

1 **A mitochondrial genetic divergence proxy predicts the reproductive**
2 **compatibility of mammalian hybrids**

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33

34 **Abstract**

35 Despite the significant genetic barriers that arise during the process of reproductive
36 isolation between species, numerous hybrid offspring of divergent mammalian
37 species pairs have been catalogued. In some cases, these hybrids are only able to
38 produce offspring through backcrosses with a parent species due to unisexual
39 sterility (Haldane's Rule). In other instances, F1 hybrids are able to produce F2s
40 through matings with F1s. Here, we explicitly tested whether genetic distance can be
41 used as a proxy to predict the relative fertility of the hybrid offspring resulting from
42 numerous pairs of terrestrial mammals. We assessed the proxy's predictive power
43 using a well characterised felid hybrid system, and applied it to modern and ancient
44 hominins. Our results revealed a small overlap in mitochondrial genetic divergence
45 values that distinguish species pairs whose divergence values fall within two
46 categories: those whose hybrid offspring follow Haldane's Rule, and those whose
47 hybrid offspring can produce F2s. The strong correlation between genetic distance
48 and hybrid fertility demonstrated here suggests that this proxy can be employed to
49 predict whether the hybrid offspring of two mammalian species will follow Haldane's
50 Rule.

51

52 **Introduction**

53 Though hybrids between mammalian species have been catalogued for decades [1],
54 the extent and frequency of gene flow between evolutionarily divergent taxa has only
55 been recognised since the availability of high-coverage nuclear genomes. Recent
56 studies have revealed rampant gene flow between multiple species of bears[2],
57 canids[3], felids[4–6], and cetaceans[7,8]. Genome analyses of other vertebrate and
58 invertebrate lineages beyond mammals have also revealed similarly extensive
59 patterns of ancient and contemporary introgression [9–14].

60

61 This demonstrated frequency of genomic introgression is perhaps surprising given
62 the significant barriers that maintain reproductive isolation in species pairs that
63 diverged up to millions of years ago. In mammals, genomic barriers manifest in
64 accordance with Haldane's Rule [15] as the unisexual sterility of the heterogametic
65 sex (XY males) in F1 hybrid offspring. In cases where matings between F1s fail to
66 produce F2s, fertile offspring can often be produced through backcrosses between
67 the fertile F1 females and males from one of the parent species. Occasionally,

68 however, F1s produced from interbreeding between closely related mammalian
69 species pairs can result in viable and fertile F2 offspring.

70

71 If genetic divergence values between two species correlated with the ability of their
72 hybrid offspring to produce F2s, these values could serve as a proxy to predict this
73 occurrence. Though at least one study [16] reported that genetic divergence values
74 do generally correlate with species boundaries, others [17,18] have questioned
75 whether this correlation exists, and have instead stated that measures of species
76 divergence are not reliable predictors of hybrid sterility. A recent empirical study of
77 damselflies, however, demonstrated a strong correlation between the genetic
78 distances between species pairs and their relative reproductive isolation [19].

79

80 Establishing whether genetic distance and reproductive isolation are correlated is
81 also critical for our understanding of the genetic architecture of reproductive
82 isolation. Doing so firstly requires knowing whether any two species are capable of
83 producing viable or fertile offspring, but there is a general paucity of captive breeding
84 experiments or field data that have unequivocally established this. An alternative
85 approach is to develop a metric that can accurately predict the relative fertility of the
86 F1 hybrids of any two species that makes use of interspecific crosses whose
87 offspring have been reproductively characterized. Here we developed a robust,
88 quantitative framework based on the correlation between mitochondrial genetic
89 distance between mammalian species known to produce F1spairs to obtain a
90 quantitative measure of whether F1 hybrids of both sexes are likely to be capable of
91 breeding, or if they instead manifest Haldane's Rule. We tested the accuracy of the
92 proxy in a well characterised felid hybrid system, and then applied it to a hominin
93 case study to assess the relative potential sterility of hybrids between humans and
94 their closest extinct relatives.

95

96 **Results and Discussion**

97 ***Categorizing Hybrid Incompatibility***

98 We first explicitly defined two dichotomous categories along the spectrum of hybrid
99 compatibility. Category 1 is defined by mammalian species pairs capable of
100 producing fertile F1 offspring of both sexes that can reproduce without backcrossing
101 with a parent species (even if there are observed asymmetries in gene flow and

102 variation in male fertility amongst the hybrids) (Table S1). Category 2 is defined by
103 pairs of species that can produce viable F1 offspring, but follow Haldane's Rule, and
104 thus only female F1s can reproduce by backcrossing with a parent species.
105 Category 2 also includes species pairs whose hybrids are infertile (Table S1). We
106 determined the categorical assignment of each species pair (Table S1, Fig. S1) by
107 following a flowchart (Fig. S2) based upon empirical evidence derived from
108 experimental studies of F1 hybrid fertility. We confidently placed seven species pairs
109 into Category 1, and six others into Category 2.

