; 30 min) did not decrease the ACh-evoked endothelial response at 60 mmHg (Fig. 6D) – as expected from previous findings in larger arteries (Hartmannsgruber *et al.* 2007; Loot *et al.* 2008).

Endothelial cells are stimulated almost constantly by extrinsic blood-borne bioactive molecules amid a background of haemodynamic mechanical forces. The combined effect of chemical activators and mechanical force is presumably incorporated into the endothelium's biological response, but details are largely unknown. We addressed whether a change in endothelium-dependent

Ca^{2+} signalling, evoked by a bioactive molecule (ACh), occurs with variation in transmural pressure, and hence mechanical force. At 60 mmHg, ACh ($100 \mu\text{M}$) evoked asynchronous Ca^{2+} waves and oscillations that originated in macrodomains (Fig. 7A; Movie S2). As transmural pressure increased (60 to 110 to 160 mmHg), both the amplitude of the Ca^{2+} rise that occurred in each cell and the temporal spread of total activation decreased (Fig. 7A–D; Movie S3). The suppression of the amplitude of the Ca^{2+} signals reversed as pressure decreased (not shown). These results suggest that increased pressure in the intact artery suppressed IP_3 -evoked Ca^{2+} signals. IP_3R itself is not reported to be stretch sensitive, and mechanical activation of IP_3 -generating processes is unlikely to explain

our observations as transient increases in IP_3 production and IP_3 -evoked Ca^{2+} release occur typically (Jena *et al.* 1997).

The question arises as to how increased pressure suppressed IP_3 -evoked Ca^{2+} signals. Significant changes in endothelial cell shape occurred with increased pressure (Fig. 8). Increased circumferential stretch, with increased pressure, elongated cells around the artery, and cell thickness was significantly decreased to maintain volume (Fig. 8A and B). Increased transmural pressure may have also compressed the endothelial cells. The change in cell geometry is likely to alter Ca^{2+} dynamics in the endothelial cells at the microdomain level near IP_3R . The effect of changes in cell geometry on IP_3 -evoked Ca^{2+} signals

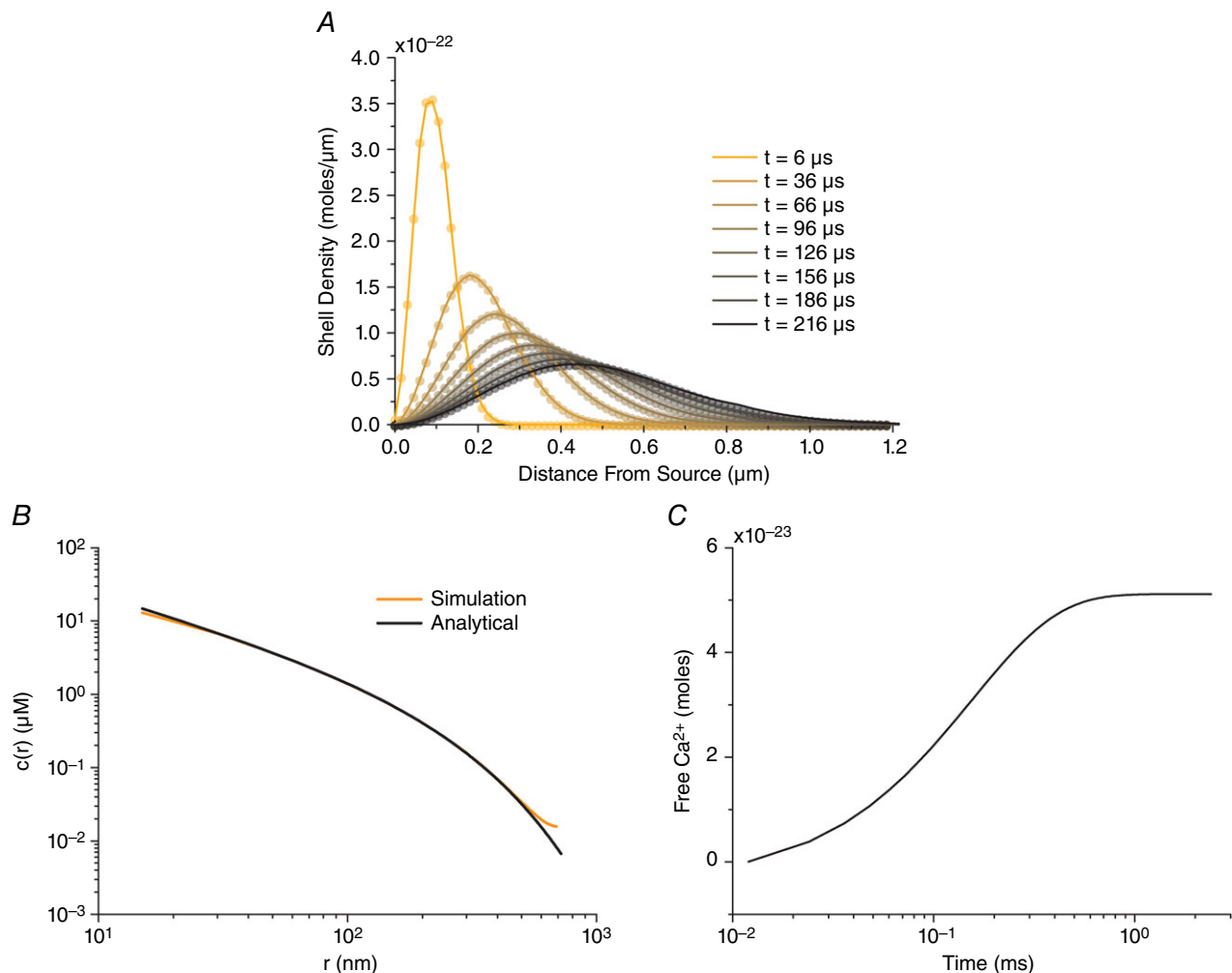


Figure 3. IP_3 -evoked Ca^{2+} release simulation

A, shell density plotted against distance from source (i.e. IP_3R). Theoretical (analytical test case; lines) plots of shell density for a slug of mass released at the origin shows good agreement with our simulated results (circles). B, Ca^{2+} concentration as a function of radial distance from IP_3Rs . When equilibrated, the Ca^{2+} profile may be approximated by an analytical case (black line), which compares accurately to an examination of our simulation (yellow line). C, free Ca^{2+} concentration at an IP_3R as a function of time from receptor opening. The total cytosolic Ca^{2+} rises immediately after IP_3R opening then plateaus as the current flowing through IP_3R and cytosolic Ca^{2+} buffering reaches equilibrium (~ 0.1 ms). See Methods and Table 1 for equations, source references and parameters used to generate the figures.

was therefore considered further. To explore the effects of changes in the cells' geometry on IP₃-mediated Ca²⁺ release at the relevant scale, which exists below functional optical microscopy, we computationally simulated events at an IP₃R cluster.

IP₃Rs are arranged in clusters and the sensitivity of IP₃Rs to Ca²⁺ is such that the [Ca²⁺]_c in the microdomain of a single open IP₃R (or cluster) may be high enough to activate nearby IP₃Rs (or clusters) (Callamaras *et al.* 1998). This process of Ca²⁺ activation may become regenerative and carry a wave of activity across cells (Figs 4 and 5). In the transmission process, IP₃R activity is exquisitely sensitive to distance between channels on the ER and to the cytosolic space between the ER and membrane structures, such as

mitochondria and the plasma membrane (McGeown *et al.* 1996; Rizzuto *et al.* 1998; Mazel *et al.* 2009; Olson *et al.* 2010, 2012). These features (clustering, cytosolic space) create a local diffusive environment (i.e. microdomain) in which Ca²⁺ and Ca²⁺ buffers regulate the activity of IP₃Rs and the release of Ca²⁺ from the ER due to the local [Ca²⁺]_c.

The opening of a single IP₃R introduced a point source of Ca²⁺ that equilibrated rapidly by diffusion and aggressive cytosolic buffering, to produce a localized, quasi-static region of elevated [Ca²⁺]_c (microdomain; Fig. 9A and B) around the channel. The Ca²⁺ concentration decreased sharply with distance from the IP₃R with a characteristic scale (the distance at which [Ca²⁺]_c remains

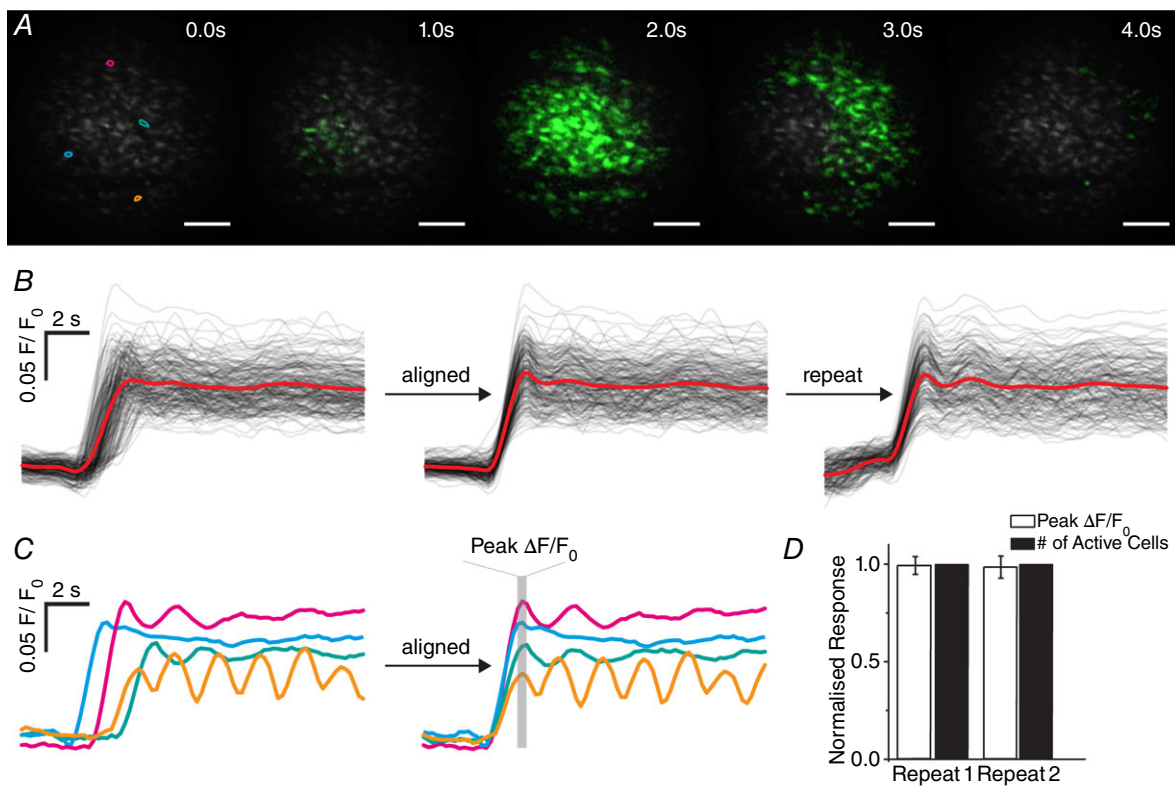


Figure 4. Endothelial Ca²⁺ imaging in intact, pressurized (60 mmHg) arteries

A, time series Ca²⁺ images of endothelial cells showing the progression of the Ca²⁺ response evoked by ACh (100 μM; bath applied; 1 μM estimated at the vessel lumen) recorded at 60 mmHg. Images are composed of instantaneous Ca²⁺ activity (green) overlaid on standard deviation images (greyscale) indicative of total Ca²⁺ activity. The rise began in a distinct cluster of cells (see 1 s) and progressed from there. Each bright spot is an individual cell. Scale bar: 100 μm. B, ACh-evoked Ca²⁺ signals from the same cells shown in A. In the left panel (unaligned data) the range of times for Ca²⁺ to increase for each of the ~200 cells is approximately 4 s. The spread of responses resulted in mean data representing the data poorly. The position of the first peak of derivative signals was used to align Ca²⁺ signals. This procedure synchronizes the Ca²⁺ rises occurring in each due to the action of ACh (middle panel 'aligned') and illustrates total Ca²⁺ activity (thick red line) with increased clarity. On successive application of ACh (20 min reequilibration following wash), the response to ACh was reproducible (right panel). C, select examples of unaligned (left) and aligned (right) cellular Ca²⁺ responses from ROIs shown in A. The grey box illustrates the time point of measurements from aligned signals. D, summary data illustrating the reproducibility of Ca²⁺ responses upon repeat application (wash + 20 min reequilibration) with ACh. Data are presented as means ± SEM and normalized to control (*n* = 5; first ACh application, 1; not shown), *P* < 0.05 was considered significant.

elevated by at least one order of magnitude) on the order of 100 nm (Fig. 9B). The presence of many such point sources resulted in a greater $[\text{Ca}^{2+}]_c$ in the vicinity of IP_3R clusters than isolated receptors (Fig. 9C). The raised $[\text{Ca}^{2+}]_c$ that occurred as a result of IP_3R clustering reduced the local ion gradient between the store ($[\text{Ca}^{2+}]_{\text{ER}}$) and the cytosol, and so reduced the entropic force driving Ca^{2+} release through the channel. In addition, the elevated $[\text{Ca}^{2+}]_c$ merged the microdomain of each IP_3R into a unified, larger domain with scales approaching the 500 nm thickness typical of endothelial cells.

In wide, thin cells (e.g. endothelium) IP_3R s release Ca^{2+} into the cytosolic space between the ER and the cellular plasma membrane, where proximity of the IP_3R to the cellular plasma membrane may create a restricted environment limiting the diffusion of Ca^{2+} . The decreased cell height as a result of the change in cell shape (Fig. 8) reduces the distance between the IP_3R s and the opposing plasma membrane, restricting Ca^{2+} diffusion away from

IP_3R and, as a result, increasing the $[\text{Ca}^{2+}]_c$ at the receptors (Fig. 9A and C). Thus, the decreased height reduced the total Ca^{2+} liberated from the ER by reducing the entropic force driving Ca^{2+} release (Fig. 9D). In this way, a mechanical force (circumferential stretch) can reduce Ca^{2+} release via IP_3R , in the absence of any force- or stretch-sensitive molecular machinery. Notably, the clustering of IP_3R s extends this effect, when compared to isolated receptors, from sub-100 nm thick cells to those with a 100–500 nm thickness typical of the endothelium (Fig. 9).

Discussion

The endothelium's control over cardiovascular structure and function requires cooperation among endothelial cells to both sense and communicate stimuli efficiently with itself and other cell types (Ledoux *et al.* 2008; Behringer & Segal, 2012; Tran *et al.* 2012; Socha *et al.*

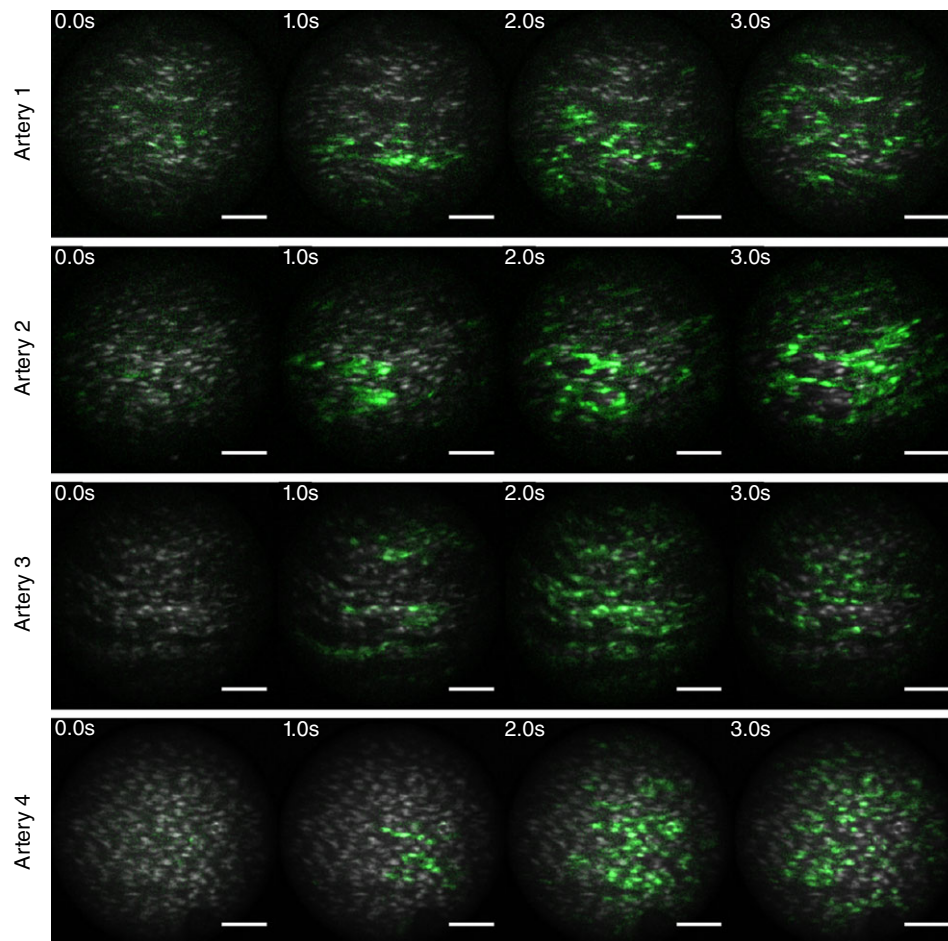


Figure 5. Large-scale, ACh-evoked endothelial Ca^{2+} waves in four separate arteries

In each artery, a time series of Ca^{2+} images from the endothelium illustrates that large-scale, ACh-evoked (100 μM ; bath application) Ca^{2+} waves originate in distinct clusters of cells and propagate from there across the endothelium. The images are composed of instantaneous Ca^{2+} activity (green) overlaid on standard deviation images (grayscale), which are indicative of total Ca^{2+} activity. Each bright spot is a single endothelial cell. Scale bars: 100 μm .

2012*b*; Qian *et al.* 2013). The cooperative behaviour is not well understood because of the difficulties in studying large numbers of endothelial cells in a physiological configuration. The miniature, wide-field GRIN imaging probe described permits endothelial Ca^{2+} activity from within an intact, pressurized artery to be observed. The microprobe bypasses the difficulties created by the

artery wall in imaging blood vessels in a physiological configuration. Furthermore, the absence of biological tissue between the end of the probe and the endothelium minimized the light intensity required. The microprobe has a large depth-of-focus (141 μm) and delivers a 0.5 mm field of view to allow 200 cells to be viewed simultaneously.