110

111 Many additional live hybrid offspring have been reported in the literature than are
112 included in Fig. 1 or in Fig. S1. We identified 17 species pairs known to produce
113 viable offspring, but for which there was insufficient evidence to confidently assign
114 them into either category (Table S2, Fig. S3). The framework and threshold values
115 depicted in Fig. 1 allow us to predict the fertility of these offspring given the
116 definitions described above and their placement into Categories 1 or 2. These pairs
117 are listed in Table S2 and their relative positions are depicted in Fig. S3.

118

119 We then obtained published sequence data across all species (Table S3) from both
120 the cytochrome b gene (*CYTB*) (n=1795) and complete mitochondrial genomes
121 (n=30) (excluding the control region) from multiple individuals per species. By
122 matching the phylogenies derived from the alignments to available nuclear species
123 trees, and by only including sequences that fell into reciprocally monophyletic clades,
124 we ensured that the selected mitochondrial sequences for each species were neither
125 mislabelled, nor nuclear copies of mitochondrial genes (NUMTs), nor derived from
126 hybrid populations. Using the sequence alignments, we then calculated average
127 pairwise genetic distances between each species pair using both the number of raw
128 differences, and differences scaled by several different nucleotide substitution
129 models. In addition, we calculated genetic distances for four nuclear loci (*CHRNA1*,
130 *GHR*, *ZFX*, *ZFY*) available between ten of the primate species pairs to test the
131 correlation between the mitochondrial proxy and the nuclear genome estimates [20]
132 (Fig. S4).

133

134 Plotting the calculated divergence estimates using *CYTB* revealed a small overlap in
135 values associated with the two categories (Fig 1, Fig. S1). More specifically, the

136 Category 1 pair with the greatest divergence value was a pair of guinea pig species
137 that were 8.0% divergent, and the Category 2 pair with the lowest divergence was a
138 pair of vole species that were 7.2% divergent. Within this 0.8% overlapping region
139 were several pairs of species that fall into both categories suggesting that this level
140 of CYTB divergence is the region where F1 offspring begin to require a backcross to
141 generate an F2. The existence of a genetic distance threshold separating the two
142 categories also held true for the complete mitogenomes (Fig. S4).

143

144 In addition, both the male and female hybrid offspring of one of the two most
145 divergent pairs, Domestic Pig (*Sus domesticus*) x Babirusa (*Babirusa celebensis*),
146 were shown to be infertile, and an exhaustive study that attempted to produce
147 hybrids between mountain hares and European rabbits failed completely [21]. The
148 significant divergence values between these species pairs corroborates previous
149 studies showing that along the continuum of speciation, infertility in both sexes
150 evolves prior to inviability [22–24]. In addition, a Mann-Whitney U test showed
151 significantly lower genetic divergence values of species pairs in Category 1 relative
152 to those in Category 2 ($p < 6.3e-4$).

153

154 Importantly, the two categories of fertility defined here are not strictly linked with
155 gene flow. For instance, though both male and female Category 1 hybrid offspring
156 can reproduce without requiring a backcross with a parent species, gene flow
157 asymmetries have been demonstrated in virtually all of these species pairs including
158 house mice (*Mus musculus musculus* x *Mus musculus domesticus*) [25], and
159 between brown bears (*Ursus arctos*) and polar bears (*Ursus maritimus*) [2] (Table
160 S1). Gene flow has also been demonstrated between Category 2 species (including
161 *Mus musculus* and *M. spretus* [26]). Since both fertility and the potential for gene
162 flow vary along a continuum, it is striking that the divergence values associated with
163 the two fertility categories defined here do not overlap more substantially.