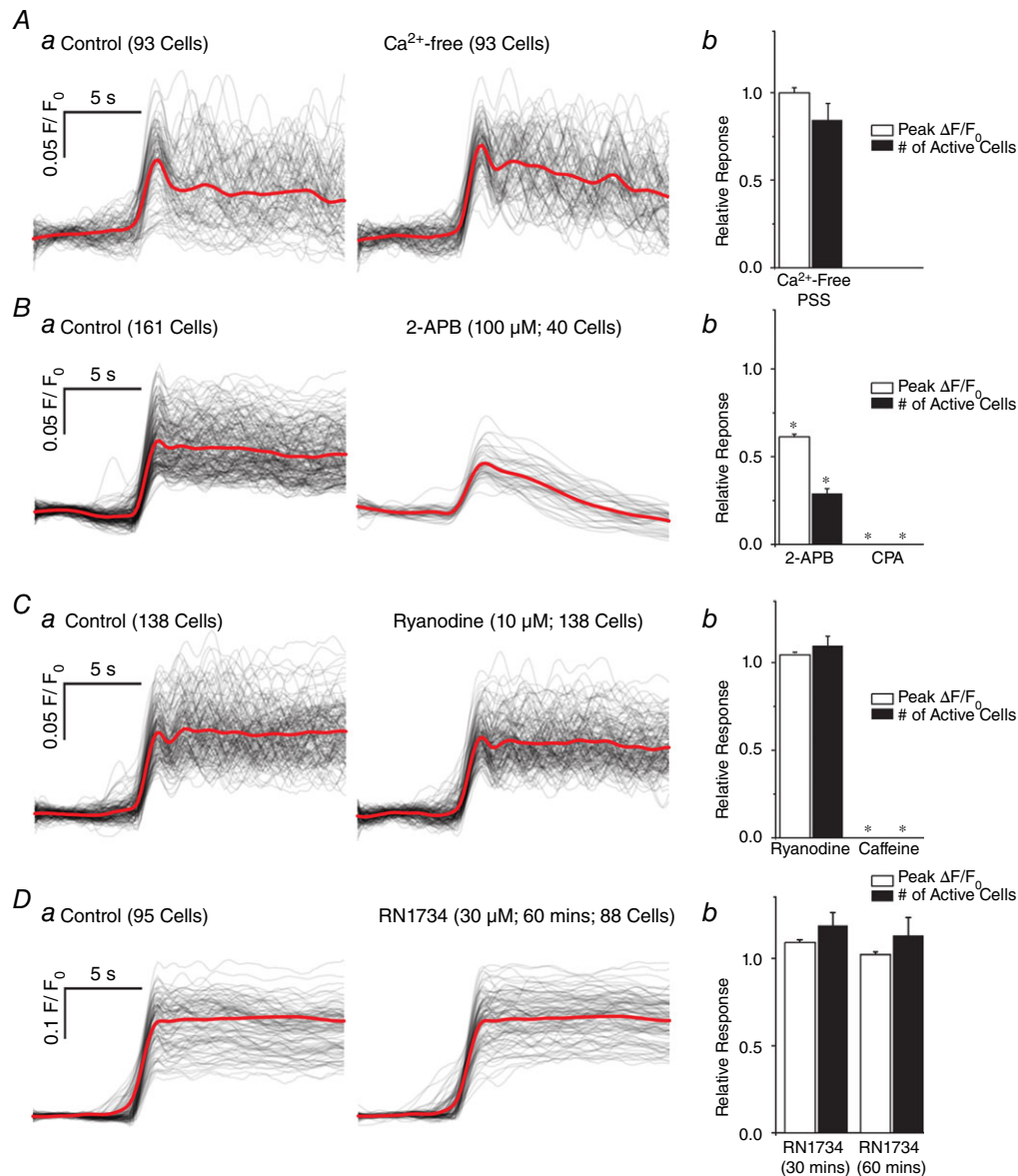


Figure 6. The effects of Ca^{2+} -free bath solution and pharmacological activators and inhibitors on ACh-evoked Ca^{2+} increases

Individual Ca^{2+} traces are shown on the left (a) and summarized data are presented as means \pm SEM and normalized to control (no treatment, 1; not shown) on the right (b). *Aa* and *b*, the ACh-evoked Ca^{2+} was comparable before and after the removal of Ca^{2+} from the bath solution (1 mM EGTA; $n = 4$). *Ba* and *b*, the ACh-evoked Ca^{2+} rise was substantially reduced by aminoethoxydiphenyl borate (2-APB, 100 μM , $n = 3$) and inhibited by the SERCA blocker cyclopiazonic acid (*Bb*; CPA, 10 μM , $n = 3$). *Ca* and *b*, ACh-evoked responses persisted in the presence of ryanodine (10 μM ; $n = 3$). Caffeine (10 mM; $n = 3$; *Cb*) failed to evoke any Ca^{2+} response. *Da* and *b*, ACh-evoked responses were not reduced by the TRPV4 blocker RN1734 (30 μM ; $n = 3$). Individual cellular Ca^{2+} traces are shown in grey with average overlaid in red. $P < 0.05$ was considered significant.

The detection of chemical stimuli appears to be distributed among endothelial cells. In response to ACh, Ca^{2+} rises initiate in small regions of the endothelium and expand from these regions to recruit most of the cells in the field. The initiator regions (macrodomains) appear to be more sensitive to ACh. The path of the Ca^{2+} signal from the sensitive regions at times appeared poorly coordinated among endothelial cells—even chaotic. Nonetheless, the path and pattern were repeatable on successive application of ACh suggesting an encoded signal communicated among cells. The response to ACh was derived from an IP_3 -mediated release of Ca^{2+} from the internal store. The Ca^{2+} rise persisted in a Ca^{2+} -free bath solution and was abolished by the SERCA pump inhibitor

cyclopiazonic acid. The ACh-evoked Ca^{2+} rise persisted in ryanodine and caffeine failed to evoke a Ca^{2+} rise. However, release was inhibited by the IP_3 R blocker 2-APB. The transmission of the Ca^{2+} signal among endothelial cells may involve Ca^{2+} or IP_3 or both.

In addition to receiving and communicating chemical stimuli, arteries are under permanent mechanical activation from shear stress and blood pressure. While cells often respond to the activation of mechanically sensitive components (e.g. via shear stress) with increased ion channel and cell activity, the mechanical response to pressure differs and endothelial activity is suppressed. For example, transient and chronic increases in pressure each inhibit endothelial nitric oxide-mediated vasodilatation