164

165 The lack of available nuclear sequences (relative to mitochondria) limited our ability
166 to test whether nuclear genes generally produced the same pattern as the
167 mitochondria across all our species pairs. Despite this limitation, we were able to
168 identify four nuclear loci: Zinc finger Y-chromosomal protein (*ZFY*), Zinc finger X-
169 chromosomal protein (*ZFX*), Growth Hormone Receptor (*GHR*), Cholinergic

170 Receptor Nicotinic Alpha (*CHRNA1*), that have been sequenced in 10 primate pairs
171 (Fig. S4) known to produce viable hybrid offspring [20]. We generated pairwise
172 distances for each of these genes using the same method employed in the
173 mitochondrial distance calculation. We then assigned each species pair to Category
174 1 or Category 2 based upon their *CYTB* divergence values within the original
175 framework. In each case, though the order of the taxa based upon pairwise
176 divergence values varied relative to the pattern generated using *CYTB* (owing to
177 significantly fewer variable sites and thus smaller divergence values in nuclear loci),
178 for the two most variable nuclear loci, *ZFX* and *GHR*, there was no overlap in
179 divergence values between the two categories, consistent with the mitochondrial
180 assessment (Fig. S4). The other two loci, *ZFY* and *CHRNA1*, possessed very limited
181 interspecific nucleotide variability, but generally followed the same overall pattern.

182

183 ***Testing the proxy using known mammalian hybrids***

184 In order to further substantiate both this correlation and the robustness of *CYTB*
185 divergence as a proxy for hybrid sterility, we tested the utility of this system for
186 predicting fertility in a well-known hybrid system. Cat breeders have crossed
187 domestic cats (*Felis catus*) with several wild felids, including the Jungle cat (*F.*
188 *chaus*), Leopard cat (*Prionailurus bengalensis*), and Serval (*Leptailurus serval*) [27],
189 to create three exotic cat breeds: Chausies, Bengals, and Savannahs, respectively.
190 In all cases, the F1 male hybrids are sterile. To regain fertility while maintaining
191 some wild felid characteristics, breeders must backcross the F1 female offspring with
192 male domestic cats to establish a breeding population of pets [27]. Given that
193 multiple generations of unidirectional backcrossing was required for all three crosses
194 to generate a fertile population, our proxy would firstly predict that the *CYTB*
195 distances between all three pairs should be close to or greater than ~7.2%, and that
196 they should all fall into the range encompassed by Category 2. Secondly, the pairs
197 with larger genetic distance values should require a greater number of backcrosses
198 with domestic cats (halving the wild cat ancestry with each subsequent generation)
199 before fertility is restored in hybrid males and a breeding pet population is
200 established.

201

202 Both of these predictions are borne out by the data (Fig 1, Table S3). All three pairs
203 show *CYTB* distances greater than or equal to 7.5% and the increasing molecular

204 distances between the pairs correlate with an increase in the number of required
205 backcross generations to regain fertility. Specifically, distances between domestic
206 cats and Jungle cats, Leopard cats and Serval (7.54%, 10.94%, and 11.28%
207 respectively) are consistent with both the observed minimum (2, 3, and 4
208 respectively) and average (3, 4, and 5, respectively) number of backcrosses with
209 domestic cats required for hybrid males to acquire fertility [27]. These results are
210 also consistent with an early hybrid experiment using guinea pigs in which hybrids
211 between *Cavia fulgida* and *C. porcellus* (8.0% *CYTB* divergence) were able to regain
212 male fertility after three generations of backcrossing [28](Table S1).

213

214 Accidental hybrids in zoos also confirm the predictive power of this proxy. In 2006,
215 the Copenhagen Zoo placed a domestic sow (*Sus domesticus*) in a pen with a male
216 babirusa (*Babirusa celebensis*) with the expectation that the two species were
217 sufficiently evolutionarily divergent that they would be incapable of producing
218 offspring. Months later, however, five piglets were born and though two died from
219 maternally induced trauma, the other three (two males, and one female), all survived
220 and were shown to be infertile [29] (Table S1, Fig. S1). Historically, hybrid offspring
221 between distantly related species have accidentally been produced in zoos (Table
222 S2), though the relative fertility of the F1s was rarely established. In this case, the
223 *CYTB* divergence between the two species (12.9%) is not much greater than the
224 value between rhesus macaques and hamadryas baboons (12.5%) which were able
225 to produce a live offspring, thus suggesting that live offspring between these suids
226 was possible.

227

228 ***Assessing hominin hybrid incompatibility***

229 The initial discovery of Neanderthals led some anthropologists, as early as the turn
230 of the 20th century, to speculate that anatomically modern humans (AMH) and their
231 closest extinct relatives were capable of producing hybrid offspring [30]. The
232 absence of Neanderthal mitochondrial genomes in the extant human population,
233 however, led some to suggest that AMH and Neanderthals did not hybridise [31–33].
234 More recent analyses of whole ancient genome sequences have demonstrated that,
235 in fact, archaic hominins, including Neanderthals and Denisovans, did produce
236 hybrid offspring, not only with AMH [34–36], but also with each other [37]. The
237 generation of these ancient genomes has also allowed for an assessment of the role

238 that incompatibility may have played in the selection for and against hybrid
239 introgression in modern humans [38]. The genomic confirmation of the existence of
240 hominin hybrids supported the conclusions of two studies [39,40] that used a
241 qualitative correlation between the divergence times between species pairs and the
242 fertility of their hybrid offspring to suggest that, given their relatively recent temporal
243 divergence, AMH and Neanderthals could have retained the ability to produce fertile
244 offspring of both sexes.