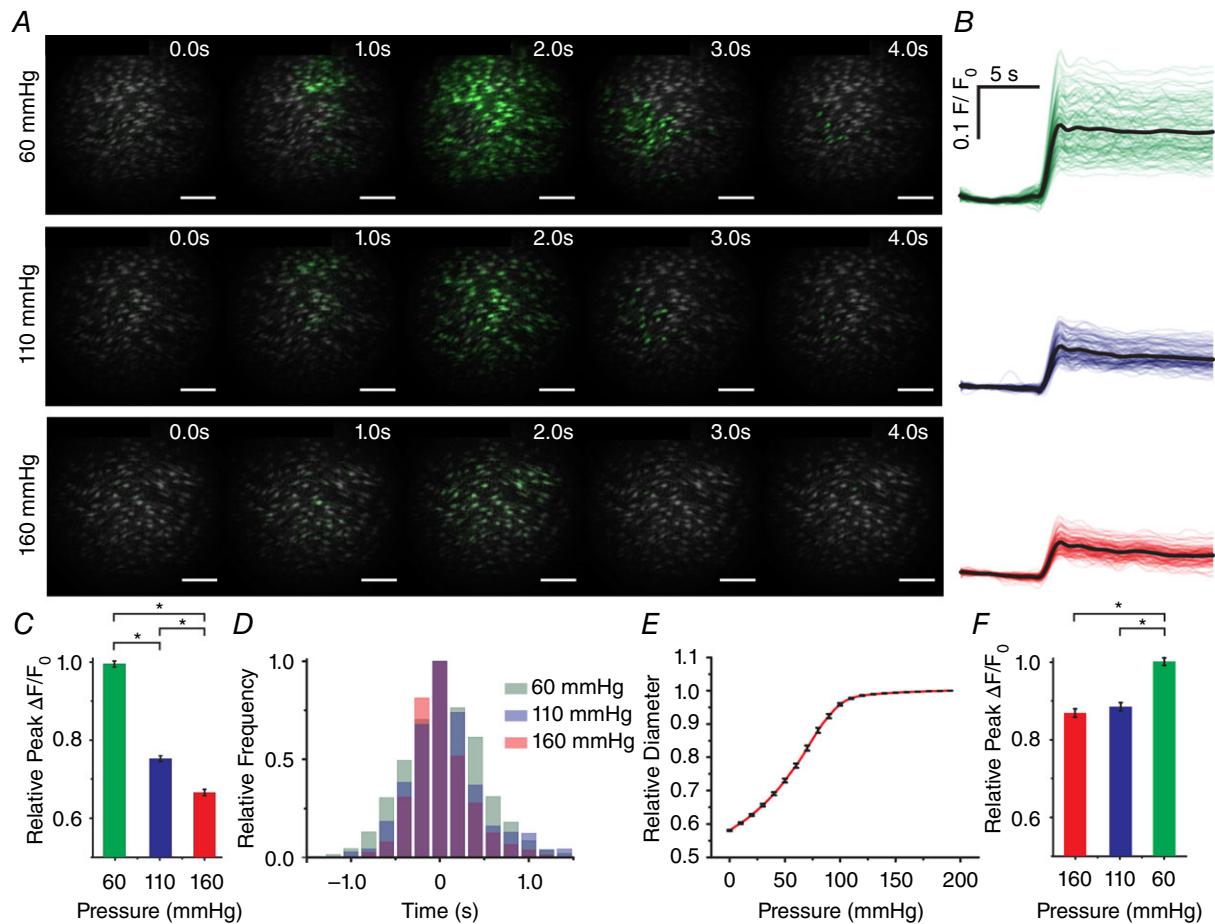


Figure 7. Increased transmural pressure decreases endothelial Ca^{2+} signalling

A, ACh-evoked Ca^{2+} images from endothelial cells at 60 mmHg (top), 110 mmHg (middle) and 160 mmHg (bottom). Ca^{2+} activity decreased as pressure increased in the three repeated activations of the same artery (20 min wash/re-equilibration period between each ACh application). The data are shown as described in Fig. 2A. Scale bar 100 μm . B, baseline corrected and aligned Ca^{2+} signals (average overlaid in black and shown in colour in the bottom panel) from the Ca^{2+} images shown in A. C, summary data illustrating the pressure-dependent decrease in peak $\Delta F/F_0$ ($n = 8$, $\pm\text{SEM}$). D, the temporal spread of cellular activation decreases with increased pressure. Note the fourth colour in the histogram (purple) arises from overlap of colours from the data in 110 mmHg (blue) and 160 mmHg (red). E, summary data illustrating a logistic change in vessel diameter as pressure is increased (from 0 to 200 mmHg; $n = 3$, $\pm\text{SEM}$), measured using the Vessel Diameter plugin for ImageJ. Artery diameter was $1135 \pm 110 \mu\text{m}$ at 60 mmHg, $1429 \pm 92 \mu\text{m}$ at diameter at 110 mmHg and $1460 \pm 83 \mu\text{m}$ at 160 mmHg ($n = 3$). F, summary data illustrating an increase in peak $\Delta F/F_0$ as pressure is decreased ($n = 6$, $\pm\text{SEM}$).

(Hishikawa *et al.* 1992; De Bruyn *et al.* 1994; Huang *et al.* 1998; Paniagua *et al.* 2000; Jurva *et al.* 2006; Phillips *et al.* 2011). How pressure may suppress endothelial activity is unresolved but increased reactive oxygen species may contribute (Huang *et al.* 1998; Vecchione *et al.* 2009; Zhao *et al.* 2015). Here we report that endothelial Ca^{2+} signalling is suppressed when pressure is elevated. The suppressed Ca^{2+} signalling will presumably result in reduced production of nitric oxide and reduced endothelium-dependent dilatation (Hishikawa *et al.* 1992; Sun *et al.* 2001).

The sensing system(s) responsible for detecting changes in pressure in the endothelium are poorly understood. Pressure-evoked mechanical *deactivation* of ion channels has been proposed to account for some physiological responses (Hoffman *et al.* 2011; Bagher *et al.* 2012). We propose that changes in cell shape may be sufficient to underlie the detection of pressure-dependent mechanical forces by the endothelium. The changes in geometric shape of the cell in response to pressure changes are substantial: endothelial cells may be flattened either by circumferential stretch, as the height of endothelial cells

is reduced to maintain volume, or by compression due to the radial force of pressure or both. Such changes in the geometric shape of the cell will bring the endoplasmic reticulum and plasma membrane closer together and create a region of restricted diffusion (see also Hong *et al.* 2008; Qi *et al.* 2015). As shown in Fig. 9, the region of restricted diffusion changes the local diffusive environment surrounding IP_3R clusters and results in a higher concentration of Ca^{2+} near the IP_3R microdomain. This higher concentration results in a reduced Ca^{2+} gradient between the store and the cytosol. As a result, the entropic force driving Ca^{2+} release is reduced and so the amplitude of IP_3 -evoked Ca^{2+} signals is also reduced (Fig. 10). Reduced IP_3 -evoked Ca^{2+} release may explain the suppression of artery level endothelium-derived responses at high transmural pressure (De Bruyn *et al.* 1994; Huang *et al.* 1998; Paniagua *et al.* 2000; Zhao *et al.* 2015). A specific force- or stretch-sensitive component may not be required, but rather changes in cell shape may alter the local concentrations of second messengers (e.g. Ca^{2+}) within the cell to modulate the overall response of the cell.

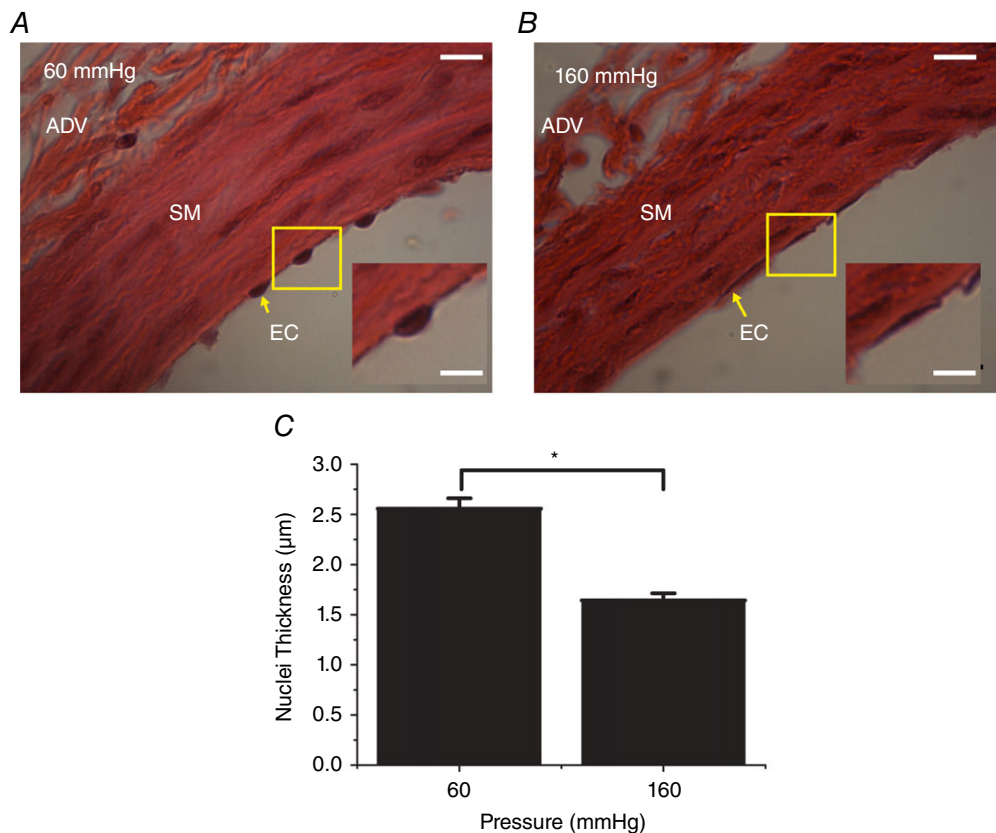


Figure 8. Endothelial cell flattening as intraluminal pressure is increased

Arteries pressurized to 60 mmHg (A) and 160 mmHg (B) then fixed show flattening of the endothelial cells (EC) at higher (160 mmHg) pressure. The insets (A and B) show the yellow box expanded to illustrate the change in shape of single endothelial cells at each pressure. SM, smooth muscle; ADV, adventitia. Scale bars 10 μm main picture, inset 5 μm . C, average nuclei thickness (a measure of cell depth) at 60 mmHg and 160 mmHg. At 60 mmHg, $n = 37$ cells from 3 animals; 160 mmHg $n = 35$ cells from 3 animals ($*P < 0.05$).

The pressure-sensitive change in IP₃-evoked Ca²⁺ signalling provides an additional level of complexity and subtly to mechanosensitive signalling, which may induce further signals to generate a functional response. For example, the changes in [Ca²⁺]_c as a result of geometric modulation of IP₃-evoked Ca²⁺ release may modulate the activity of plasma membrane-located Ca²⁺-sensitive ion channels to explain, at least in part, some contradictory observations of apparent mechanical activation (shear stress, Strotmann *et al.* 2000; Alessandri-Haber *et al.* 2003; Loukin *et al.* 2010) and inhibition (pressure increases, Bagher *et al.* 2012) of ion channels. In the latter study, when pressure decreased from 80 mmHg to 5 mmHg, the frequency of transient localized endothelial Ca²⁺ rises

increased (Bagher *et al.* 2012). The authors attributed the Ca²⁺ changes to influx via TRPV4. The mechanosensor responsible for the pressure-induced change in TRPV4 activity was unclear but the authors suggested reduced radial compression of endothelial cells may activate the channel (Bagher *et al.* 2012). TRPV4 is Ca²⁺ sensitive and activated by increases in [Ca²⁺]_c (Strotmann *et al.* 2003). Interestingly, in their study Bagher *et al.* (2012) found the local endothelial Ca²⁺ events studied were blocked fully by a phospholipase C inhibitor (U-73122), and an IP₃R blocker (xestospongine C) almost abolished Ca²⁺ events at 80 mmHg and significantly inhibited responses at 5 mmHg. These results suggest that IP₃-mediated Ca²⁺ release may contribute significantly to the spontaneous

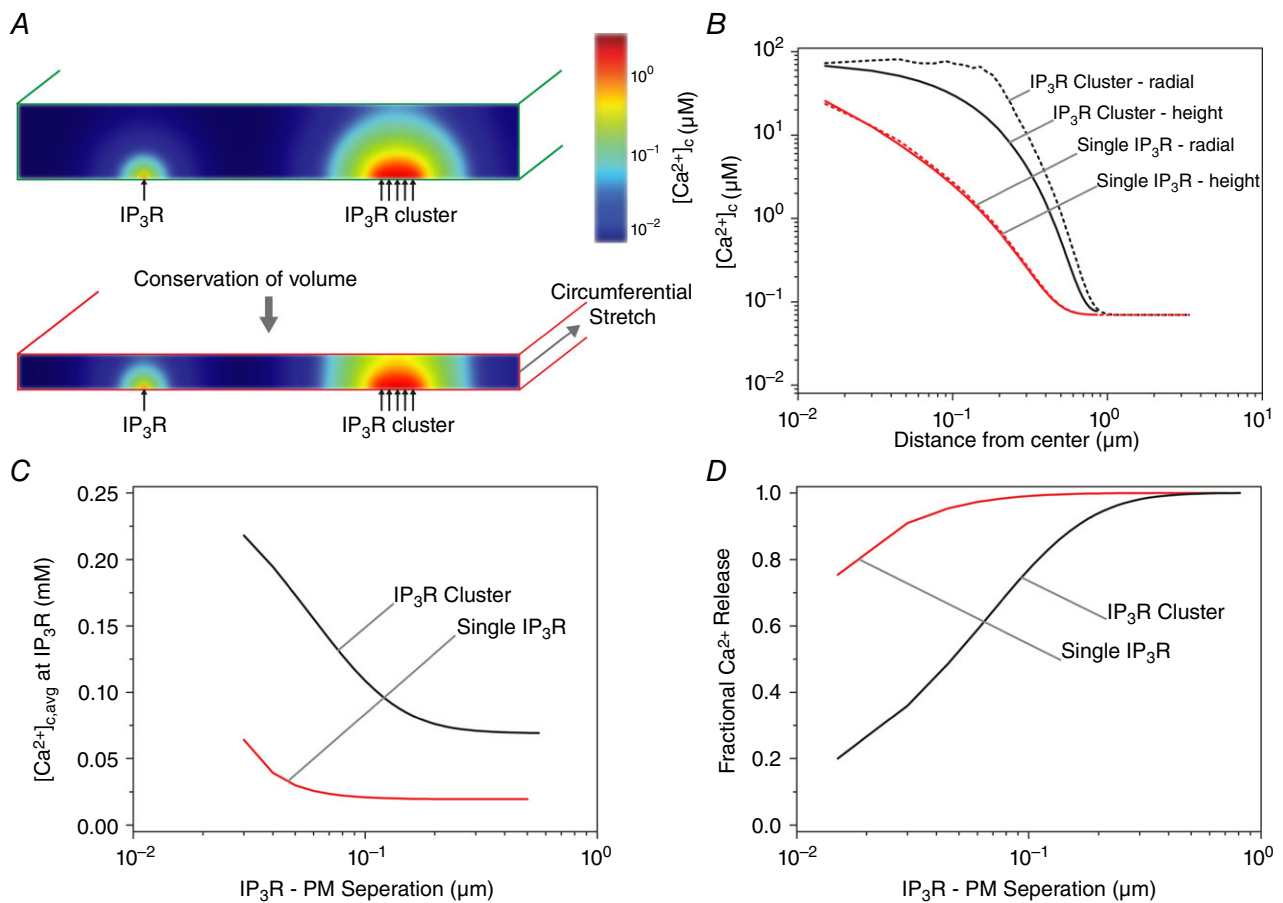


Figure 9. Modelling endoplasmic reticulum Ca²⁺ release from solitary and clusters of IP₃ receptors in cells subject to pressure-induced stretch

A, cartoon illustrating the cytosolic Ca²⁺ distribution during Ca²⁺ release through a single IP₃R and a cluster of IP₃Rs as a cell (top, cell height: 0.57 μm) is stretched in the circumferential direction and thins to conserve volume (bottom, cell height: 0.27 μm). B, concentration profiles for [Ca²⁺]_c surrounding individual and clustered IP₃Rs shows that the microdomain of elevated [Ca²⁺]_c is significantly larger for clusters of receptors. C, the average [Ca²⁺]_c in the vicinity of IP₃Rs is sensitive to an increasing proximity between the ER and the plasma membrane (cell thinning). D, this results in a sensitivity of fractional Ca²⁺ release from internal stores that changes with receptor–plasma membrane distance (cell height) for single IP₃R and a cluster of IP₃Rs, with clusters having sensitivity over significantly larger cell heights, on the order of 500 nm.

Ca²⁺ events. Geometric regulation of IP₃-evoked Ca²⁺ release may provide the pressure sensor for the change in TRPV4 activity.

Endothelial cells are acknowledged to be sensitive to their geometry and changes are known to regulate proliferation and cell death (Chen *et al.* 1997). Changes in cell shape alter tension in the cell membrane to modify components within membranes (Sheetz, 1993; Mederos y Schnitzler *et al.* 2008) with consequences for cell behaviour (Mossman *et al.* 2005). Alternatively, changes in links between the plasma membrane and cytoskeleton or mechanosensitive ion channels may detect changes in membrane tension (Patel *et al.* 2001). Several proteins also have domains (BAR or ALPS) found in membranes that are curved and may contribute to detecting cell shape changes. The extent of membrane binding of these protein domains changes with membrane curvature (Zhao *et al.* 2011). The protein domains may either induce or are sensitive to membrane curvature (Zimmerberg & Kozlov, 2006). Guanine nucleotide exchange factors and GTPase activating proteins with curvature sensing protein domains may alter the activity of GTPases in a tension-dependent manner (de Kreuk *et al.* 2011) to explain geometry sensitivity of cells. We propose changes

in cell geometry may also alter ion diffusional gradients, which act then to sense stimuli and evoke vascular responses.

For normal function, cells in the vascular wall must act in a coordinated manner. The spatial relationships among extracellular chemical stimuli, cellular architecture and Ca²⁺ signals are central to signal processing and the coordinated behaviour. Indeed, although ‘sensing’ may occur on the microdomain scale, the macroscopic configuration of the intact endothelium is essential for physiological mechanotransduction. The detection of mechanical forces like pressure is critically dependent on the organization of the sensing proteins and the contact of cells with other cells. In other investigations on mechanotransduction, a single type of protein was found to evoke different responses when either organized in 3D filaments or arranged on a flat surface (Cukierman *et al.* 2001). Some types of mechanosensitivity seen in isolated single cells were lost when cells are in contact with one another (Yeung *et al.* 2005). Our results in intact arteries in a physiological configuration suggest that mechanical activation by pressure may be merged into the agonist-evoked response in the endothelium. The pressure dependence of Ca²⁺ release provides a mechanism for transmural pressure to

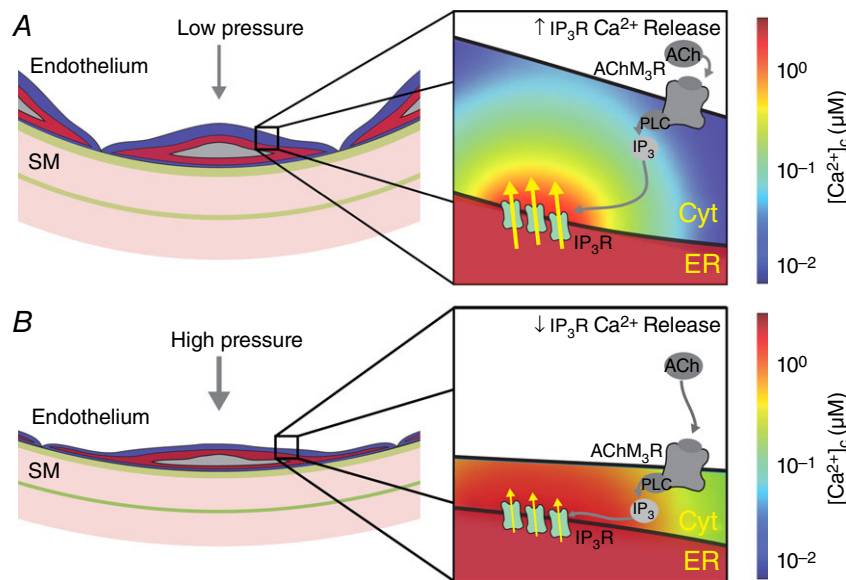


Figure 10. Model of pressure-dependent, geometric suppression of Ca²⁺ release through IP₃Rs

As pressure is increased, from low (A) to high (B), arterial diameter increases and endothelial cells are stretched along the circumferential direction. In addition, the radial force of pressure acts upon the cells. To conserve volume, the height of endothelial cells decrease (B). The insets depict the processes leading to changes in ACh-induced Ca²⁺ release in an endothelial cell at low (A, top) and high (B, bottom) pressure. At low pressure, activation of IP₃R, after ACh application, results in Ca²⁺ release from the endoplasmic reticulum (ER). Ca²⁺ diffuses rapidly from the release site to maintain a gradient for continued release. The microdomains of Ca²⁺ operate over sizes (300 nm) comparable to the height of an endothelial cell (see text). When pressure is increased (B), the height of the cell decreases and so the distance between the ER and the plasma membrane (PM) also decreases. The decrease in ER–PM distance restricts the diffusion of Ca²⁺ from the release site (IP₃Rs), effectively increasing the local Ca²⁺ microdomain concentration and decreasing the entropic force (indicated by size of yellow arrows) driving Ca²⁺ release through IP₃Rs. The resultant effect is a decrease in Ca²⁺ release from the ER as pressure is increased. SM: smooth muscle.

be encoded and integrated to a common, coordinated output without imposing a requirement for mechanosensitivity on any of the molecular machinery in the signalling pathway. The time course of this response would be rapid and may form an early event in an altered mechanical environment. Furthermore, our results suggest how hypertensive conditions may limit the ability of cells to engage in Ca²⁺ signalling, offering insight into the associated degeneration of endothelial response (Luscher & Vanhoutte, 1986; Wallace *et al.* 2007) and the role that IP₃R clustering may play.