245

246 We quantitatively assessed the relative fertility of hybrids between pairs of modern
247 and ancient hominin lineages using the proxy established in this study. To do so, we
248 calculated the average pairwise divergence in *CYTB* sequences between AMH and
249 three extinct hominin lineages: Neanderthals, Denisovans, and the ancient
250 population from the Sima de los Huesos cave in Spain [29,41]. To avoid
251 overestimating the genetic divergence resulting from the comparison of modern and
252 extinct populations, we generated distance values using the *CYTB* sequences
253 derived solely from ancient AMH found in archaeological contexts (Supp Table 3).

254

255 The divergence values for each pairing of three *Homo* groups (Sima de los Huesos,
256 Neanderthals, and AMH) occupy the bottom of the Category 1 range and are less
257 than the divergence values between Polar Bears and Brown Bears, and between
258 subspecific crosses of *Mus musculus* (Fig 1, Fig. S1, Table S1). When placed within
259 this context, our data predict that ancient hominin lineages were likely not sufficiently
260 divergent from each other to expect a significant biological impediment to the
261 generation of fertile offspring. This is consistent with the ancient genomic evidence,
262 which has shown not only that archaic populations interbred with AMH on at least
263 four occasions [42], but also that introgression took place in both directions [43]. In
264 addition, the divergence values of Denisovan-Neanderthal and Denisovan-AMH are
265 the largest of the *Homo* pairings, and are consistent with the suggestion that
266 Denisovans possessed a mitochondrial lineage that may have introgressed from
267 another source population [44].

268

269 We also assessed hybrid sterility between more distantly diverged hominin lineages.
270 Specifically, we calculated divergence values between humans and our two closest
271 living relatives: chimpanzees (*Pan troglodytes*) and bonobos (*P. paniscus*). Female

272 chimpanzees inseminated with human sperm during a Soviet experiment in the
273 1920s failed to produce any offspring, and the reverse experiment did not progress
274 beyond the planning stage [45]. Recent molecular clock assessments have
275 suggested that AMH and chimps diverged ~5-6 Mya [46], well beyond both the two-
276 million-year threshold suggested by other studies as the upper limit to hybrid fertility
277 [39,40], and the average time to speciation [47]. Our analysis places the divergence
278 values between AMH and chimps, and AMH and bonobos within Category 2
279 suggesting that even if hybrids could be produced, they would likely follow Haldane's
280 Rule (Fig 1, Fig S1).

281

282 **Conclusions**

283 The correlation demonstrated here between *CYTB* divergence (but also genetic
284 divergence in general) and relative hybrid sterility suggests that distance values can
285 be used as a proxy to accurately and rapidly predict the relative sterility of hybrids
286 resulting from matings between pairs of mammalian species. Our emphasis here on
287 mitochondrial DNA should not be misinterpreted as a claim that *CYTB* plays a
288 causative role in hybrid sterility, though recent studies have proposed that speciation
289 may be mediated by mitonuclear interactions [48,49]. Nor can the use of genetic
290 divergence values as a proxy be perfectly predictive. For example, under the
291 Dobzhansky-Müller model incompatibility can arise from as few as two mutations in
292 isolated populations irrespective of time since divergence, meaning that it would be
293 possible for closely related populations to be incapable of generating fertile hybrids
294 [48,50], though no such examples have been described.

295

296 The value of any proxy is determined by both its predictive power and the ease of
297 generating the proxy data. Publicly available mtDNA sequences from thousands of
298 mammalian taxa already exist and calculating pairwise divergence values is
299 inexpensive, simple and fast. As a result, mitogenomic distances have substantial
300 value as a means to predict the potential for any two mammalian species to produce
301 fertile offspring, and the relative degree of sterility in one or both sexes. As whole
302 genomes become available from the same set of species, this analysis can be
303 extended to determine which regions of the nuclear genome may also be more or
304 less predictive.

305

306 The discovery of additional extinct hominin populations that survived into the last
307 250,000 years, including *H. floresiensis* [51] and *H. naledi* [52], has raised interest in
308 understanding the limits to fertility and hybridization between extinct and extant
309 *Homo* spp [53]. If and when mitochondrial genomes from these samples can be
310 obtained, the approach described here may provide an answer, even if nuclear
311 genomic data are not obtainable. Lastly, establishing which species pairs violate the
312 predictions of the framework will identify unique systems that may lead to a better
313 understanding of the process of reproductive isolation and the biological
314 mechanisms responsible for hybrid sterility.

315

316 **Materials and Methods**

317 ***Assessment of Hybrid Fertility and Rationale of Assignment into Categories***

318 In order to ascertain if there was a correlation between genetic divergence and the
319 fertility of hybrid offspring between species, we first collected published examples of
320 species pairs that were capable of producing live offspring. We then split the hybrid
321 pairings into two categories. Category one consisted of seven species pairs that are
322 capable of producing fertile F1 offspring of both sexes, and for which we were able to
323 obtain evidence of captive breeding experiments showing that the F1s could mate to
324 produce F2s. The evidence and rationale for placing each of these pairs into
325 Category 1 is listed in Table S1 and the flow chart we used to determine the
326 categorisation is shown in Table S2.

327

328 The hybrid offspring of all of six pairs of species in Category 2 are either completely
329 infertile, or require one or more generations of female hybrid backcrosses with the
330 male of a parent species to produce fertile offspring. For these pairs, we obtained
331 evidence demonstrating no successful F2s from F1 hybrid pairings, an inability to
332 produce offspring other than by backcrossing to a parent species, or other biological
333 measurements (including histological assessments of the testes from the hybrid
334 males) that demonstrated complete infertility (Table S1, Supp Fig. 5).

335

336 ***Genetic Distance Calculation***

337 Both *CYTB* sequences and full mitogenomes (excluding the control region) of
338 multiple individuals of each species were collected from Genbank (Table S4) and
339 aligned using Clustal Omega version 1.2.4 [54]. In order to ensure that none of the

340 sequences were either mislabelled, or were nuclear copies of mitochondrial genes,
341 we constructed Neighbour-Joining trees using Geneious version 6.1.8 [55] and
342 removed all individuals that did not fall into monophyletic clades consisting of
343 individuals from each species. We first used *pModelTest* version 1.04 [56] to
344 determine the best model for the alignment of each set of sequences for both
345 species. We then calculated pairwise distances between each species pair using
346 RAXML version 8 [57] and FASTTree version 2.1 [58]. We also generated raw
347 distance values using the Hamming distance method which sums the number of
348 base pair differences (ignoring transition or transversion status) and divides that
349 number by the sequence length.

350

351 The distances were generated from the *CYTB* and nuclear gene alignments for each
352 set of species pairings in fasta file format using a Python 2.7 wrapper to automate
353 the terminal based programs RAXML, FastTree, and pModelTest. A custom Python
354 3.6 program was written to calculate Hamming distances of sequences making use
355 of the distance v0.1.3 [59] and Biopython v1.66 [60] modules. Gaps in the aligned
356 sequences were treated as missing data.

357

358 The mean distance and standard errors for each pairwise comparison were
359 calculated using the bootstrapping method on the assumption that the sets of
360 pairwise distances between related species would not be normally distributed. Each
361 pairwise comparison group containing Hamming distances was randomly resampled
362 into sets of equal sample size and processed using a helper function in the custom
363 software which made use of the bootstrapped v0.0.2, NumPy v1.10.1 [61] and SciPy
364 v0.16.0 [62] Python modules. The source code is available at
365 <https://github.com/BeebBenjamin/MrHamming>. The *CYTB* distances were validated
366 through comparison with data generated using the `compute between group mean
367 distance` method in MEGA X for GNU/Linux [63], using the following settings:

- 368 a) Variance Estimation Method: Bootstrap Method
- 369 b) No of Bootstrap Replications: 500
- 370 c) Substitutions Type: Nucleotide
- 371 d) Model/Method: p-distance
- 372 e) Substitutions to Include: d: Transitions + Transversions
- 373 f) Rates among Sites: Uniform Rates
- 374 g) Gaps/Missing/Data Treatment: Complete deletion

375 h) Select Codon Positions: 1st, 2nd, 3rd, Noncoding Site
376 Using a Student's t-test (two tailed), the differences between the results were found
377 to be statistically non-significant ($t=-0.11222$, $p= 0.912504$).

378

379 ***Mann-Whitney test of statistical difference between *CYTB* distance in hybrid***
380 ***categories***

381 The statistical significance of observed differences in *CYTB* divergence between
382 Categories 1 and 2 was tested using the Mann-Whitney U test ($p=6.216e-4$)
383 implemented in the R-software package [64].