References

- Alessandri-Haber N, Yeh JJ, Boyd AE, Parada CA, Chen X, Reichling DB & Levine JD (2003). Hypotonicity induces TRPV4-mediated nociception in rat. *Neuron* **39**, 497–511.
- Angus JA, Campbell GR, Cocks TM & Manderson JA (1983). Vasodilatation by acetylcholine is endothelium-dependent: a study by sonomicrometry in canine femoral artery in vivo. *J Physiol* **344**, 209–222.
- Bagher P, Beleznai T, Kansui Y, Mitchell R, Garland CJ & Dora KA (2012). Low intravascular pressure activates endothelial cell TRPV4 channels, local Ca²⁺ events, and IKCa channels, reducing arteriolar tone. *Proc Natl Acad Sci USA* **109**, 18174–18179.
- Balluffi R, Allen S & Carter W (2005). *Kinetics of Materials*. John Wiley and Sons, Hoboken.
- Behringer EJ & Segal SS (2012). Tuning electrical conduction along endothelial tubes of resistance arteries through Ca²⁺-activated K⁺ channels. *Circ Res* **110**, 1311–1321.
- Behringer EJ & Segal SS (2015). Membrane potential governs calcium influx into microvascular endothelium: Integral role for muscarinic receptor activation. *J Physiol* **593**, 4531–4548.
- Billaud M, Lohman AW, Johnstone SR, Biber LA, Mutchler S & Isakson BE (2014). Regulation of cellular communication by signaling microdomains in the blood vessel wall. *Pharmacol Rev* **66**, 513–569.
- Bortolozzi M, Lelli A & Mammano F (2008). Calcium microdomains at presynaptic active zones of vertebrate hair cells unmasked by stochastic deconvolution. *Cell Calcium* **44**, 158–168.
- Bradley KN, Currie S, MacMillan D, Muir TC & McCarron JG (2003). Cyclic ADP-ribose increases Ca²⁺ removal in smooth muscle. *J Cell Sci* **116**, 4291–4306.
- Bubolz AH, Mendoza SA, Zheng X, Zinkevich NS, Li R, Gutterman DD & Zhang DX (2012). Activation of endothelial TRPV4 channels mediates flow-induced dilation in human coronary arterioles: role of Ca²⁺ entry and mitochondrial ROS signaling. *Am J Physiol Heart Circ Physiol* **302**, H634–642.
- Callamaras N, Marchant JS, Sun XP & Parker I (1998). Activation and co-ordination of InsP₃-mediated elementary Ca²⁺ events during global Ca²⁺ signals in *Xenopus* oocytes. *J Physiol* **509**, 81–91.
- Chataigneau T, Feletou M, Duhault J & Vanhoutte PM (1998a). Epoxyeicosatrienoic acids, potassium channel blockers and endothelium-dependent hyperpolarization in the guinea-pig carotid artery. *Br J Pharmacol* **123**, 574–580.
- Chataigneau T, Feletou M, Thollon C, Villeneuve N, Vilaine JP, Duhault J & Vanhoutte PM (1998b). Cannabinoid CB1 receptor and endothelium-dependent hyperpolarization in guinea-pig carotid, rat mesenteric and porcine coronary arteries. *Br J Pharmacol* **123**, 968–974.
- Chen CS, Mrksich M, Huang S, Whitesides GM & Ingber DE (1997). Geometric control of cell life and death. *Science* **276**, 1425–1428.
- Chien S (2007). Mechanotransduction and endothelial cell homeostasis: the wisdom of the cell. *Am J Physiol Heart Circ Physiol* **292**, H1209–1224.
- Corey DP, Garcia-Anoveros J, Holt JR, Kwan KY, Lin SY, Vollrath MA, Amalfitano A, Cheung EL, Derfler BH, Duggan A, Geleoc GS, Gray PA, Hoffman MP, Rehm HL, Tamasauskas D & Zhang DS (2004). TRPA1 is a candidate for the mechanosensitive transduction channel of vertebrate hair cells. *Nature* **432**, 723–730.
- Craig J & Martin W (2012). Dominance of flow-mediated constriction over flow-mediated dilatation in the rat carotid artery. *Br J Pharmacol* **167**, 527–536.
- Cukierman E, Pankov R, Stevens DR & Yamada KM (2001). Taking cell-matrix adhesions to the third dimension. *Science* **294**, 1708–1712.
- De Bruyn VH, Nuno DW, Cappelli-Bigazzi M, Dole WP & Lamping KG (1994). Effect of acute hypertension in the coronary circulation: role of mechanical factors and oxygen radicals. *J Hypertens* **12**, 163–172.
- de Kreuk BJ, Nethe M, Fernandez-Borja M, Anthony EC, Hensbergen PJ, Deelder AM, Plomann M & Hordijk PL (2011). The F-BAR domain protein PACSIN2 associates with Rac1 and regulates cell spreading and migration. *J Cell Sci* **124**, 2375–2388.
- Deanfield JE, Halcox JP & Rabelink TJ (2007). Endothelial function and dysfunction: testing and clinical relevance. *Circulation* **115**, 1285–1295.
- Dedman A, Sharif-Naeini R, Folgering JH, Duprat F, Patel A & Honore E (2009). The mechano-gated K_{2P} channel TREK-1. *Eur Biophys J* **38**, 293–303.
- Dunn WR, Wellman GC & Bevan JA (1994). Enhanced resistance artery sensitivity to agonists under isobaric compared with isometric conditions. *Am J Physiol Heart Circ Physiol* **266**, H147–155.
- Duza T & Sarelius IH (2004). Localized transient increases in endothelial cell Ca²⁺ in arterioles in situ: implications for coordination of vascular function. *Am J Physiol Heart Circ Physiol* **286**, H2322–2331.
- Earley S & Brayden JE (2015). Transient receptor potential channels in the vasculature. *Physiol Rev* **95**, 645–690.
- Edelstein A, Amodaj N, Hoover K, Vale R & Stuurman N (2010). Computer control of microscopes using microManager. In *Current Protocols in Molecular Biology*, ed. Ausubel MF *et al.*, Ch. 14, Unit 14.20.
- Falcone JC, Kuo L & Meininger GA (1993). Endothelial cell calcium increases during flow-induced dilation in isolated arterioles. *Am J Physiol Heart Circ Physiol* **264**, H653–659.
- Faraci FM & Heistad DD (1990). Regulation of large cerebral arteries and cerebral microvascular pressure. *Circ Res* **66**, 8–17.