384

385 **Figure Legends**

386

387 **Figure 1**

388 A depiction of the correlation between *CYTB* divergence between mammalian
389 species pairs and the relative fertility of their hybrid offspring. In column A, the green
390 circles represent species capable of producing fully fertile F1 offspring which can
391 reproduce independently of their parent species (Category 1). Brown circles
392 represent species pairs that follow Haldane's Rule and require backcrossing of a
393 female F1 with a parent species, or both sexes are sterile (Category 2). The green
394 and brown shaded regions represent the range of divergence values of the two
395 categories. Column B depicts the divergence between three wild felid species and
396 domestic cats, as well as the minimum number of generations of backcrosses with
397 domestic cats before full fertility of the hybrid is restored. The white circles in Column
398 C depict the divergence between three ancient hominins and AMH, as well as the
399 distances between AMH and chimpanzees and bonobos (in Category 2). The
400 asterisks represent those pairs that include modern samples of AMH. The lack of an
401 asterisk signifies that only sequences derived from archaeological AMH were used to
402 compute the divergence values. Details regarding the specific species pairs are
403 listed in Fig. S1 and Table S1.

404

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413

414 **References**

- 415 1. Gray AP. 1971 *Mammalian hybrids: A check-list with bibliography*. 2nd, revised, ed.
416 Farnham Royal: Commonwealth Agricultural Bureaux.
- 417 2. Cahill JA, Stirling I, Kistler L, Salamzade R, Ersmark E, Fulton TL, Stiller M, Green RE,
418 Shapiro B. 2015 Genomic evidence of geographically widespread effect of gene flow
419 from polar bears into brown bears. *Mol. Ecol.* **24**, 1205–1217.
- 420 3. Gopalakrishnan S *et al.* 2019 Interspecific Gene Flow Shaped the Evolution of the
421 Genus *Canis*. *Curr. Biol.* **29**, 4152.
- 422 4. Li G, Davis BW, Eizirik E, Murphy WJ. 2016 Phylogenomic evidence for ancient
423 hybridization in the genomes of living cats (Felidae). *Genome Res.* **26**, 1–11.
- 424 5. Figueiró HV *et al.* 2017 Genome-wide signatures of complex introgression and adaptive
425 evolution in the big cats. *Sci Adv* **3**, e1700299.
- 426 6. Li G, Figueiró HV, Eizirik E, Murphy WJ. 2019 Recombination-aware phylogenomics
427 reveals the structured genomic landscape of hybridizing cat species. *Molecular Biology*
428 *and Evolution*. (doi:10.1093/molbev/msz139)
- 429 7. Árnason Ú, Lammers F, Kumar V, Nilsson MA, Janke A. 2018 Whole-genome
430 sequencing of the blue whale and other rorquals finds signatures for introgressive gene
431 flow. *Sci Adv* **4**, eaap9873.
- 432 8. Skovrind M, Castruita JAS, Haile J, Treadaway EC, Gopalakrishnan S, Westbury MV,
433 Heide-Jørgensen MP, Szpak P, Lorenzen ED. 2019 Hybridization between two high
434 Arctic cetaceans confirmed by genomic analysis. *Sci. Rep.* **9**, 7729.
- 435 9. Martin SH *et al.* 2013 Genome-wide evidence for speciation with gene flow in *Heliconius*
436 butterflies. *Genome Res.* **23**, 1817–1828.
- 437 10. Martin SH, Davey JW, Salazar C, Jiggins CD. 2019 Recombination rate variation
438 shapes barriers to introgression across butterfly genomes. *PLoS Biol.* **17**, e2006288.
- 439 11. Edelman NB *et al.* 2019 Genomic architecture and introgression shape a butterfly
440 radiation. *Science* **366**, 594–599.
- 441 12. Fontaine MC *et al.* 2015 Mosquito genomics. Extensive introgression in a malaria vector
442 species complex revealed by phylogenomics. *Science* **347**, 1258524.
- 443 13. Lamichhaney S *et al.* 2015 Evolution of Darwin's finches and their beaks revealed by
444 genome sequencing. *Nature* **518**, 371–375.
- 445 14. Runemark A, Trier CN, Eroukhanoff F, Hermansen JS, Matschiner M, Ravinet M,
446 Elgvin TO, Sætre G-P. 2018 Variation and constraints in hybrid genome formation. *Nat*
447 *Ecol Evol* **2**, 549–556.

- 448 15. Haldane JBS. 1922 Sex ratio and unisexual sterility in hybrid animals. *J. Genet.* **12**,
449 101–109.
- 450 16. Bradley RD, Baker RJ. 2001 A Test of the Genetic Species Concept: Cytochrome-b
451 Sequences and Mammals. *J. Mammal.* **82**, 960–973.
- 452 17. Jančúchová-Lásková J, Landová E, Frynta D. 2015 Are genetically distinct lizard
453 species able to hybridize? A review. *Curr. Zool.* **61**, 155–180.
- 454 18. Edmands S. 2002 Does parental divergence predict reproductive compatibility? *Trends*
455 *Ecol. Evol.* **17**, 520–527.
- 456 19. Sánchez-Guillén RA, Córdoba-Aguilar A, Cordero-Rivera A, Wellenreuther M. 2014
457 Genetic divergence predicts reproductive isolation in damselflies. *J. Evol. Biol.* **27**, 76–
458 87.
- 459 20. Perelman P *et al.* 2011 A molecular phylogeny of living primates. *PLoS Genet.* **7**,
460 e1001342.
- 461 21. Castle WE. 1925 The Hare-Rabbit, A Study in Evolution by Hybridization. *Am. Nat.* **59**,
462 280–283.
- 463 22. Presgraves DC. 2002 Patterns of postzygotic isolation in Lepidoptera. *Evolution* **56**,
464 1168.
- 465 23. Price TD, Bouvier MM. 2002 The evolution of F1 postzygotic incompatibilities in birds.
466 *Evolution* **56**, 2083–2089.
- 467 24. Coyne JA, Orr HA. 1997 'Patterns of Speciation in *Drosophila*' Revisited. *Evolution* **51**,
468 295–303.
- 469 25. Teeter KC *et al.* 2008 Genome-wide patterns of gene flow across a house mouse hybrid
470 zone. *Genome Res.* **18**, 67–76.
- 471 26. Song Y, Endepols S, Klemann N, Richter D, Matuschka F-R, Shih C-H, Nachman MW,
472 Kohn MH. 2011 Adaptive introgression of anticoagulant rodent poison resistance by
473 hybridization between old world mice. *Curr. Biol.* **21**, 1296–1301.
- 474 27. Davis BW, Seabury CM, Brashear WA, Li G, Roelke-Parker M, Murphy WJ. 2015
475 Mechanisms Underlying Mammalian Hybrid Sterility in Two Feline Interspecies Models.
476 *Mol. Biol. Evol.* **32**, 2534–2546.
- 477 28. Detlefsen JA. 1914 *Genetic studies on a cavy species cross*. Washington, D. C.:
478 Carnegie Institution of Washington.
- 479 29. Thomsen PD, Schauser K, Bertelsen MF, Vejlsted M, Grøndahl C, Christensen K. 2011
480 Meiotic studies in infertile domestic pig-babirusa hybrids. *Cytogenet. Genome Res.* **132**,
481 124–128.
- 482 30. Arthur K. 1911 *Ancient Types of Man*. London: Harper & Brothers.
- 483 31. Currat M, Excoffier L. 2004 Modern Humans Did Not Admix with Neanderthals during
484 Their Range Expansion into Europe. *PLoS Biol.* **2**, e421.
- 485 32. Serre D, Langaney A, Chech M, Teschler-Nicola M, Paunovic M, Mennecier P, Hofreiter
486 M, Possnert G, Pääbo S. 2004 No evidence of Neandertal mtDNA contribution to early
487 modern humans. *PLoS Biol.* **2**, E57.