- Faraci FM, Orgren K & Heistad DD (1994). Impaired relaxation of the carotid artery during activation of ATP-sensitive potassium channels in atherosclerotic monkeys. *Stroke* **25**, 178–182.
- Fischer MJ, Uchida S & Messlinger K (2010). Measurement of meningeal blood vessel diameter in vivo with a plug-in for ImageJ. *Microvasc Res* **80**, 258–266.
- Flusberg BA, Nimmerjahn A, Cocker ED, Mukamel EA, Barretto RP, Ko TH, Burns LD, Jung JC & Schnitzer MJ (2008). High-speed, miniaturized fluorescence microscopy in freely moving mice. *Nat Methods* **5**, 935–938.
- Francis M, Qian X, Charbel C, Ledoux J, Parker JC & Taylor MS (2012). Automated region of interest analysis of dynamic Ca^{2+} signals in image sequences. *Am J Physiol Cell Physiol* **303**, C236–243.
- Gunduz F, Meiselman HJ & Baskurt OK (2008). High intravascular pressure attenuates vascular dilation responses of small mesenteric arteries in the rat. *Circ J* **72**, 482–486.
- Hartmannsgruber V, Heyken WT, Kacik M, Kaistha A, Grgic I, Harteneck C, Liedtke W, Hoyer J & Kohler R (2007). Arterial response to shear stress critically depends on endothelial TRPV4 expression. *PLoS One* **2**, e827.
- Hishikawa K, Nakaki T, Suzuki H, Saruta T & Kato R (1992). Transmural pressure inhibits nitric oxide release from human endothelial cells. *Eur J Pharmacol* **215**, 329–331.
- Hoffman BD, Grashoff C & Schwartz MA (2011). Dynamic molecular processes mediate cellular mechanotransduction. *Nature* **475**, 316–323.
- Hong D, Jaron D, Buerk DG & Barbee KA (2008). Transport-dependent calcium signaling in spatially segregated cellular caveolar domains. *Am J Physiol Cell Physiol* **294**, C856–866.
- Huang A, Sun D, Kaley G & Koller A (1998). Superoxide released to high intra-arteriolar pressure reduces nitric oxide-mediated shear stress- and agonist-induced dilations. *Circ Res* **83**, 960–965.
- Huang TY, Chu TF, Chen HI & Jen CJ (2000). Heterogeneity of $[Ca^{2+}]_i$ signaling in intact rat aortic endothelium. *FASEB J* **14**, 797–804.
- Janssen DA, Hoenderop JG, Jansen KC, Kemp AW, Heesakkers JP & Schalken JA (2011). The mechanoreceptor TRPV4 is localized in adherence junctions of the human bladder urothelium: a morphological study. *J Urol* **186**, 1121–1127.
- Jena M, Minore JF & O'Neill WC (1997). A volume-sensitive, IP_3 -insensitive Ca^{2+} store in vascular endothelial cells. *Am J Physiol Cell Physiol* **273**, C316–322.
- Jurva JW, Phillips SA, Syed AQ, Syed AY, Pitt S, Weaver A & Gutterman DD (2006). The effect of exertional hypertension evoked by weight lifting on vascular endothelial function. *J Am Coll Cardiol* **48**, 588–589.
- Kim P, Chung E, Yamashita H, Hung KE, Mizoguchi A, Kucherlapati R, Fukumura D, Jain RK & Yun SH (2010). In vivo wide-area cellular imaging by side-view endomicroscopy. *Nat Methods* **7**, 303–305.
- Knudsen HL & Frangos JA (1997). Role of cytoskeleton in shear stress-induced endothelial nitric oxide production. *Am J Physiol Heart Circ Physiol* **273**, H347–355.
- Kohler R, Heyken WT, Heinau P, Schubert R, Si H, Kacik M, Busch C, Grgic I, Maier T & Hoyer J (2006). Evidence for a functional role of endothelial transient receptor potential V4 in shear stress-induced vasodilatation. *Arterioscler Thromb Vasc Biol* **26**, 1495–1502.
- Kusche-Vihrog K, Jeggle P & Oberleithner H (2014). The role of ENaC in vascular endothelium. *Pflügers Arch* **466**, 851–859.
- Lamping KG & Faraci FM (2001). Role of sex differences and effects of endothelial NO synthase deficiency in responses of carotid arteries to serotonin. *Arterioscler Thromb Vasc Biol* **21**, 523–528.
- Ledoux J, Taylor MS, Bonev AD, Hannah RM, Solodushko V, Shui B, Tallini Y, Kotlikoff MI & Nelson MT (2008). Functional architecture of inositol 1,4,5-trisphosphate signaling in restricted spaces of myoendothelial projections. *Proc Natl Acad Sci USA* **105**, 9627–9632.
- Li J, Hou B, Tumova S, Muraki K, Bruns A, Ludlow MJ, Sedo A, Hyman AJ, McKeown L, Young RS, Yuldasheva NY, Majeed Y, Wilson LA, Rode B, Bailey MA, Kim HR, Fu Z, Carter DA, Bilton J, Imrie H, Ajuh P, Dear TN, Cubbon RM, Kearney MT, Prasad RK, Evans PC, Ainscough JF & Beech DJ (2014). Piezo1 integration of vascular architecture with physiological force. *Nature* **515**, 279–282.
- Loot AE, Popp R, Fisslthaler B, Vriens J, Nilius B & Fleming I (2008). Role of cytochrome P450-dependent transient receptor potential V4 activation in flow-induced vasodilatation. *Cardiovasc Res* **80**, 445–452.
- Loukin S, Zhou X, Su Z, Saimi Y & Kung C (2010). Wild-type and brachyolmia-causing mutant TRPV4 channels respond directly to stretch force. *J Biol Chem* **285**, 27176–27181.
- Luscher TF & Vanhoutte PM (1986). Endothelium-dependent contractions to acetylcholine in the aorta of the spontaneously hypertensive rat. *Hypertension* **8**, 344–348.
- McCarron JG, Chalmers S, MacMillan D & Olson ML (2010). Agonist-evoked Ca^{2+} wave progression requires Ca^{2+} and IP_3 . *J Cell Physiol* **224**, 334–344.
- McGeown JG, Drummond RM, McCarron JG & Fay FS (1996). The temporal profile of calcium transients in voltage clamped gastric myocytes from *Bufo marinus*. *J Physiol* **497**, 321–336.
- Marchenko SM & Sage SO (2000). Effects of shear stress on $[Ca^{2+}]_i$ and membrane potential of vascular endothelium of intact rat blood vessels. *Exp Physiol* **85**, 43–48.
- Maroto R, Raso A, Wood TG, Kurosky A, Martinac B & Hamill OP (2005). TRPC1 forms the stretch-activated cation channel in vertebrate cells. *Nat Cell Biol* **7**, 179–185.
- Mazel T, Raymond R, Raymond-Stintz M, Jett S & Wilson BS (2009). Stochastic modeling of calcium in 3D geometry. *Biophys J* **96**, 1691–1706.
- Mchedlishvili G (1986). *Arterial Behavior and Blood Circulation in the Brain*. Plenum Publishing Corp, New York.
- Means S, Smith AJ, Shepherd J, Shadid J, Fowler J, Wojcikiewicz RJ, Mazel T, Smith GD & Wilson BS (2006). Reaction diffusion modeling of calcium dynamics with realistic ER geometry. *Biophys J* **91**, 537–557.
- Mederos Y, Schnitzler M, Storch U, Meibers S, Nurwakagari P, Breit A, Essin K, Gollasch M & Gudermann T (2008). Gq-coupled receptors as mechanosensors mediating myogenic vasoconstriction. *EMBO J* **27**, 3092–3103.