- 488 33. Krings M, Stone A, Schmitz RW, Krainitzki H, Stoneking M, Pääbo S. 1997 Neandertal
489 DNA sequences and the origin of modern humans. *Cell* **90**, 19–30.
- 490 34. Green RE *et al.* 2010 A draft sequence of the Neandertal genome. *Science* **328**, 710–
491 722.
- 492 35. Meyer M *et al.* 2012 A high-coverage genome sequence from an archaic Denisovan
493 individual. *Science* **338**, 222–226.
- 494 36. Fu Q *et al.* 2015 An early modern human from Romania with a recent Neanderthal
495 ancestor. *Nature* **524**, 216–219.
- 496 37. Slon V *et al.* 2018 The genome of the offspring of a Neanderthal mother and a
497 Denisovan father. *Nature* **561**, 113–116.
- 498 38. Petr M, Pääbo S, Kelso J, Vernot B. 2019 Limits of long-term selection against
499 Neandertal introgression. *Proc. Natl. Acad. Sci. U. S. A.* **116**, 1639–1644.
- 500 39. Holliday TW. 2006 Neanderthals and modern humans: an example of a mammalian
501 syngameon? In *Neanderthals Revisited: New Approaches and Perspectives* (eds J-J
502 Hublin, K Harvati, T Harrison), pp. 281–297. Dordrecht: Springer Netherlands.
- 503 40. Holliday TW, Gautney JR, Friedl L. 2014 Right for the Wrong Reasons. *Curr. Anthropol.*
504 **55**, 696–724.
- 505 41. Meyer M *et al.* 2013 A mitochondrial genome sequence of a hominin from Sima de los
506 Huesos. *Nature* **505**, 403–406.
- 507 42. Vernot B *et al.* 2016 Excavating Neandertal and Denisovan DNA from the genomes of
508 Melanesian individuals. *Science* **352**, 235–239.
- 509 43. Kuhlwilm M *et al.* 2016 Ancient gene flow from early modern humans into Eastern
510 Neanderthals. *Nature* **530**, 429–433.
- 511 44. Prüfer K *et al.* 2014 The complete genome sequence of a Neanderthal from the Altai
512 Mountains. *Nature* **505**, 43–49.
- 513 45. Etkind A. 2008 Beyond eugenics: the forgotten scandal of hybridizing humans and apes.
514 *Stud. Hist. Philos. Biol. Biomed. Sci.* **39**, 205–210.
- 515 46. Scally A *et al.* 2012 Insights into hominid evolution from the gorilla genome sequence.
516 *Nature* **483**, 169–175.
- 517 47. Hedges SB, Marin J, Suleski M, Paymer M, Kumar S. 2015 Tree of life reveals clock-like
518 speciation and diversification. *Mol. Biol. Evol.* **32**, 835–845.
- 519 48. Hill GE. 2016 Mitonuclear coevolution as the genesis of speciation and the
520 mitochondrial DNA barcode gap. *Ecol. Evol.* **6**, 5831–5842.
- 521 49. Ma H *et al.* 2016 Incompatibility between Nuclear and Mitochondrial Genomes
522 Contributes to an Interspecies Reproductive Barrier. *Cell Metab.* **24**, 283–294.
- 523 50. Orr HA, Turelli M. 2001 The evolution of postzygotic isolation: accumulating
524 Dobzhansky-Muller incompatibilities. *Evolution* **55**, 1085–1094.
- 525 51. Aiello LC. 2010 Five years of Homo floresiensis. *Am. J. Phys. Anthropol.* **142**, 167–179.

- 526 52. Berger LR, Hawks J, Dirks PH, Elliott M, Roberts EM. 2017 Homo naledi and
527 Pleistocene hominin evolution in subequatorial Africa. *eLife*. **6**. (doi:10.7554/elife.24234)
- 528 53. Ovchinnikov IV. 2013 Hominin evolution and gene flow in the Pleistocene Africa.
529 *Anthropologischer Anzeiger*. **70**, 221–227. (doi:10.1127/0003-5548/2013/0313)
- 530 54. Sievers F *et al.* 2011 Fast, scalable generation of high-quality protein multiple sequence
531 alignments using Clustal Omega. *Mol. Syst. Biol.* **7**, 539.
- 532 55. Kearse M *et al.* 2012 Geneious Basic: an integrated and extendable desktop software
533 platform for the organization and analysis of sequence data. *Bioinformatics* **28**, 1647–
534 1649.
- 535 56. Serra F. 2011 *pModelTest*. See <https://github.com/et toolkit/pmodeltest>.
- 536 57. Stamatakis A. 2014 RAxML version 8: a tool for phylogenetic analysis and post-analysis
537 of large phylogenies. *Bioinformatics* **30**, 1312–1313.
- 538 58. Price MN, Dehal PS, Arkin AP. 2010 FastTree 2--approximately maximum-likelihood
539 trees for large alignments. *PLoS One* **5**, e9490.
- 540 59. Meyer M. 2013 *distance*. See <https://github.com/doukremt/distance>.
- 541 60. Cock PJA *et al.* 2009 Biopython: freely available Python tools for computational
542 molecular biology and bioinformatics. *Bioinformatics* **25**, 1422–1423.
- 543 61. Oliphant TE. 2015 *Guide to NumPy*. 2nd edn. USA: CreateSpace Independent
544 Publishing Platform.
- 545 62. Virtanen P *et al.* 2019 SciPy 1.0--Fundamental Algorithms for Scientific Computing in
546 Python. *arXiv [cs.MS]*.
- 547 63. Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018 MEGA X: Molecular Evolutionary
548 Genetics Analysis across Computing Platforms. *Mol. Biol. Evol.* **35**, 1547–1549.
- 549 64. R Core Team. 2017 *R: A language and environment for statistical computing*. Vienna,
550 Austria: R Foundation for Statistical Computing. See <https://www.R-project.org/>.

Figure 1