- Mendoza SA, Fang J, Gutterman DD, Wilcox DA, Bubolz AH, Li R, Suzuki M & Zhang DX (2010). TRPV4-mediated endothelial Ca²⁺ influx and vasodilation in response to shear stress. *Am J Physiol Heart Circ Physiol* **298**, H466–476.
- Moccia F, Berra-Romani R & Tanzi F (2012). Update on vascular endothelial Ca²⁺ signalling: A tale of ion channels, pumps and transporters. *World J Biol Chem* **3**, 127–158.
- Mossman KD, Campi G, Groves JT & Dustin ML (2005). Altered TCR signaling from geometrically repatterned immunological synapses. *Science* **310**, 1191–1193.
- Muller JM, Davis MJ, Kuo L & Chilian WM (1999). Changes in coronary endothelial cell Ca²⁺ concentration during shear stress- and agonist-induced vasodilation. *Am J Physiol Heart Circ Physiol* **276**, H1706–1714.
- Ohashi M, Faraci F & Heistad D (2005). Peroxynitrite hyperpolarizes smooth muscle and relaxes internal carotid artery in rabbit via ATP-sensitive K⁺ channels. *Am J Physiol Heart Circ Physiol* **289**, H2244–2250.
- Olson ML, Chalmers S & McCarron JG (2010). Mitochondrial Ca²⁺ uptake increases Ca²⁺ release from inositol 1,4,5-trisphosphate receptor clusters in smooth muscle cells. *J Biol Chem* **285**, 2040–2050.
- Olson ML, Sandison ME, Chalmers S & McCarron JG (2012). Microdomains of muscarinic acetylcholine and Ins(1,4,5)P₃ receptors create 'Ins(1,4,5)P₃ junctions' and sites of Ca²⁺ wave initiation in smooth muscle. *J Cell Sci* **125**, 5315–5328.
- Paniagua OA, Bryant MB & Panza JA (2000). Transient hypertension directly impairs endothelium-dependent vasodilation of the human microvasculature. *Hypertension* **36**, 941–944.
- Patel AJ, Lazdunski M & Honore E (2001). Lipid and mechano-gated 2P domain K⁺ channels. *Curr Opin Cell Biol* **13**, 422–428.
- Phillips SA, Das E, Wang J, Pritchard K & Gutterman DD (2011). Resistance and aerobic exercise protects against acute endothelial impairment induced by a single exposure to hypertension during exertion. *J Appl Physiol* **110**, 1013–1020.
- Plane F, Wiley KE, Jeremy JY, Cohen RA & Garland CJ (1998). Evidence that different mechanisms underlie smooth muscle relaxation to nitric oxide and nitric oxide donors in the rabbit isolated carotid artery. *Br J Pharmacol* **123**, 1351–1358.
- Popp R, Fleming I & Busse R (1998). Pulsatile stretch in coronary arteries elicits release of endothelium-derived hyperpolarizing factor: a modulator of arterial compliance. *Circ Res* **82**, 696–703.
- Qi H, Li L & Shuai J (2015). Optimal microdomain crosstalk between endoplasmic reticulum and mitochondria for Ca²⁺ oscillations. *Sci Rep* **5**, 7984.
- Qian X, Francis M, Solodushko V, Earley S & Taylor MS (2013). Recruitment of dynamic endothelial Ca²⁺ signals by the TRPA1 channel activator AITC in rat cerebral arteries. *Microcirculation* **20**, 138–148.
- Ranade SS, Qiu Z, Woo SH, Hur SS, Murthy SE, Cahalan SM, Xu J, Mathur J, Bandell M, Coste B, Li YS, Chien S & Patapoutian A (2014). Piezo1, a mechanically activated ion channel, is required for vascular development in mice. *Proc Natl Acad Sci USA* **111**, 10347–10352.
- Rizzuto R, Pinton P, Carrington W, Fay FS, Fogarty KE, Lifshitz LM, Tuft RA & Pozzan T (1998). Close contacts with the endoplasmic reticulum as determinants of mitochondrial Ca²⁺ responses. *Science* **280**, 1763–1766.
- Saunter CD, Semprini S, Buckley C, Mullins J & Girkin JM (2012). Micro-endoscope for in vivo widefield high spatial resolution fluorescent imaging. *Biomed Opt Express* **3**, 1274–1278.
- Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, Saalfeld S, Schmid B, Tinevez JY, White DJ, Hartenstein V, Eliceiri K, Tomancak P & Cardona A (2012). Fiji: an open-source platform for biological-image analysis. *Nat Methods* **9**, 676–682.
- Sheetz MP (1993). Glycoprotein motility and dynamic domains in fluid plasma membranes. *Annu Rev Biophys Biomol Struct* **22**, 417–431.
- Shiu YT, Li S, Marganski WA, Usami S, Schwartz MA, Wang YL, Dembo M & Chien S (2004). Rho mediates the shear-enhancement of endothelial cell migration and traction force generation. *Biophys J* **86**, 2558–2565.
- Smith GD (1996). Analytical steady-state solution to the rapid buffering approximation near an open Ca²⁺ channel. *Biophys J* **71**, 3064–3072.
- Socha MJ, Behringer EJ & Segal SS (2012a). Calcium and electrical signalling along endothelium of the resistance vasculature. *Basic Clin Pharmacol Toxicol* **110**, 80–86.
- Socha MJ, Domeier TL, Behringer EJ & Segal SS (2012b). Coordination of intercellular Ca²⁺ signaling in endothelial cell tubes of mouse resistance arteries. *Microcirculation* **19**, 757–770.
- Sonkusare SK, Bonev AD, Ledoux J, Liedtke W, Kotlikoff MI, Heppner TJ, Hill-Eubanks DC & Nelson MT (2012). Elementary Ca²⁺ signals through endothelial TRPV4 channels regulate vascular function. *Science* **336**, 597–601.
- Spassova MA, Hewavitharana T, Xu W, Soboloff J & Gill DL (2006). A common mechanism underlies stretch activation and receptor activation of TRPC6 channels. *Proc Natl Acad Sci USA* **103**, 16586–16591.
- Strotmann R, Harteneck C, Nunnenmacher K, Schultz G & Plant TD (2000). OTRPC4, a nonselective cation channel that confers sensitivity to extracellular osmolarity. *Nat Cell Biol* **2**, 695–702.
- Strotmann R, Schultz G & Plant TD (2003). Ca²⁺-dependent potentiation of the nonselective cation channel TRPV4 is mediated by a C-terminal calmodulin binding site. *J Biol Chem* **278**, 26541–26549.
- Sun D, Huang A, Recchia FA, Cui Y, Messina EJ, Koller A & Kaley G (2001). Nitric oxide-mediated arteriolar dilation after endothelial deformation. *Am J Physiol Heart Circ Physiol* **280**, H714–721.
- Tanko LB, Mikkelsen EO & Simonsen U (1999). A new experimental approach in endothelium-dependent pharmacological investigations on isolated porcine coronary arteries mounted for impedance planimetry. *Br J Pharmacol* **128**, 165–173.

- Thi MM, Tarbell JM, Weinbaum S & Spray DC (2004). The role of the glycocalyx in reorganization of the actin cytoskeleton under fluid shear stress: a “bumper-car” model. *Proc Natl Acad Sci USA* **101**, 16483–16488.
- Toda N, Inoue S, Okunishi H & Okamura T (1990). Intra- and extraluminally-applied acetylcholine on the vascular tone or the response to transmural stimulation in dog isolated mesenteric arteries. *Naunyn Schmiedebergs Arch Pharmacol* **341**, 30–36.
- Toda N, Minami Y & Onoue H (1988). Extraluminally applied acetylcholine and oxyhemoglobin on the release and action of EDRF. *Eur J Pharmacol* **151**, 123–126.
- Tran CH, Taylor MS, Plane F, Nagaraja S, Tsoukias NM, Solodushko V, Vigmond EJ, Furstenhaupt T, Brigdan M & Welsh DG (2012). Endothelial Ca²⁺ wavelets and the induction of myoendothelial feedback. *Am J Physiol Cell Physiol* **302**, C1226–1242.
- Tu H, Wang Z, Nosyreva E, De Smedt H & Bezprozvany I (2005). Functional characterization of mammalian inositol 1,4,5-trisphosphate receptor isoforms. *Biophys J* **88**, 1046–1055.
- Tzima E, Irani-Tehrani M, Kiosses WB, Dejana E, Schultz DA, Engelhardt B, Cao G, DeLisser H & Schwartz MA (2005). A mechanosensory complex that mediates the endothelial cell response to fluid shear stress. *Nature* **437**, 426–431.
- Vecchione C, Carnevale D, Di Pardo A, Gentile MT, Damato A, Cocozza G, Antenucci G, Mascio G, Bettarini U, Landolfi A, Iorio L, Maffei A & Lembo G (2009). Pressure-induced vascular oxidative stress is mediated through activation of integrin-linked kinase 1/ β PIX/Rac-1 pathway. *Hypertension* **54**, 1028–1034.
- Wallace SM, Yasmin, McEniery CM, Maki-Petaja KM, Booth AD, Cockcroft JR & Wilkinson IB (2007). Isolated systolic hypertension is characterized by increased aortic stiffness and endothelial dysfunction. *Hypertension* **50**, 228–233.
- Yamamoto K, Sokabe T, Matsumoto T, Yoshimura K, Shibata M, Ohura N, Fukuda T, Sato T, Sekine K, Kato S, Isshiki M, Fujita T, Kobayashi M, Kawamura K, Masuda H, Kamiya A & Ando J (2006). Impaired flow-dependent control of vascular tone and remodeling in P2X4-deficient mice. *Nat Med* **12**, 133–137.
- Yeung T, Georges PC, Flanagan LA, Marg B, Ortiz M, Funaki M, Zahir N, Ming W, Weaver V & Janmey PA (2005). Effects of substrate stiffness on cell morphology, cytoskeletal structure, and adhesion. *Cell Motil Cytoskeleton* **60**, 24–34.
- Zhao H, Pykalainen A & Lappalainen P (2011). I-BAR domain proteins: linking actin and plasma membrane dynamics. *Curr Opin Cell Biol* **23**, 14–21.
- Zhao Y, Flavahan S, Leung SW, Xu A, Vanhoutte PM & Flavahan NA (2015). Elevated pressure causes endothelial dysfunction in mouse carotid arteries by increasing local angiotensin signaling. *Am J Physiol Heart Circ Physiol* **308**, H358–363.
- Zimmerberg J & Kozlov MM (2006). How proteins produce cellular membrane curvature. *Nat Rev Mol Cell Biol* **7**, 9–19.

Additional information

Competing interests

None.

Author contributions

All authors have approved the final version of the manuscript and agree to be accountable for all aspects of the work. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed. The manuscript was prepared and written with contributions from all authors. All authors approved the final version of the manuscript.

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Supporting information

The following supporting information is available in the online version of this article.

Movie S1. ACh-evoked endothelial Ca²⁺ signals in a pressurised (60 mmHg) rat carotid artery. Activation of IP₃-evoked endothelial Ca²⁺ signals, by bath application of ACh (100 μ M), causes a rise in cytoplasmic Ca²⁺ concentration in the majority of cells across the field of view. The movie is composed of raw data, to which a 5-frame rolling average has been applied, that has been linearly contrast adjusted for visualisation. Data was acquired at 5 Hz and the scale bar is 100 μ m.

Movie S2. ACh activated IP₃-evoked endothelial Ca²⁺ signals which were recorded from inside pressurized arteries. At 60 mmHg, ACh caused an initial burst of Ca²⁺ increases that propagate as waves. As ACh-evoked Ca²⁺ waves expanded collision between adjacent cells occurred and wave annihilation occurred. As a result of multiple waves progressions and annihilation events, complex spatiotemporal patterns of Ca²⁺ signaling developed. The movie is composed of a sequential subtraction of the

temporally smoothed Ca²⁺ activity (green) overlaid on standard deviation image (grayscale). Data was acquired at 5 Hz and the scale bar is 100 μm . **Movie S3.** ACh activated IP₃-evoked Ca²⁺ signals were significantly attenuated as the artery transmural pressure increased from 60 mmHg to 110 mmHg to 160 mmHg. After each application of ACh the bath solution was washed with >20 times the bath volume, and the artery

was allowed to re-equilibrate for 20 minutes before the next pressure change and ACh addition. The movie corresponds to Ca²⁺ traces and time-series data shown in Figure 7A and Figure 7B respectively. The movie is a time series of Ca²⁺ wave activity (green) overlaid on standard deviation images (STDev) (grayscale). Note that STDev images only show cells that exhibit Ca²⁺ activity. Data was acquired at 5 Hz and the scale bar corresponds to 100 μm .